

Handbook of protocols for standardized measurement of terrestrial invertebrate functional traits

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41 **Summary**

- 42 1. Trait-based approaches are increasingly being used to test mechanisms underlying species
43 assemblages and biotic interactions across a wide range of organisms including terrestrial
44 arthropods and to investigate resulting ecosystem processes. However, such an approach relies on
45 the standardized measurement of functional traits that can be applied across taxa and regions.
46 Currently, unified methods of trait measurements are lacking for terrestrial arthropods.
- 47 2. Here, we present a comprehensive review and detailed protocol for a set of 28 traits known to be
48 sensitive to global stressors and/or affecting ecosystem processes and services. We give
49 recommendations how to measure these traits under standardized conditions across various
50 terrestrial invertebrate taxonomic groups.
- 51 3. We provide considerations and approaches that apply to almost all traits described, such as the
52 selection of species and individuals needed for the measurements, the importance of intraspecific
53 trait variability, how many populations or communities to sample and over which spatial scales.
- 54 4. The approaches outlined here provide a means to improve the reliability and predictive power of
55 functional traits to explain community assembly, species diversity patterns, and ecosystem
56 processes and services within and across taxa and trophic levels, allowing comparison of studies
57 and running meta-analyses across regions and ecosystems.
- 58 5. This handbook is only a first step towards standardizing trait methodology across the most studied
59 terrestrial invertebrate groups, and the protocols are aimed to balance general applicability and
60 requirements for special cases or particular taxa. Therefore, we envision this handbook as a
61 common platform to which everyone is kindly invited to provide methodological input for
62 additional special cases.

63

64 **Key-words:** species features, species characteristics, physiology, morphology, feeding, behaviour,
65 life-history, functional diversity.

66

67

68 **Introduction**

69 Over the last decade strong calls have been made to shift the research focus of community ecology
70 from species-based to trait-based ecology (among others Lavorel & Garnier 2002; McGill *et al.* 2006;
71 Diaz *et al.* 2007b; Suding *et al.* 2008; Webb *et al.* 2010; Chown 2012; Mouillot *et al.* 2013). This call
72 is driven by an increasing awareness that trait-based approaches can significantly enhance our
73 mechanistic understanding and predictive capabilities of the processes that play a major role in
74 community ecology. Moving from a taxonomical approach to a functional trait approach reduces
75 context dependency and therefore enables generalization across communities and ecosystems that is
76 needed to address macro-ecological questions (McGill *et al.* 2006; Suding *et al.* 2008; Kunstler *et al.*
77 2016). For example, traits can help explain the effects of environmental gradients and stressors on the
78 distribution of species and community (dis)assembly (e.g., Dias *et al.* 2013; Astor *et al.* 2014;
79 Woodcock *et al.* 2014), as well as the effect of community composition on ecosystem processes and
80 the provision of ecosystem services across ecological scales (Naeem & Wright 2003; Messier, McGill
81 & Lechowicz 2010; Luck *et al.* 2012; Brittain *et al.* 2013; Deraison *et al.* 2015). Trait-based
82 approaches have recently also been advocated as promising tools also in ecotoxicology and
83 environmental risk assessment of chemical substances (Rubach *et al.* 2011; Van den Brink *et al.*
84 2013).

85 Recent developments in trait-based ecology have been led by plant ecologists, as plant traits have
86 become effective predictors of community assemblages (de Bello *et al.* 2012; HilleRisLambers *et al.*
87 2012) and ecosystem processes (Lavorel 2013), and are now widely used. The prime utilization of
88 plant functional traits is to identify abiotic and biotic mechanisms that determine species composition,
89 ecosystem processes and service delivery (Lavorel & Garnier 2002; Diaz *et al.* 2007a; Luck *et al.*
90 2009; de Bello *et al.* 2010; Lavorel *et al.* 2013). Plant ecologists have been able to successfully scale
91 up from individual plant physiological traits to vegetation processes such as competition and
92 community assembly, as well as ecosystem processes such as decomposition, across a wide range of
93 plant communities (Diaz *et al.* 2004; Cornwell *et al.* 2008; Kunstler *et al.* 2016), and link trait
94 variability to global carbon cycle and climate models (Atkin *et al.* 2015). The early success of the
95 plant trait approach has fuelled the discussion on which traits need to be measured and how they

96 should be quantified in a standardized way. The development of large online trait databases in plant
97 ecology, such as LEDA (Kleyer *et al.* 2008) and TRY (Kattge *et al.* 2011), now provide quick access to
98 plant trait values, allowing comparisons even between ecosystems and biomes. Following this success
99 in plant ecology, interest has been growing among ecologists to adopt a similar trait-based approach in
100 other taxonomic groups (e.g., Vandewalle *et al.* 2010; Aubin *et al.* 2013; Pakeman & Stockan 2014;
101 Pey, Laporte & Hedde 2014; Fournier *et al.* 2015). Particularly for terrestrial invertebrates, attempts to
102 develop trait frameworks for specific taxa, e.g., Fountain-Jones, Baker & Jordan (2015) for beetles, or
103 to construct trait databases have been published, e.g., Falkner *et al.* (2001) for snails, Bouget, Brustel
104 and Zagatti (2008) for saproxylic beetles, Speight and Castella (2010) for hoverflies, Bertelsmeier *et*
105 *al.* (2013) for ants, Homburg *et al.* (2014) for carabid beetles, and Pey, Laporte and Hedde (2014) for
106 soil invertebrates.

107 Invertebrates have crucial roles as consumers of primary producers (e.g., herbivores, fungivores,
108 granivores etc.) and the products of animals and plants (i.e., leaf-litter, dead wood, dung and carrion),
109 they provide a staple food for higher trophic levels (e.g., for predators, parasites and parasitoids) and
110 are recognised as both facilitators of primary production (i.e. pollinators and detritivores) and as
111 ecosystem engineers (e.g., soil bioturbators) (see Gagic *et al.* 2015 for an overview). Hence,
112 knowledge of invertebrate traits are key to understanding multi-trophic processes and ecosystem
113 functioning (e.g., Lavorel *et al.* 2013; Schmitz *et al.* 2015). Current invertebrate trait databases are
114 often built, around a set of basic traits from a mixture of studies and observations, that are obtained
115 without uniform methodology and with little consistence in which traits were chosen for
116 measurements. In addition, functional trait values, such as species temperature tolerance and drought
117 resistance, are often missing or inferred from the abiotic conditions at the (micro)habitats where they
118 have been observed. However, (micro)habitat selection of species and realized niche in general might
119 result from interactions between species rather than physiological and phenological characteristics of
120 single individuals and populations (Colwell & Fuentes 1975; Ellers, Dias & Berg 2010; Araujo *et al.*
121 2013; Colas *et al.* 2014; He & Bertness 2014), but see Warren, Giladi & Bradford (2010). The use of
122 such inferred traits as predictors of community and ecosystem processes has been strongly
123 discouraged (Violle *et al.* 2007). The arguments above raise the urgent need for reliable and unified

124 methods to measure functional traits that are directly linked to species performance. A coherent,
125 unified and standardized trait approach for various types of terrestrial invertebrates requires consensus
126 on 1) what the appropriated functional traits are and, particularly, on 2) how they should be measured.
127 A key element in this process has been the provision of a handbook of standardized plant functional
128 traits that detail the methods and definitions of key traits worldwide (Cornelissen *et al.* 2003), and its
129 recent update with additional traits and measuring techniques (Pérez-Harguindeguy *et al.* 2013).
130 The present work aims to provide a similar incentive to the trait-based approach for terrestrial
131 invertebrates by describing a set of standardized trait measurements in easy to use protocols to
132 improve the reliability and predictive power of functional traits to explain community and ecosystem
133 processes within and across taxa and trophic levels, allowing comparison of studies and running meta-
134 analyses across regions and ecosystems.

135

136 *Overall approach to the handbook*

137 This handbook aims to provide a set of protocols for trait measurements that can be used across a wide
138 range of terrestrial invertebrate species, including the major taxonomic groups of Insecta, Aranea,
139 Crustaceae, Myriapoda, Gastropoda and Oligochaeta. We recognise that the wide variety of life forms
140 encompassed by the present handbook makes it a challenging undertaking. In general, invertebrate
141 traits are harder to determine and calibrate compared to plant traits, since animals can respond to
142 environmental changes by movement and behaviour. Therefore, the trait protocols contain
143 recommendations for adjustments to accommodate the biology of particular taxonomic groups, while
144 maintaining comparability and standardization across taxa. The handbook does not include specific
145 methods for measuring traits of nematodes, parasites and aquatic invertebrates, although some of the
146 protocols may be used for these groups as well.

147 The handbook is meant as a first step to advance the trait-based approach to trophic groups other than
148 plants, and to stimulate discussion about additional traits that should be included in the handbook. We
149 foresee that this set of traits might be expanded in the future as the use of the functional approach
150 becomes increasingly used among animal ecologists. Moreover, the trait protocols are designed for
151 easy and standardized measurement of traits to facilitate widespread use by any research group, and to

152 allow high-throughput phenotyping to enable measurements on large numbers of species. For this
153 reason, some of the most advanced technological methods that are currently used by specialized
154 research groups for specific taxonomic groups are not part of the standardized methods, but included
155 as special cases in the protocols. We would like to emphasize that the handbook's main purpose is to
156 maximize comparability of measurements across a wide range of taxa.

157 Below, we first give an overview of the criteria and concepts used for selecting the set of traits,
158 subsequently we describe the standard format of the protocols, followed by some general
159 recommendations. The protocols themselves are provided as an electronic appendix (Appendix S1).

160

161 *Trait selection*

162 We reviewed the literature on ecology of terrestrial invertebrates, and selected the 28 traits (see Table
163 1) for which we found clear evidences that they directly link organism performance with
164 environmental conditions or ecosystem processes. These traits have been then further discussed among
165 a group of specialist scientists working on the ecology, ecophysiology, and evolutionary aspects of
166 predominantly terrestrial invertebrate fauna at different trophic levels with the aim to standardize the
167 methods for their unambiguous use in any terrestrial biome and for the majority of its constituents.

168 Overall, the selected set of traits largely covers the primary functions related to species performance
169 and interactions between trophic levels at various spatial scales from plots to landscape and even
170 biomes. Traits can be separated into response traits which determine the response of the species to an
171 environmental change, and effect traits which contribute to ecosystem function (Lavorel & Garnier
172 2002; Naeem & Wright 2003). In Table 1 and trait protocols we defined response and effect properties
173 of the traits selected. We focus on several effect and response traits, which based on the literature, are
174 among the most widely used or are in urgent need of standardized measurement protocols that can be
175 applied across taxa. Most of the selected traits are quantitative and directly measurable on an
176 individual under standardized conditions; others are categorical (e.g., activity time and feeding guild)
177 or ordinal (e.g., ontogeny and respiration system).

178 Broadly, the selected traits can be grouped into five categories, i.e., morphology, feeding, life history,
179 physiology, and behaviour. *Morphological traits* such as eye morphology, body pigmentation or body

180 size are important features of an organism's interaction with the abiotic and biotic environment. For
181 example, body size is a predictor of multiple ecological processes (de Bello *et al.* 2010), and strongly
182 correlated with an individual's metabolic rate (Peters 1983; Brown *et al.* 2004). Body size also scales
183 with many other life history traits (Ellers & Jervis 2003) and determines the structure and function of
184 ecological networks (Woodward *et al.* 2005). *Feeding traits* are related to the trophic position of a
185 species and describe aspects of the morphology and behaviour associated with their diet. Feeding
186 related traits can therefore be important to better understand niche partitioning, trophic interactions as
187 well as shape the structure of ecological networks (Stang *et al.* 2009; Ibanez 2012; Ibanez *et al.* 2013).
188 *Life history traits* describe the age schedule of reproduction of an organism, including key
189 reproductive aspects such as age at maturity, clutch size, and life span (Stearns 1992). These traits
190 have strong links to fitness and are expected to be among the most sensitive to environmental stress,
191 making them useful to assess the vulnerability of species to global change. For instance, egg size
192 varies enormously between species (Fox & Czesak 2000) and affects hatching success (Fischer *et al.*
193 2006) and resistance to desiccation (Fischer *et al.* 2006) and heat (Lieferting *et al.* 2010). Moreover,
194 trade-offs exist between reproductive traits and dispersal (Guerra 2011), leading to a reduced
195 reproductive investment in some insects with strong range expansion under the influence of global
196 warming (Hughes, Hill & Dytham 2003).

197 *Physiological traits* refer to features that allow species to tolerate variations in abiotic conditions
198 (resistance adaptations), as well as biochemical modifications that adjust the rate of
199 metabolic function (capacity adaptations) in response to environmental changes (Somero 1992).
200 Physiological tolerance traits, such as heat tolerance and desiccation resistance have been successfully
201 applied in predicting species distribution patterns along abiotic gradients (Dias *et al.* 2013), while
202 growth rate can determine an individuals' susceptibility to predation (Denno *et al.* 2002; Coley,
203 Bateman & Kursar 2006) and temperature fluctuations (Fordyce & Shapiro 2003). Further,
204 physiological tolerances can be affected by changes in diet (Verdu *et al.* 2010).

205 Finely, *Behavioural traits* enable flexible, rapid responses to environmental change without any
206 associated changes to physiological or morphological phenotypes. Traits such as activity time,
207 aggregation, and locomotion allow organisms to seek out preferred microhabitats to avoid (a)biotic

208 stress. Behavioural strategies can also increase tolerance to abiotic stresses, for instance through
209 adopting flight strategies that maximize heat dissipation (Verdu, Alba-Tercedor & Jimenez-Manrique
210 2012) or by choosing microclimates to achieve nutritional homeostasis (Clissold, Coggan & Simpson
211 2013). Yet in soil fauna species, stratification in soil interacts with other traits, such as physiological
212 traits, thus modifying the individual response to changes in environmental conditions (Cloudsley-
213 Thompson 1962) and vulnerability to extreme temperature events (van Dooremalen *et al.* 2012).

214

215 **The handbook protocols**

216 The trait protocols are described using a standard format aimed to facilitate comparisons among traits.
217 Each protocol includes four main sections. The section *Definition and relevance* provides a formal
218 definition and a short, non-exhaustive justification why that particular trait is of ecological
219 significance based on its role in responding to stressors and/or effecting trophic interactions or
220 ecosystem processes. This section also describes the main approaches to measure a particular trait. The
221 section *What and how to measure* describes the standardized method, and provides the units of
222 expression and, if applicable, mathematical formulas for trait value calculations. The section
223 *Additional notes* contains, if available, alternative techniques, often more expensive and challenging,
224 and mainly used by more specialized research groups to answer deeper questions. This section may
225 also list modifications of the methods for specific taxonomic groups and draws attention to potential
226 caveats. Finally, the *References* list a number of key papers which are cited in the protocol.

227

228 *Standardization of measurements and acclimation of animals*

229 Invertebrates respond to a multitude of external environmental factors, leading to differences in trait
230 values due to trait plasticity, learning and shifts in physiological status. As a consequence, trait values
231 may depend on the immediate conditions an organism is subjected to at the place or time of collection.
232 To achieve standardized trait measurements it is necessary to provide the same conditions for all
233 individuals measured, which for many traits requires an acclimation period in order to remove the
234 effect of local conditions. Therefore, the handbook starts off with a standardization paragraph that

235 describes recommendations for pre-treating and acclimating animals to obtain comparable values
236 within and among species for all taxonomic groups.

237 For traits with survival time as the unit of expression, such as inundation resistance, all individuals
238 should have the same nutritional status at the start of the measurements and should either be fully fed
239 or subjected to a short starvation period to empty their gut prior to trait measurements. When
240 measuring feeding traits (e.g., food preference, ingestion rate) it is necessary that all individuals are
241 acquainted with the food items used during the feeding assays. For traits that are strongly temperature-
242 dependent such as metabolic rate, food ingestion rate and locomotion speed, thermal acclimation is
243 absolutely necessary, although the acclimation time depends on the organisms and specific life cycles,
244 as well as on the trait and ontogenetic stage of interest. As trait plasticity can occur during an
245 organisms' ontogeny, it might be necessary to raise animals under controlled conditions (controlled
246 environmental rooms) and measure traits in individuals born into these rooms.

247 Obviously, in cases where the research interest is focused on the actual survival time when animals are
248 exposed to drought in their habitat, the actual diet composition in the field, or the dispersal distance
249 under natural conditions, then standardized measurements will not need to be imposed, except perhaps
250 for serving as a baseline to measure the extent by which field conditions depart from basal adaptations.

251

252 *Selection of specimens and number of individuals per species*

253 A key consideration is selecting the appropriate specimens for trait measurements. Aiming to compare
254 standardized trait measurements across studies and taxa of any developmental stage and sex, we
255 recommend selecting healthy, well-shaped, and full-developed individuals of the ontogenetic stage of
256 interest, without any signs of damage and diseases. The use of interception trapping devices, such as
257 pitfall traps, windowpane traps and Malaise traps to collect species for trait measurements should be
258 regarded with caution as the performance of a trap depends on its construction, location, time of day,
259 season or year, and weather (Gibb & Oseto 2006), and, most importantly, they might be selective for
260 specimen with certain traits. We recommend that the sampling methods should be reported in detail
261 and that additional information on trapping efficiency should be provided together with the trait
262 measurements.

263 When laboratory strains are used for measurements, extreme care should be taken as laboratory
264 adaptation may cause spurious changes in life history and physiological traits of species (Sgro &
265 Partridge 2001; Griffiths, Schiffer & Hoffmann 2005). The type of culturing method, the size of the
266 stock population and the length of the period of laboratory culture are all factors that determine the
267 magnitude of selection response in laboratory population, and therefore these factors need to be
268 reported meticulously with the trait measurements.

269 Sample size is a general issue in trait-based approaches and has already been covered in other
270 publications, although mainly on plants (e.g., Pakeman & Quasted 2007; Bolnick *et al.* 2011; de Bello
271 *et al.* 2011; Fu *et al.* 2013; Pérez-Harguindeguy *et al.* 2013). If one would like to capture the full
272 spatiotemporal variability around a species trait mean, a proportional number of individuals should be
273 measured from different populations, seasons, communities, and ecosystems (Pakeman & Quasted
274 2007; de Bello *et al.* 2011; Violle *et al.* 2012). This number will further increase if other sources of
275 intraspecific variation will be included, e.g. polymorphism, sexual dimorphism and ontogenetic stages
276 (Yang & Rudolf 2010; Violle *et al.* 2012), which are all particularly important among invertebrates. In
277 general, the minimal number of individuals to be measured for a given species will depend on the
278 variation of the trait values. The higher the variation, the higher the numbers of individuals to be
279 measured for reliable estimates of the species mean trait value.

280

281 **Future perspectives**

282 This handbook is a first step towards standardizing trait methodology across the most studied
283 terrestrial invertebrate groups. We are aware that its protocols are far from covering all special cases
284 and may miss information for particular taxa. Below we highlight three fields that we hope to develop
285 further with the aid of this handbook and provide some future perspective on the field of trait research.

286

287 *Incorporating intraspecific trait variability*

288 There is increasing evidence that intraspecific trait variability determine community assembly and the
289 distribution of individuals across different spatio-temporal scales, as well having implications for
290 ecosystem processes (Bolnick *et al.* 2011; de Bello *et al.* 2011; Violle *et al.* 2012; Siefert *et al.* 2015).

291 Within-species variability may originate from spatial variability in trait values within a species range,
292 or may be due to genetic or environmental variation within a population at a single site. Information
293 on both types of variability is extremely valuable, e.g. for understanding the mechanisms underlying
294 community assembly or as input for models on functional consequences of global drivers (Yang &
295 Rudolf 2010). Until now the lack of standardized measurements for invertebrate traits, as well as the
296 tiny sample size for many traits, has prohibited a clear indication of the trait variability beyond the
297 single species level. We believe that the use of the standardized protocols can overcome this gap.

298

299 *Definition and validation of effect traits*

300 Quantifying community variation in response traits, the redundancy among species sharing similar
301 effect traits, as well as the overlap between response and effect traits is important for enhancing
302 predictability of ecosystem functioning under environmental change (Folke, Holling & Perrings 1996;
303 Elmquist *et al.* 2003; Mori, Furukawa & Sasaki 2013). While our knowledge on response traits of
304 terrestrial invertebrates is relatively good, information on the extent to which response traits and effect
305 traits can be linked within taxa, either via trait correlations or trait trade-offs, is still largely lacking.
306 Even less is known about response-to-effect models across trophic levels (Schmitz 2008; Lavorel *et al.*
307 2013; Moretti *et al.* 2013; Pakeman & Stockan 2014; Deraison *et al.* 2015), although the degree of
308 overlap between the two types of traits will determine our ability to predict changes in key ecosystem
309 processes under variable environmental conditions. The current definition of response and effect traits
310 in invertebrates is based on literature and expert knowledge, but validation based on controlled
311 experiments is urgently needed.

312

313 *Construction of an invertebrate trait database*

314 The benefits of standardized trait measurements to the research community can be amplified if this
315 information is compiled in a communal database. Following the successful example of the worldwide
316 TRY initiative, we propose that increased access to trait information collected with standardized
317 protocols will promote the interest to use this data. However, construction and maintenance of such a
318 large database is a major undertaking that requires a dedicated staff and long-term funding. We hope

319 that an enthusiastic and regular use of this first handbook of protocols for standardized measurement
 320 of terrestrial invertebrate functional traits will encourage researchers and funding agencies alike to
 321 taking this crucial long term option.

322

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542 **Table 1** – List of the terrestrial invertebrate traits selected for the handbook and considered to be key in responding to the environment (RT,
 543 response traits) and/or effecting ecosystem processes and services (ET, effect traits) at various scales from local plots, to landscapes and biomes.
 544 Symbols: “-” no relation with response to or effect on environment, “+” affinity with response to or effect on environment, “++” strong affinity with
 545 response to or effect on environment; the evaluation is based on qualitative expert knowledge.

| <i>Trait type Trait</i> | Definition | RT | ET | Comment |
|--------------------------|---|-----------|-----------|--|
| <i>Morphology</i> | | | | |
| Body size | Size of the body. It includes body length, body width, body mass, and body volume | ++ | ++ | Environmental conditions affect body size which will influence amount and composition of resources used |
| Eye morphology | Form of the eye. It includes: eye number, eye size, eye sight | + | + | Eye morphology can be filtered by environmental conditions which will reflect prey and/or predator recognition |
| Respiration system | Structures developed to perform gas exchange | ++ | - | Type of respiration mode directly affect drought tolerance and desiccation resistance |
| Hairiness | Degree of hair coverage. It includes: hair length and hair density | + | + | Abiotic condition and biotic interactions (pollination) affect hairiness providing fitness and performance |
| Colour | Body coloration. It includes: colour, intensity, contrast | + | + | Abiotic condition and biotic interactions (e.g. predation) affect pigmentation providing fitness and performance |
| <i>Feeding</i> | | | | |
| Feeding guild | Food type, upon which species feed. It informs about “who eats what or whom” | ++ | ++ | Feeding guild is a good surrogate for trophic level and position in the food web. It determines the quality of resources, which influences a species growth, reproduction and survival |
| Ingestion rate | Quantity of food consumed in a given period | ++ | ++ | The rate of food ingested by an organism reflects its nutritional and energetic requirements and is related to species responses to food quality |

| | | | | |
|------------------------|---|----|----|---|
| Biting force | Biomechanical force exerted on food items by the tip of the mouth-parts, claws or fore legs | + | ++ | Biting force mainly determines the effect on trophic network interactions and thus on ecosystem function |
| <i>Life history</i> | | | | |
| Ontogeny | Developmental history. It includes type and number of developmental stages | ++ | + | Response to environmental stressors and effects on the ecosystem can change significantly across an organism's life history. Changes in environmental conditions can affect ontogeny and ecosystem processes |
| Clutch size | Number of eggs or juveniles produced in one reproductive event | ++ | ++ | Clutch size respond significantly to environmental conditions which affect number of offspring and their impact on the ecosystems |
| Egg size | Size dimension or mass of an egg | ++ | + | Resistance to environmental and particularly climatic conditions increase with egg size, which indirectly determines impact on the ecosystem via changes in population sizes |
| Life span | Amount of time an adult individual lives, from emergence from last instar until death | ++ | ++ | Stressors can heavily affect life span which is reflected in different ecosystem functions |
| Age at maturity | Age at first reproductive event | ++ | + | Time of first reproductive event can be changed under environmental stress, with consequences for population size and ecosystem processes |
| Parity | The number of times a females lays eggs or gives birth | ++ | + | The spreading of reproductive events over a life time has fitness consequences that are related to the trade-off between current and future reproduction |
| Reproduction mode | Mode by which new offspring are produced (sexual or asexual) | + | + | Mode of reproduction can be changed under environmental stress, with consequences for population sizes and ecosystem processes |
| <i>Physiology</i> | | | | |
| Resting metabolic rate | Amount of energy expended by an organism at rest | ++ | + | Metabolic rate is related to several organism features such as behaviour, longevity and reproduction output and its reaction norm with temperature can indicate how organisms differ in their response to environmental changes |

| | | | | |
|-------------------------|--|----|---|---|
| Relative growth rate | Increasing in mass of an organism per unit of time | ++ | + | Relative growth rate is related to other several life history traits, such as body size and age at maturity. Therefore, growth rate can influence different fitness components such as fecundity and survival |
| Desiccation resistance | Ability to withstand dry conditions | ++ | - | Physiological capacity to resist dry conditions is related to species distribution along water availability gradients and to species response to changes in water availability |
| Inundation resistance | Ability of terrestrial organisms to survive under water | ++ | - | Flooding and increased frequency and intensity of extreme precipitation can impose strong restrictions on survival |
| Salinity resistance | Ability to withstand conditions of high salinity | ++ | - | Ability to withstand conditions of high salinity determines species survival under high salt stress and will influence growth and reproduction via trade-offs |
| Temperature tolerance | Ability to survive at any temperature. It includes: hot and cold | ++ | - | Tolerance of Hot and cold temperatures determines species survival under stress and will influence growth and reproduction via trade-offs |
| pH resistance | Ability to withstand acidic or alkaline conditions | ++ | - | Ability to withstand acidic or alkaline conditions determines species survival under acidity stress and will influence growth and reproduction via trade-offs |
| <i>Behaviour</i> | | | | |
| Activity time | Activity period of a species within 24h | ++ | + | Environmental conditions, e.g. climatic conditions, determine the activity time. This can affect ecosystem function through asynchrony, e.g. spatiotemporal mismatch in biotic interactions |
| Aggregation | Clustering of individuals | + | + | Clustering of individual reduces microclimatic stress, especially overcoming cold and drought and can locally result in enhanced ecosystem process rates via high population sizes |
| Dispersal mode | The form of self-directed movements an animal uses to move from one place to another | ++ | - | Dispersal mode influences access to new habitat, resources and suitable environments, mates, and shelters, and opportunities to escape adverse environmental conditions |

| | | | | |
|----------------------|--|----|----|--|
| Locomotion speed | The pace of self-propelled movement of an organism | ++ | + | Habitat conditions and biotic interactions influence locomotion speed which reflect behaviours critical for survival, including efficient use of resources, foraging, predator avoidance, fitness and survival |
| Sociality | Degree of interactive behaviour with other members of its species to the point of having a recognizable and distinct society | + | ++ | Disturbance and land use changes are expected to affect sociality. High levels of sociality are expected to have a bigger impact on ecosystem function |
| Annual activity time | Period in an organism's life cycle when growth, development, and physical activity are temporarily stopped | ++ | - | Offers the possibility to overcome unfavourable environmental conditions in a resting stage |

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