Evaluation of a full-scale wastewater treatment plant upgraded with

ozonation and biological post-treatments: Abatement of micropollutants,

formation of transformation products and oxidation by-products

- Marc Bourgin¹, Birgit Beck¹, Marc Boehler¹, Ewa Borowska¹, Julian Fleiner¹, Elisabeth Salhi¹, Rebekka
- 6 Teichler¹, Urs von Gunten^{1,2,3}, Hansruedi Siegrist¹, Christa S. McArdell^{1,*}

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- ¹Eawag, Swiss Federal Institute of Aquatic Science and Technology, CH-8600 Dübendorf, Switzerland
- ²School of Architecture, Civil and Environmental Engineering (ENAC), Ecole Polytechnique Fédérale
- de Lausanne (EPFL), CH-1015 Lausanne, Switzerland
- ³Institute of Biogeochemistry and Pollutant Dynamics (IBP), ETH Zurich, CH-8092 Zurich, Switzerland

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* Corresponding author: Christa S. McArdell, christa.mcardell@eawag.ch

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ABSTRACT

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To protect the ecosystem and drinking water resources in Switzerland and in the countries of the downstream catchments, a new Swiss water protection act entered into force in 2016 aiming to reduce the discharge of micropollutants from wastewater treatment plants (WWTPs). As a consequence, selected WWTPs must be upgraded by an advanced treatment for micropollutant abatement with suitable and economic options such as (powdered) activated carbon treatment or ozonation. WWTP Neugut (105'000 people equivalent) was the first WWTP in Switzerland to implement a long-term fullscale ozonation. Differing specific ozone doses in the range of 0.35-0.97 g O₃/g DOC were applied to determine the adequate ozone dose to fulfill the requirements of the Swiss water protection act. Based on this assessment, a specific ozone dose of 0.55 g O₃/g DOC is recommended at this plant to ensure an average abatement of the twelve selected indicator substances by $\geq 80\%$ over the whole treatment. A monitoring of 550 substances confirmed that this dose was very efficient to abate a broad range of micropollutants by >79% on average. After ozonation, an additional biological post-treatment is required to eliminate possible negative ecotoxicological effects generated during ozonation caused by biodegradable ozonation transformation products (OTPs) and oxidation by-products (OBPs). Three biological treatments (sand filtration, moving bed, fixed bed) and granular activated carbon (GAC, fresh and pre-loaded) filtration were evaluated as post-treatments after ozonation. In parallel, a fresh GAC filter directly connected to the effluent of the secondary clarifier was assessed. Among the three purely biological post-treatments, the sand filtration performed best in terms of removal of dissolved organic carbon (DOC), assimilable organic carbon (AOC) and total suspended solids (TSS). The fresh activated carbon filtration ensured a significant additional micropollutants abatement after ozonation due to sorption. The relative abatement of the indicator substances ranged between 20-89% after 27'000 bed volumes (BV) and was still substantial at 50'000 BV. In an identical GAC filter running in parallel and being fed with the effluent of the secondary clarifier, the elimination was less efficient. Seven primary OTPs (chlorothiazide and six N-oxides) formed during ozonation could be quantified thanks to available reference standards. Their concentration decreased with increasing specific ozone doses with the concomitant formation of other OTPs. The seven OTPs were found to be stable compounds and were not abated in the biological post-treatments. They were sorbed in the fresh GAC filter, but less

- efficiently than the corresponding parent compounds. Two OBPs, bromate (BrO₃⁻) and N-
- nitrosodimethylamine (NDMA), were formed during ozonation but did not exceeded 5 μ g/L for bromate
- and 30 ng/L for NDMA at the recommended specific ozone dose of 0.55 g O₃/g DOC. NDMA was well
- 49 abated in all post-treatments (minimum 41% during fixed bed filtration, maximum 83% during fresh
- 50 GAC filtration), while bromate was very stable as expected.

INTRODUCTION

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More than 100'000 compounds are registered today in Europe (European Chemical Agency 2017), 52 53 including pharmaceuticals, personal care products, food additives, and industrial chemicals. A large 54 number of them are frequently observed as persistent compounds in surface waters in levels of ng/Lμg/L (Petrie et al. 2014, Richardson and Kimura 2016). Therefore, sensitive species in the aquatic 55 ecosystem may be adversely affected by, e.g., a damage of the nervous systems, feminization and 56 57 disruption of reproduction, or inhibition of photosynthesis (Stalter et al. 2013). Furthermore, micropollutants in surface waters and in groundwaters can contaminate the drinking water resources 58 59 (Lopez et al. 2015). Many micropollutants observed in water bodies originate from WWTP discharges, where polar 60 61 micropollutants are generally poorly abated by conventional treatments (Loos et al. 2013), as WWTPs 62 are not designed to eliminate micropollutants. The task of conventional WWTPs is to remove nutrients 63 (inorganic nitrogen and total phosphorus), suspended solids and to reduce the biological and chemical 64 oxygen demand (BOD₅ and COD, respectively) (Ternes et al. 2017). 65 In Switzerland, a new water protection act entered into force as of January 2016 with the objective to improve surface water quality by reducing the load of micropollutants from WWTPs. Within the next 66 twenty years, about 100 of the 700 existing Swiss WWTPs will have to implement an advanced 67 68 treatment step for micropollutant abatement, following four criteria: (i) large WWTPs to significantly 69 reduce the load; (ii) WWTPs in the catchment of lakes; (iii) WWTPs at rivers with a high fraction of 70 wastewater to protect ecosystems; and (iv) WWTPs at surface waters which serve as drinking water 71 resources. 72 The aim is to remove organic micropollutants by 80% on average over the whole WWTP from influent 73 to effluent (Eggen et al. 2014). To ensure the efficiency of the upgraded wastewater treatment, the Swiss 74 Federal Office of Environment (FOEN) has established a shortlist of 12 indicator substances (SI, Table 75 S1), which must be abated on average by 80% (FOEN 2017). The 12 compounds were specifically 76 chosen as non-easily biodegradable substances and are therefore normally not well removed during 77 conventional wastewater treatments.

Adsorption by powdered activated carbon (PAC) and ozonation are currently the recommended technologies that lead cost-efficiently to a significant abatement of the micropollutant load and toxicity (Hollender et al. 2009, Logar et al. 2014, Margot et al. 2013, Schindler Wildhaber et al. 2015). Besides, a treatment with granular activated carbon filtration has been recently considered as a potential alternative and is already applied on a full-scale WWTP in Germany (Benstoem et al. 2017). Ozone reacts quickly with electron-rich moieties, e.g., olefins, tertiary amines, thioethers and activated aromatics (von Sonntag and von Gunten 2012), while hydroxyl radicals (•OH), which are formed from self-decomposition of ozone (Hoigné and Bader 1975, Staehelin et al. 1984, Staehelin and Hoigné 1982), react additionally with alkanes, amides and non-activated aromatic compounds (Buxton et al. 1988). If the mechanism and the kinetics of the reactions of a substance or moiety with ozone are known, the possible OTPs and the rate of abatement of a substance can in principle be predicted (Hübner et al. 2015, Lee et al. 2017, Lee and von Gunten 2016, von Sonntag and von Gunten 2012). Even though there is a lot of mechanistic information available, there are still significant knowledge gaps on certain functional groups. Furthermore, ozonation may yield oxidation/disinfection by-products (OBPs) by reaction of ozone and hydroxyl radicals with the different components of the water matrix. Some OBPs such as BrO₃⁻ and nitrosamines, e.g., N-nitrosodimethylamine (NDMA), are known to be (potential) human carcinogens (Kurokawa et al. 1990, Tricker and Preussmann 1991). Therefore, the formation of these OBPs needs to be minimized during ozonation. In Switzerland, the formation of BrO₃ and NDMA during ozonation needs to be assessed according to a recently introduced test procedure to evaluate the treatability of wastewater with ozone (Schindler Wildhaber et al. 2015). Bromate is formed from the successive oxidative reactions of bromide (Haag and Hoigné 1983, von Gunten and Hoigné 1994) present in water bodies and wastewaters in µg/L-mg/L levels (Soltermann et al. 2016). The main bromide sources in Switzerland are from wastewaters discharged from municipal waste incinerators, industrial landfills and chemical industries (Soltermann et al. 2016). A bromate drinking water standard/guideline value for BrO₃ was set to 10 μg/L (Commission Directive 2003, U.S. EPA 2006, WHO 2011) and acute and chronic environmental quality standards (EQS) of 50 µg/L were proposed by the Ecotox Centre Eawag-EPFL (Ecotox Center 2017). NDMA is formed during ozonation of precursors such as hydrazines and

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106 sulfamides (yield >50%) (Kosaka et al. 2009, Krasner et al. 2013, Lim et al. 2016, Schmidt and Brauch 107 2008, von Gunten et al. 2010). The NDMA guideline value in drinking water was set to 100 ng/L by 108 WHO (2008) and even lower values of 10 ng/L are proposed in countries such as the USA and Germany 109 (California Department of Public Health 2010, Massachusetts Department of Environmental Protection 110 2004, Planas et al. 2008). 111 Some toxicity endpoints may increase after ozonation due to the formation of aldehydes and organic 112 acids. It has been recently concluded that OBPs are toxicologically probably more relevant than OTPs (Lee and von Gunten 2016). However, toxicity was found to decrease again after biological post-113 114 treatments (Stalter et al. 2010a, Stalter et al. 2010b), because of the biodegradability of toxic OBPs and 115 OTPs. Therefore, a biological post-treatment is required with the goal to eliminate biodegradable, toxic 116 OTPs and OBPs leading to negative ecotoxicological effects. 117 The present study aims at evaluating a full-scale wastewater treatment with installed ozonation and post-118 treatment. To this end, three purely biological post-treatments (a sand filter, a moving bed reactor, and a 119 fixed bed reactor) were investigated as well as GAC filtration with additional sorption capacity (a fresh 120 and a pre-loaded GAC filter). Additionally, a fresh GAC filter was run in parallel directly with the 121 secondary effluent for comparison. The effects of the differing treatment steps were assessed on the 122 chemical (current paper) and biological (Kienle et al. in preparation) wastewater quality. Additionally, 123 non-target peaks across the wastewater treatment train were characterized (Schollee et al. 2017). Here, 124 the abatement of 43 micropollutants frequently detected in wastewater, including the indicator 125 substances, was first investigated at various ozone doses to determine the required specific ozone dose to abate the indicator substances by 80% on average, as requested by the new Swiss water protection act. 126 127 Thereafter, the abatement of micropollutants from an exhaustive list of 550 substances was studied at the 128 optimum specific ozone dose for an overall evaluation of the advanced treatment. With a target and 129 suspect screening approach, the formation of known OTPs and the OBPs BrO₃ and NDMA during 130 ozonation was investigated. The fate of micropollutants, OTPs and OBPs over the differing biological 131 post-treatments and in the GAC filters was studied and compared.

MATERIAL AND METHODS

Wastewater treatment plant

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Neugut WWTP (Dübendorf, Switzerland) (Fig. S1, Text S1) treats wastewater of about 50'000 persons and 105'000 people equivalent (maximum capacity of 150'000 people equivalent) with an approximate industrial contribution of 50% in COD load (mainly from food industries). The flow in the treatment plant ranges between 13'000 and 57'000 m³/day with an average of 19'000 m³/day at dry weather conditions. The wastewater treatment originally consisted of a primary clarifier, a conventional activated sludge treatment with a secondary clarifier and a sand filtration. The average sludge retention time in the conventional biological treatment is about 13 days and the hydraulic retention time (HRT) is 18 h (without settling tank). Dissolved organic carbon (DOC) after the conventional biological treatment was 3.5 – 6 mg/L and nitrite below 0.04 mg N/L; other quality parameters are given in Table S2). Although the industrial contribution is relatively high in this WWTP, the quality of the secondary effluent is similar to typical Swiss domestic wastewater (Schindler Wildhaber et al. 2015, Soltermann et al. 2016). In 2014, an ozonation reactor (V = 530 m³, divided in 6 reactors in series) was installed between the secondary clarifiers and the four sand filters (4x31.5 m³ operated in parallel, quartz sand, 0.7-2.0 mm granular size with an average empty bed contact time (EBCT) of 10 min). Ozone is produced with two ozone generators (OZAT-CFV5, Ozonia, Dübendorf, Switzerland) from oxygen fed by a liquid oxygen tank. The ozone reactor has an average HRT of 43 minutes at dry weather conditions and of 13 minutes at maximum flow. Furthermore, 4 post-treatments, i.e., a moving bed (MB, 300 L, average EBCT 21 min), a fixed bed (FB, 220 L, average EBCT 25 min), a fresh GAC filter (GAC_{fresh/OZO}, 77 L, i.e. 34.65 kg Cyclecarb 401, density 450 g/L, 0.425-2.36 mm granular size, average EBCT 14.5 min) and a preloaded GAC filter (GAC_{loaded/OZO}, 85 L, i.e. 40.8 kg Norit GCN 830, density 480 g/L, 1.4-2.36 mm granular size, average EBCT 18 min) were installed in pilot-scale for a comparison with sand filter, run flow proportionally at dry weather water flow. An identical fresh GAC filter (GAC_{fresh/bio}, 77 L, Cyclecarb 401, 0.425-2.36 mm granular size, average EBCT 14 min) was installed directly after the secondary clarifier. More details about the differing reactors and the operational parameters are given in Text S1.

Wastewater sampling and chemical analyses

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161 Wastewater sampling 162 Samples were taken at the effluent of the primary clarifier (WWTP-INF), of the secondary clarifier (BIO-EFF), of the ozonation reactor (OZO-EFF) and of the post-treatments mentioned above (SF-EFF, 163 164 MB-EFF, FB-EFF, GAC_{fresh/Dio}-EFF, GAC_{fresh/OZO}-EFF and GAC_{loaded/OZO}-EFF). On 37 sampling days over a 10-month period (September 2014-June 2015), samples (1 L, 24-h flow proportional) were 165 166 collected from all sampling points in intervals from 1 day at the beginning to 5 weeks at the end. Note 167 that WWTP-INF was always sampled 1 day before the others to account for the HRT of the plant. 168 Four sampling campaigns (September/October 2014) were conducted collecting 24-h flow proportional 169 samples (1 L) at WWTP-INF, BIO-EFF, OZO-EFF and SF-EFF on 3 consecutive dry days at differing 170 ozone doses of 2.0 (October 3-5), 3.0 (September 24-26), 4.0 (September 11-13) and 4.9 mg O₃/L 171 (September 15-17), corresponding to specific ozone doses of 0.35±0.02, 0.54±0.04, 0.67±0.02 and 0.97±0.07 g O₃/g DOC, respectively. Thereafter, ozonation was run at 2.7 mg/L ozone (about 0.54 g 172 O₃/g DOC), the ozone dose recommended for continuous full-scale application at the WWTP Neugut. In 173 February and March 2015, two sampling campaigns (48-h flow proportional samples for all sampling 174 175 points) were carried out to screen for 550 environmentally relevant micropollutants. The samples were collected at 4°C and were maintained at this temperature until analyses. If chemical 176 analyses could not be carried out within a week, the samples were frozen at -20°C and thawed just 177 178 before sample preparation. 179 Ouantification of 51 micropollutants and OTPs using an automatized extraction (online SPE-LC-HRMS, 180 *method I*) 181 The concentration of 43 micropollutants (acesulfame, aliskiren amisulpride, atenolol, atenolol acid, 182 azithromycin, benzotriazole, bezafibrate, candesartan, carbamazepine, carbendazim, cetirizine, 183 citalopram, clarithromycin, DEET, diclofenac, diuron, eprosartan, fexofenadine, gabapentin, hydrochlorothiazide, iopromide, irbesartan, lamotrigine, levetiracetam, losartan, mecoprop, metoprolol, 184

methylbenzotriazole, N4-acetyl-sulfamethoxazole, oxazepam, phenazone, primidone, ranitidine,

sucralose, sulfamethoxazole, telmisartan, tramadol, triclosan, trimethoprim, valsartan, valsartan acid, venlafaxine, including parent compounds and metabolites, Table S5) and eight OTPs with reference standards (4'-/5-hydroxydiclofenac, amisulpride *N*-oxide, cetrizine *N*-oxide, clarithromycin *N*-oxide, fexofenadine *N*-oxide, chlorothiazide, tramadol *N*-oxide and venlafaxine *N*-oxide) was determined according to Jeon et al. (2013) (details of method I are presented in Text S2, SI). Shortly, samples were filtered through 2 glass microfiber filters, *i.e.*, GF/D (top, 2.7 µm pore size, Whatman) and GF/F (bottom, 0.7 µm pore size, Whatman). After dilution and spiking with internal standards (IS), the samples were extracted by online solid-phase extraction with a combination of four adsorbents (Oasis HLB and a Strata-X-AW/Strata-X-CW/ENV+ mixture). The extracts were directly analyzed by liquid chromatography (Atlantis T3 column, Waters), coupled to high-resolution mass spectrometry (Q-Exactive, ThermoScientific). Details on quality control measurements (limit of quantification and recoveries) are given in Table S5.

Additionally to the eight OTPs mentioned above, other OTPs, presented in the literature (Table S6), without reference standards were screened with their exact mass of the protonated or deprotonated molecular ion. MS/MS spectra from fragmentation did not show high enough intensities for a

Ouantification of 550 micropollutants using a manual extraction (offline SPE-LC-HRMS, method II)

The fate of 550 environmentally-relevant chemicals was investigated with an offline SPE-LC-HRMS method, adapted from Kern et al. (2009) and described in Text S3, SI. This list included 240 pharmaceuticals, 234 pesticides, 38 industrial chemicals and 60 biological TPs. Their physical-chemical properties cover a wide range of molecular weight (120-915 g/mol), $\log K_{ow}$ (-2.2 – 5.7), and charge at circumneutral pH (50% neutral, 50% ionic). In brief, 1 L of sample was filtered (as described above) and spiked with IS. The samples were then extracted through a multilayered solid-phase cartridge, consisting of ENVI-Carb activated carbon (bottom), Strata-X-AW/Strata-X-CW/ENV+ mixture (middle layer), and Oasis HLB (top layer). Chemical analyses were carried out with the same LC-HRMS system as described above.

confirmation.

- 213 Determination of bromide and bromate
- For the quantification of bromide and bromate, 1 mL samples (24- and 48-h flow proportional,
- September 2014-June 2015, n=30) were filtered with a 13-mm syringe filter (regenerated cellulose
- membrane, 0.45µm porosity, Infochroma AG), diluted 100 times with ultrapure water and stored at
- 217 +4°C until analyses. Bromide and bromate were separated by capillary ion chromatography (Thermo
- Dionex ICS-4000) using an anion-exchange IC column (IonPac AS19-4μm, Dionex) and detected by a
- 219 modified tandem mass spectrometry (Thermo TSQ-Vantage) to reach lower detection limit (Shah et al.
- 220 2015). The quantification limits of bromide and bromate in wastewater were 25 μg/L and 2 μg/L,
- respectively, for the 100-fold dilution (0.25 and 0.02 µg/L, respectively, in ultrapure water without
- dilution).
- 223 Determination of N-nitrosodimethylamine (NDMA) by LC-HRMS/MS
- The concentrations of NDMA were determined in several samples collected flow proportionally in
- September-October 2014 (*n*=2, 24-h composite; n=3 72-h composite) and in February-April 2015 (*n*=3,
- 48-h composite) with LC-HRMS/MS according to a method described previously (Krauss and Hollender
- 227 2008), obtaining a limit of quantification at 5 ng/L (Text S5).
- 228 Determination of second-order rate constants
- For some relevant compounds without available kinetic data, apparent second-order rate constants at pH
- 7 ($k_{O3,pH7}$) were determined: 1.5×10^5 M⁻¹ s⁻¹ for amisulpride, 4.9×10^5 M⁻¹ s⁻¹ for eprosartan, 2.1×10^5
- M^{-1} s⁻¹ for losartan, 1.2×10^5 M⁻¹ s⁻¹ for telmisartan, 560 M⁻¹ s⁻¹ for candesartan, and 24 M⁻¹ s⁻¹ for
- 232 irbesartan (See Text S4 for details). These and other rate constants from literature are compiled in Table
- 233 S4.

RESULTS AND DISCUSSION

Micropollutant abatement.

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236 Fate of micropollutants during ozonation and over the whole wastewater treatment 237 The abatement of 43 wastewater relevant micropollutants (Table 1, parent compounds and metabolites) 238 in biological treatment with conventional activated sludge (BIO) and during ozonation (OZO) with 4 239 specific ozone doses is presented in Table 1 (concentrations can be found in Table S5). 240 During the biological treatment, the abatements were relatively constant for single compounds over the 241 sampling period (n=12), but varied considerably between -131% and >99% among them (SI, Table S8). 242 A limited number of compounds (acesulfame, bezafibrate, DEET, eprosartan, levetiracetam, N4-acetyl-243 sulfamethoxazole, triclosan, trimethoprim and valsartan) were almost completely eliminated in 244 biological treatment (relative abatement >80%). Half of the compounds (n=22) were not significantly 245 eliminated (relative abatement <20%). Some investigated compounds, e.g., lamotrigine, phenazone and 246 the TP valsartan acid, were even significantly formed during this treatment step (negative elimination or 247 detection of the compound only after BIO). For lamotrigine, this is in agreement with recent studies 248 (Bollmann et al. 2016, Zonja et al. 2016) which showed that non-conjugated and conjugated forms 249 (mainly as glucuronide) enter the WWTPs. In conventional biological treatment, conjugated forms were 250 cleaved to lamotrigine, which is relatively stable. Therefore, its concentration significantly increases, typically from about 300 to 800 ng/L (Table S5). Phenazone was also found to be formed from other 251 252 pharmaceuticals (from metamizole or human metabolites) during biological treatment (Gros et al. 2010, Kovalova et al. 2012, Prasse et al. 2010), and valsartan acid was identified as a biodegradation product 253 254 of valsartan (Kern et al. 2010). 255 During ozonation, the extent of abatement increased with increasing ozone doses (Table 1 and Fig. S4). 256 At the lowest specific ozone dose of 0.35 g O₃/g DOC, 20 out of the 43 substances were abated by less 257 than 80%. However, this number decreased significantly to 13, 11 and 5 with higher specific ozone 258 doses of 0.54 g O₃/g DOC, 0.67 g O₃/g DOC and 0.97 g O₃/g DOC, respectively. Substances with high 259 reactivity with ozone at circumneutral pH (Table S4), e.g., diclofenac, carbamazepine, 260 hydrochlorothiazide, sulfamethoxazole and tertiary amines such as cetirizine, fexofenadine and

tramadol, were very well abated (>80%) even at 0.35 g O₃/g DOC. For higher specific ozone doses, these compounds were abated by >93% and not detected anymore (<LOQ). Compounds with low apparent second-order rate constant for the reaction with ozone at circumneutral pH (i.e., $k_{O3 pH7} < 10 \text{ M}^{-1}$ s⁻¹), for which the reaction with •OH is the predominant abatement pathway, e.g., iopromide, lamotrigine 264 and sucralose, were only slightly abated (<30%) at a specific ozone dose of 0.35 g O₃/g DOC. The concentration of these compounds was reduced by less than 71% at the highest specific ozone dose (0.97 $g O_3/g DOC)$. Other studies, presented in Table S9, showed similar abatements for comparable specific ozone doses (Kovalova et al. 2013, Lee et al. 2013, Reungoat et al. 2010, Ternes et al. 2017) or slightly lower (Hollender et al. 2009, Margot et al. 2013). In the latter two studies, the abatement of compounds predominantly reacting with •OH (methylbenzotriazole, benzotriazole, gabapentine, iopromide, 272 irbesartan, mecoprop, oxazepam, primidone and valsartan) was particularly low. These differences may be attributed to varying wastewater qualities, e.g., a differing composition of the dissolved organic matter leading to a lower •OH formation (Schindler Wildhaber et al. 2015) or a higher alkalinity (HCO₃⁻ being a •OH scavenger (Buxton and Elliot 1986) in these wastewaters. In the study by Margot et al. (2013), the presence of nitrite in high concentrations (average value of 0.4 mg NO₂-N/L, compared to 0.01-0.04 mg NO₂-N/L level in other studies) explains at least partially the lower abatements. Nitrite scavenges ozone rapidly in a ratio of about 1 g O₃ per g NO₂⁻ (1:1 molar ratio), therefore, a specific ozone dose of 0.83 g O_3 /g DOC corresponds to an effective specific dose of 0.63 g O_3 /g DOC (DOC 7.3 mg/L) at 1.4 $mg NO_2^{-1}/L$ (i.e., 0.4 $mg NO_2^{-1}/L$). Over the entire treatment, including biological treatment and ozonation (BIO+OZO), most of the compounds were abated by >80% at a specific ozone dose of 0.35 g O₃/g DOC, except 10 compounds (Table 1). At specific ozone doses of 0.54 g O₃/g DOC, 0.67 g O₃/g DOC or 0.97 g O₃/g DOC, 6, 7 or 2 compounds, respectively, were not abated >80%. To comply with the guidelines established by the Swiss authorities for the evaluation of advanced wastewater treatment, 12 indicator substances need to be eliminated by 80% on average over the whole treatment chain (marked in Table 1, see Table S1 and Table S8 for more details). These 12 indicator substances were abated by 2-44% during biological treatment, except benzotriazole (64±4%).

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Benzotriazole is often poorly abated in biological treatment (Herzog et al. 2014, Liu et al. 2011), except in some specific WWTPs (Mazioti et al. 2015), for unknown reasons. The average abatement increased to $85 \pm 3\%$ when the biological treatment was followed by a low specific ozone dose (0.35 g O₃/g DOC) and even up to >94% when the specific ozone dose was 0.54 g O₃/g DOC. Note that these 12 substances are not abated in sand filtration, as will be discussed in more detail below. Considering these results, a combination of biological treatment and a specific ozone dose of 0.40 g O₃/g DOC (1.5-2.5 mg O₃/L) is sufficient to reach an average abatement of >80% of the 12 indicator substances as requested by the Swiss water protection act (FOEN 2017). However, to compensate a potential nitrite peak concentration of up to 0.7 mg NO₂/L (i.e., 0.2 mg NO₂-N/L, while the usual nitrite concentration at WWTP Neugut is 0.02-0.04 mg NO₂-N/L), a specific ozone dose of 0.55 g O₃/g DOC $(2.0-3.3 \text{ mg O}_3/\text{L})$ is recommended for routine operation at this site. Two sampling campaigns (48-h flow proportional samples) were carried out to investigate the abatement of an exhaustive list of 550 micropollutants over the whole treatment chain at a specific ozone dose of 0.55 g O₃/g DOC (Fig. 1, Fig. S5, Table S7). In WWTP-INF, 175 compounds were found (>LOQ) with a total concentration of 371 µg/L. Most abundant were pharmaceuticals (133 µg/L), caffeine (130 µg/L) and food additives (85 µg/L) in the influent, while pesticides and biocides were detected only in very low concentrations (ca. 1 µg/L). Being aware that the sampling was done in winter, it may not be representative for the whole year. In biological treatment, the total concentration of target compounds decreased by 85% to 54 µg/L, although the number of quantified compounds increased to 204 substances. This increase can be explained by the formation of biological transformation products that were included in the target list, and the lower LOQ in BIO-EFF compared to WWTP-INF due to differing dilutions used for the analyses and due to higher matrix effects in WWTP-INF (see Text S3). The 4 most abundant compounds in WWTP-INF, i.e., caffeine, cyclamate, metformin and paracetamol (total concentration 260 µg/L), were eliminated by 99%. Without the contribution of these 4 compounds, the abatement of the remaining micropollutant load drops from 85% to 54% during the biological treatment. After ozonation, the total concentration was still 23 µg/L, corresponding to an abatement of the micropollutants load by 58%. In sand filtration, no significant elimination was observed, and over the whole plant, the total load decreased by 94%. Five compounds (acesulfame, sucralose, metformin,

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iopromide and tris(2-chloropropyl)phosphate) were found in concentrations $>0.9 \mu g/L$ in OZO-EFF. All were poorly abated during ozonation (<50%). Alternatively, the abatement can be considered for each single substance and an average can be calculated. As a result, the average abatement of all substances (taking a zero abatement for all substances with negative abatement into account) over the biological treatment is 34% (median 22%), during ozonation >67% (median >74%), and over the whole WWTP, with and without sand filter, >79% (median >89%) (Fig. 1). Due to the fact that 64 substances (out of 147) were not detected anymore after ozonation, only a minimal abatement relative to the LOQ could be calculated in these cases.

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	0.35±0.02 g O ₃ /g DOC			0.54±0.05 g O ₃ /g DOC			0.67±0.03 g O ₃ /g DOC			0.97±0.07 g O ₃ /g DOC	
	BIO	OZO	BIO+OZO	BIO	OZO	BIO+OZO	BIO	OZO	BIO+OZO	BIO	OZO
Acesulfame	95 ± 1	39 ± 5	97 ± 1	90 ± 8	59 ± 9	96 ± 3	95 ± 1	70 ± 8	98 ± 1	88 ± 5	>90 ± 4
Aliskiren	25 ± 4	$> 84 \pm 1$	$> 88 \pm 1$	23 ± 22	$>93 \pm 1$	>95 ± 1	7 ± 12	$> 81 \pm 1$	$> 82 \pm 3$	8 ± 11	$>92 \pm 3$
Amisulpride†	2 ± 14	91 ± 1	91 ± 2	3 ± 16	$>98 \pm 1$	>98 ± 1	-1 ± 13	$>95 \pm 1$	>95 ± 1	3 ± 8	$>98 \pm 1$
Atenolol	75 ± 3	70 ± 2	92 ± 1	70 ± 5	92 ± 3	98 ± 1	72 ± 5	93 ± 5	98 ± 1	71 ± 4	$>93 \pm 1$
Atenolol acid	76 ± 2	70 ± 3	93 ± 1	72 ± 3	91 ± 3	98 ± 1	67 ± 4	$>92 \pm 4$	$>97 \pm 1$	69 ± 3	$>98 \pm 1$
Azithromycin	8 ± 28	$>90 \pm 1$	$>90 \pm 3$	n.m.2	$>54 \pm 3$	n.m.	25 ± 14	$> 88 \pm 2$	$>91 \pm 3$	23 ± 15	$>95 \pm 1$
Benzotriazole†	66 ± 3	52 ± 1	83 ± 1	62 ± 7	74 ± 3	90 ± 3	63 ± 6	80 ± 7	93 ± 2	64 ± 3	91 ± 4
Bezafibrate	96 ± 1	$>62 \pm 5$	>98 ± 1	$>95 \pm 2$	$>75 \pm 1$	$>99 \pm 1$	$>93 \pm 2$	n.m.1	$>97 \pm 1$	>94 ± 1	$>$ 24 $^{\Delta}$
Candesartan†	0 ± 16	63 ± 1	63 ± 6	1 ± 12	82 ± 3	82 ± 5	-24 ± 20	85 ± 6	81 ± 6	-17 ± 19	94 ± 3
Carbamazepine†	-16 ± 12	95 ± 1	94 ± 1	-14 ± 19	$>98 \pm 1$	$>98 \pm 1$	-33 ± 23	$>98 \pm 1$	>98 ± 1	-24 ± 8	$>98 \pm 1$
Carbendazim	8 ± 18	82 ± 2	83 ± 5	-1 ± 23	$>94 \pm 3$	>94 ± 4	$7 \pm 16^{\#}$	>93 ± 5	$> 89 \pm 8^{\#}$	-28 ± 50	$> 89 \pm 4$
Cetirizine	6 ± 12	92 ± 1	92 ± 2	-9 ± 12	$>93 \pm 1$	>92 ± 1	-25 ± 25	$>95 \pm 1$	>94 ± 1	-19 ± 10	$>95 \pm 1$
Citalopram†	$5 \pm 4^{\#}$	91 ± 3	$89 \pm 4^{\#}$	4 ± 4	>96 ± 3	$>96 \pm 3$	-8 ± 2	>94 ± 1	>93 ± 1	4 ± 8	>97 ± 1
Clarithromycin†	51 ± 2	93 ± 2	97 ± 1	28 ± 19	$>95 \pm 1$	$>96 \pm 1$	45 ± 6	$>94 \pm 1$	$>97 \pm 1$	52 ± 8	$>97 \pm 1$
DEET	96 ± 1	49 ± 9	98 ± 1	$>90 \pm 3$	65 ± 1	$>95 \pm 2$	97 ± 1	52 ± 18	99 ± 1	97 ± 1	70 ± 15
Diclofenac†	22 ± 8	96 ± 1	97 ± 1	23 ± 8	100 ± 1	100 ± 1	11 ± 10	$>99 \pm 1$	>99 ± 1	13 ± 18	$>99 \pm 1$
Diuron	0 ± 30	64 ± 2	64 ± 10	-7 ± 7	$> 84 \pm 8$	>83 ± 10	-5 ± 14	>84 ± 6	>84 ± 4	$-40 \pm 72^{\#}$	>90 ± 6
Eprosartan	98 ± 1	$>66 \pm 3$	>99 ± 1	>93 ± 2	n.m.1	$>97 \pm 1$	$>97 \pm 1$	$>67 \pm 7$	$>99 \pm 1$	$> 88 \pm 5$	n.m.1
Fexofenadine	13 ± 16	83 ± 3	85 ± 6	9 ± 4	$>94 \pm 1$	>95 ± 1	12 ± 12	$>96 \pm 3$	$>97 \pm 2$	-1 ± 12	$>91 \pm 1$
Gabapentin	44 ± 10	44 ± 4	69 ± 6	43 ± 6	55 ± 4	75 ± 1	37 ± 5	63 ± 10	77 ± 6	44 ± 5	75 ± 6
Hydrochlorothiazide†	9 ± 13	86 ± 2	87 ± 4	13 ± 9	$>98 \pm 2$	$>98 \pm 2$	-2 ± 17	$> 97 \pm 3$	$>97 \pm 2$	1 ± 10	$>99 \pm 1$
Iopromide	72 ± 5	28 ± 1	80 ± 4	70 ± 8	43 ± 3	83 ± 5	53 ± 20	53 ± 6	78 ± 8	$64 \pm 8^{\#}$	$65 \pm 1^{\#}$
Irbesartan†	17 ± 18	57 ± 1	64 ± 8	15 ± 15	75 ± 3	78 ± 5	2 ± 20	79 ± 4	79 ± 6	0 ± 25	89 ± 4
Lamotrigine	-125 ± 31	37 ± 1	-42 ± 18	-128 ± 10	50 ± 2	-15 ± 4	-140 ± 30	57 ± 7	-3 ± 21	-132 ± 5	71 ± 6
Levetiracetam	99 ± 1	$>66 \pm 34$	100 ± 1	98 ± 1	>43 ± 36	>99 ± 1	$>99 \pm 1$	n.m.1	100 ± 1	$>99 \pm 1$	n.m.1
Losartan	71 ± 2	93 ± 1	98 ± 1	68 ± 5	$>98 \pm 1$	$>99 \pm 1$	66 ± 5	$>98 \pm 1$	$>99 \pm 1$	67 ± 4	$>98 \pm 1$

Mecoprop	$-5 \pm 2^{\#}$	59 ± 2	$56 \pm 1^{\#}$	$14 \pm 7^{\#}$	77 ± 2	$81 \pm 4^{\#}$	5 ± 30	82 ± 7	84 ± 5	$9 \pm 14^{\#}$	>91 ± 3
Metoprolol†	41 ± 7	75 ± 2	85 ± 3	38 ± 5	94 ± 3	96 ± 2	37 ± 8	94 ± 5	96 ± 3	38 ± 4	$>99 \pm 1$
Methylbenzotriazole†,*	45 ± 22	66 ± 3	81 ± 9	61 ± 4	89 ± 4	96 ± 2	2 ± 22	90 ± 1	90 ± 7	28 ± 12	98 ± 1
N4-Acetyl-Sulfamethoxazole	$>98 \pm 1$	n.m.1	$>99 \pm 1$	$>98 \pm 1$	n.m.1	$>99 \pm 1$	$>98 \pm 1$	n.m.	$>99 \pm 1$	$>95 \pm 2$	n.m.1
Oxazepam	5 ± 17	55 ± 1	57 ± 7	10 ± 10	73 ± 3	75 ± 5	-13 ± 22	77 ± 5	74 ± 6	-10 ± 9	88 ± 4
Phenazone	n.m.2	$>91 \pm 1$	n.m.1	n.m.2	$>92 \pm 1$	n.m.1	n.m.2	$>92 \pm 1$	n.m.1	n.m.2	$> 81 \pm 4$
Primidone	19 ± 12	48 ± 3	58 ± 5	15 ± 17	66 ± 1	72 ± 6	14 ± 14	70 ± 10	75 ± 7	-4 ± 8	86 ± 5
Ranitidine	82 ± 1	$>74 \pm 2$	$>95 \pm 1$	79 ± 4	$> \! 80 \pm 4$	$>96 \pm 1$	66 ± 16	$>71 \pm 2$	$>90 \pm 5$	77 ± 2	$>53 \pm 19$
Sucralose	9 ± 20	21 ± 2	28 ± 14	13 ± 13	27 ± 1	36 ± 10	-9 ± 17	38 ± 4	32 ± 12	-4 ± 13	46 ± 4
Sulfamethoxazole	55 ± 3	85 ± 2	93 ± 1	46 ± 6	$>97 \pm 1$	$>98 \pm 1$	31 ± 6	>97 ± 1	$>98 \pm 1$	34 ± 2	$>95 \pm 3$
Telmisartan	15 ± 3	66 ± 1	71 ± 1	3 ± 16	86 ± 3	86 ± 4	-24 ± 27	88 ± 5	84 ± 8	-7 ± 18	$>94 \pm 1$
Tramadol**	0 ± 11	91 ± 1	90 ± 3	2 ± 7	98 ± 1	98 ± 1	-5 ± 11	$>98 \pm 1$	$>98 \pm 1$	-6 ± 10	$>98 \pm 1$
Triclosan	n.m.1	n.m.1	n.m.1	n.m.1	n.m.1	n.m.1	n.m.1	$>56 \pm 1$	n.m.1	n.m.1	n.m.1
Trimethoprim	$> 89 \pm 4$	$>59 \pm 6$	$>95 \pm 1$	85 ± 6	$>61 \pm 15$	$>95 \pm 1$	75 ± 6	$>58 \pm 4$	$>90 \pm 1$	83 ± 3	$>79 \pm 3$
Valsartan	94 ± 1	52 ± 3	97 ± 1	93 ± 2	69 ± 3	98 ± 1	96 ± 1	68 ± 8	99 ± 1	93 ± 1	$> 86 \pm 3$
Valsartan acid	n.m.2	49 ± 1	n.m.1	n.m.2	67 ± 1	n.m.1	n.m.2	71 ± 5	n.m.1	n.m.2	76 ± 8
Venlafaxine†	4 ± 15	84 ± 2	84 ± 5	9 ± 10	$>97 \pm 2$	$>97 \pm 2$	-12 ± 17	$>96 \pm 3$	$>96 \pm 2$	1 ± 5	$>96 \pm 1$
Average of the 12 indicator	17 ± 8	79 ± 1	85 ± 3	20 ± 7	>91 ± 2	>94 ± 2	7 ± 8	>92 ± 3	>93 ± 2	14 ± 9	$>96 \pm 1$

n.m.1: abatement not measurable because the concentrations at the inlet and at the outlet of the reactor were both <LOQ. 330

substances†

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n.m.2: abatement not measurable because the concentration at the inlet is <LOQ; abatement has a negative value. 331

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^{*:} methylbenzotriazole represents the sum of 4- and 5-methylbenzotriazole, which co-elute under the analytical conditions.

**: tramadol represents the sum of tramadol and o-desmethyl-venlafaxine, which are isobaric compounds that co-eluted under the analytical conditions.

#: abatement was determined for only 2 samples due to high fluctuations of concentrations in one sample.

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^Δ: abatement was determined for only 1 sample due to high fluctuations of concentrations in two samples.

^{†:} the 12 indicator substances (**bold** in the table above) are amisulpride, benzotriazole, candesartan, carbamazepine, citalopram, clarithromycin, diclofenac, hydrochlorothiazide, irbesartan, methylbenzotriazole, metoprolol and venlafaxine.

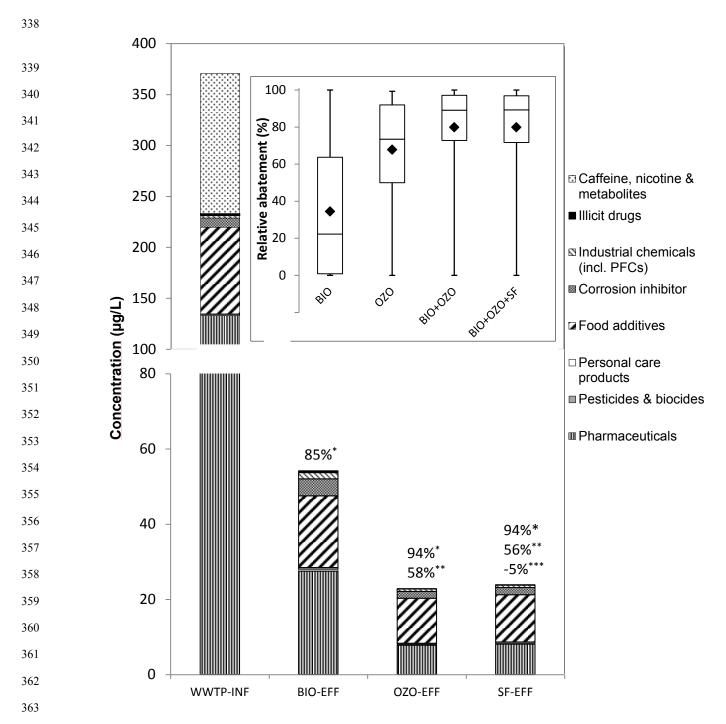


Fig. 1. Sum of concentrations of micropollutants at various WWTP sampling points (*n*=2, 48-h flow proportional samples, specific ozone dose 0.55 g O₃/g DOC, analyzed with method II; for sampling points see Fig. S1). *: abatement of total concentration relative to WWTP-INF (effluent of primary clarifier); **: abatement relative to BIO-EFF (effluent of conventional biological treatment); ***: abatement relative to OZO-EFF (effluent of ozonation reactor). Inset: Boxwhisker plot depicting the distribution of calculated abatements for all substances (minimum, 25th percentile, median, 75th percentile, maximum and (♠) average) of micropollutants (analyzed with method II) during all treatment steps. Because nearly half of the compounds were not detected anymore after ozonation, the values given for OZO, BIO+OZO and BIO+OZO+SF represent minimal abatements relative to the LOQ. Negative abatements were taken into account as zero abatements.

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376 Several post-treatments, i.e., sand filtration (SF), moving bed (MB), fixed bed (FB), a fresh granular 377 activated carbon filter (GAC_{fresh/OZO}) and a pre-loaded granular activated carbon filter (GAC_{loaded/OZO}), 378 were evaluated for their capacity to abate residues of micropollutants still present after ozonation with 379 an ozone dose of 2.7 mg/L ozone (about 0.54 g O₃/g DOC). An additional fresh granular activated carbon filter (GAC_{fresh/BIO}) was fed with secondary clarifier effluent for a comparison with GAC_{fresh/OZO}. 380 381 Amongst 20 micropollutants (including the indicator substances) frequently detected after ozonation in 382 concentrations >50 ng/L, none showed a significant relative abatement (all <16%) in the three solely 383 biologically active post-treatments, i.e., sand filtration, moving bed and fixed bed (Fig. S8). 384 Similarly, a screening of the selected 550 substances showed that these 3 post-treatments do not 385 significantly impact the number (Fig. S5) and the total concentrations of the compounds detected in the 386 ozonated effluent (22.9 μg/L after OZO vs. 24.0 μg/L after SF, 23.5 μg/L after MB and 23.5 μg/L after 387 FB, Fig. S6). The average abatement did not exceed 5% in these biological post-treatments (Fig. S7). 388 Even the compounds abated by >50% during the conventional biological treatment (Table S8), i.e., 389 acesulfame, atenolol, benzotriazole and valsartan, showed a low abatement of <7% in these three post-390 treatments, due to shorter retention times (10-25 min) compared to the conventional biological treatment 391 (18 h).The abatements in two identical fresh activated carbon filters, GAC_{fresh/BIO} and GAC_{fresh/OZO}, was 392 393 investigated for four compounds with very good sorption on activated carbon (Group I, see Table S1 for 394 definition), two compounds with good sorption (Group II) and two compounds with poor sorption 395 ability (Group III). This grouping is in agreement with the abatement by ozone, and was based on results from sorption to powdered activated carbon, because physical-chemical properties of charged, polar 396 397 micropollutants like logD_{ow} are not sufficient to predict their sorption behavior (Kovalova et al. 2013). A 398 significant difference in the breakthrough of the compounds was observed. Group I compounds (Fig. 2a 399 and b) showed a much later breakthrough compared to the other compounds with weaker sorption 400 behavior (Fig. 2c and d). Surprisingly, benzotriazole from group II was relatively well abated, as discussed later. In GAC_{fresh/OZO}, the breakthrough of Groups I, II, III (Fig. 2) and the 10 additional 401 402 substances telmisartan, valsartan, valsartan acid, primidone, atenolol, atenolol acid, gabapentin, tramadol, lamotrigine, and oxazepam (Fig. S9-S10) usually occurred later than in GAC_{fresh/BIO}. For instance, the abatement of group II/III compounds candesartan and sucralose dropped below 90% at about 3'200 BV in GAC_{fresh/BIO} but only at about 7'000 BV in GAC_{fresh/OZO}. As GAC_{fresh/OZO} followed ozonation, the concentrations of the micropollutants were lower and, at the same time, the composition of the matrix is different. It was shown that the breakthrough is independent on micropollutant concentration within certain limits (Zietzschmann et al. 2016). However, during ozonation, effluent organic matter is transformed to smaller compounds, which are generally less aromatic, more hydrophilic and charged, thereby exhibiting a poorer adsorbability on GAC. Measurements showed that small neutral organic substances are further oxidized to charged small organic acids during ozonation (Ramseier and von Gunten 2009, Zietzschmann et al. 2015). As a consequence, micropollutants in ozonated wastewater have fewer competitors for adsorption sites compared to non-ozonated wastewater and the breakthrough of micropollutants may occur later in ozonated wastewater. To compare the two GAC filters, the average relative abatement of the indicator substances was evaluated. Because some of the 12 indicator substances are present in too low concentrations after ozonation to determine an appropriate abatement, only 6 were selected: 4 compounds from Group I (diclofenac, hydrochlorothiazide, metoprolol, venlafaxine), and 2 compounds from Group II (benzotriazole and candesartan). This selection criteria is also recommended by the Swiss legislation in case of low inflow concentrations. An average relative abatement of >80% for the 6 selected indicator substances could be kept up to 10'000 BV for GAC_{fresh/BIO} and up to 15'000 BV for GAC_{fresh/OZO} (Fig. S11). At the two last sampling times for 22'000 and 28'000 BV, the sorption on GAC_{fresh/BIO} increased again. Currently, this effect cannot be explained and would require more investigations for a clarification. In the screening of the 550 substances, the decrease of total concentration in GAC_{fresh/BIO} and GAC_{fresh/OZO} was comparable (47% in GAC_{fresh/BIO} and 41% in GAC_{fresh/OZO} at similar BV in the range of 11'000-17'000, Fig. S6). Similarly, the average abatement calculated from the abatement of the same selection of substances was comparable for GAC_{fresh/BIO} and GAC_{fresh/OZO} (54% and 53%, respectively, at the same BV) (Fig. S7).

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The performance of GAC_{fresh/BIO} can be used to evaluate the effectiveness of GAC filtration. Since this GAC filter has a volume of 77 L and a filling of 34.65 kg GAC, 10'000 BV (where the average abatement of the six indicator substances fell below the limit of 80%) correspond to a carbon usage rate of 45 mg GAC/L. To be comparable to the elimination efficiency of powdered activated carbon, about 10 mg/L activated carbon (corresponding to 45'000 BV) should be reached before breakthrough of 80% of the indicator substances (Boehler et al. 2012b). However, a stepwise renewal of several GAC filter cells running in parallel (with differing elimination efficiencies between 100% and below 80%) allows optimal performance with a longer lifetime of the GAC filters (Benstoem et al. 2017). Moreover, it was demonstrated recently that a minimal EBCT is crucial to achieve a good elimination (Wunderlin et al. 2017). An EBCT of 25 minutes (compared to 15 minutes in this study) turned out to be sufficient for a good performance for a 1.18 - 2.36 mm granulation at low DOC.

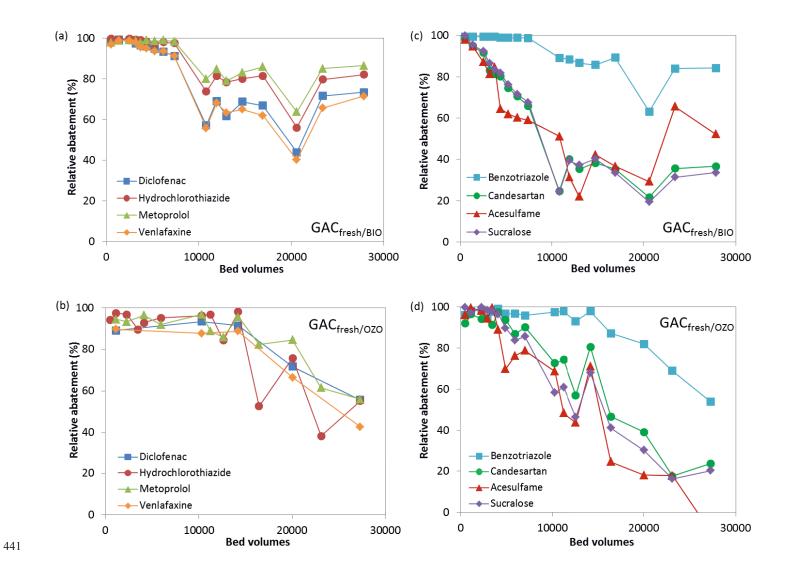


Fig. 2. Relative abatements of group I substances (diclofenac, hydrochlorothiazide, metoprolol and venlafaxine) in (a) $GAC_{fresh/BIO}$ and (b) $GAC_{fresh/OZO}$; of group II (benzotriazole and candesartan) and group III substances (acesulfame and sucralose) in (c) $GAC_{fresh/BIO}$ and (d) $GAC_{fresh/OZO}$ as a function of the bed volumes. 28'000 BV correspond to a run time of 10 months.

GAC_{loaded/OZO} had been pre-loaded for 16'000 BV with the effluent of the secondary clarifier from another WWTP (Boehler et al. 2012a), before being installed at WWTP Neugut after ozonation. During this loading phase of GAC_{loaded/OZO}, the abatement of selected micropollutants was measured and can be compared to the abatement in GAC_{fresh/BIO} at WWTP Neugut. Micropollutants abatement in GAC_{loaded/OZO} (Fig. 3 and Fig. S12, yellow zone) was less efficient than in GAC_{fresh/BIO} (Fig. 2 and Fig. S9), even though the EBCT was slightly higher in GAC_{loaded/OZO} (18 min for GAC_{loaded/OZO} vs. 15 min for GAC_{fresh/BIO}). For instance, venlafaxine and sucralose broke through immediately in GAC_{loaded/OZO} and their respective abatement at 7'500 BV were around 40% and 30% in GAC_{loaded/OZO}, while still at 91% and 68% in GAC_{fresh/BIO}, respectively. The wastewater quality (DOC 6.9 mg/L in GAC_{loaded/OZO} vs. DOC 5.5 mg/L in GAC_{fresh/BIO}) and the GAC properties (different material; higher particle size of 1.4-2.36 mm in GAC_{loaded/OZO} compared to 0.425-2.36 mm in GAC_{fresh/BIO}) likely explain why breakthrough occurred faster in GAC_{loaded/OZO}. When GAC_{loaded/OZO} was operated with ozonated wastewater (>16'000 BV, to the right of the yellow zone in Fig. 3), abatements were not significantly different to the abatements observed during the preloading phase (yellow zone in Fig. 3). Only diclofenac showed a significant increase in the relative abatement in the second phase: its relative abatement was <20% at the end of the first phase at 9'000 BV and 62% when measurements restarted at 23'000 BV. Moreover, the micropollutant's relative abatements decreased only slightly in GAC_{loaded/OZO} over the sampling period between 23'000-50'000 BV. Similarly to GAC_{fresh/BIO} and GAC_{fresh/OZO}, the relative abatement of poorly sorbing compounds such as acesulfame, candesartan, gabapentin, irbesartan, sucralose, and valsartan acid, was <20% at high BVs (Fig. 3 and Fig. S12). Conversely, abatement was generally still >60% for diclofenac, hydrochlorothiazide, metoprolol, benzotriazole, methylbenzotriazole and atenolol, even at 50'000 BV. When comparing GAC_{loaded/OZO} with GAC_{fresh/OZO} at 28'000 BV, the abatement of all substances was similar, with the exception of benzotriazole. This compound was significantly better abated in GAC_{loaded/OZO} than in GAC_{fresh/OZO} (78% vs. 54%). The results of diclofenac and benzotriazole suggest that, in addition to sorption, a biological degradation may also take place in GAC_{loaded/OZO}, in analogy to a previous publication (Reungoat et al. 2012). However, it is not possible to differentiate between sorption and degradation by just measuring changes in concentrations. Fig. S11 shows that the average

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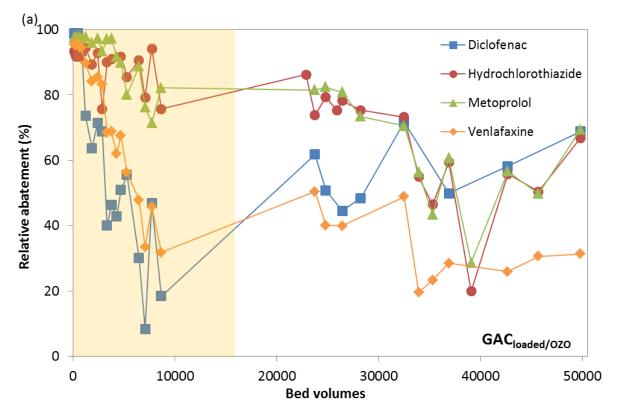
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relative abatement of the 6 selected indicator substances (benzotriazole, candesartan, diclofenac, hydrochlorothiazide, metoprolol, venlafaxine) in GAC_{loaded/OZO} was still about 40-50% at 50'000 BV, similar to GAC_{fresh/OZO} at 28'000 BV. In the screening of the 550 substances, the average abatement in GAC_{loaded/OZO} (19% at 35'000-40'000 BV; see Fig. S7) was less efficient than for the 6 selected indicator substances. The reduction of the total concentration was only 6% in GAC_{loaded/OZO} at 35'000-40'000 BV (Fig. S6). This low relative abatement in GAC_{loaded/OZO} can be attributed to the very poor abatement of the compounds still present in high concentrations in OZO-EFF (>0.9 μg/L), *i.e.*, acesulfame, sucralose, metformin, iopromide and tris(2-chloropropyl)phosphate, making up 73% of the overall concentration. These compounds are hydrophilic and known to poorly adsorb to activated carbon (Kovalova et al. 2013, Margot et al. 2013, Scheurer et al. 2010).



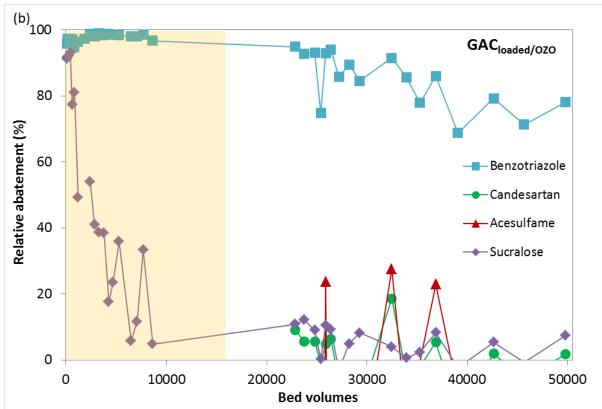


Fig. 3. Relative abatements of (a) group I substances (diclofenac, hydrochlorothiazide, metoprolol and venlafaxine) and (b) group II (benzotriazole and candesartan) and group III substances (acesulfame and sucralose) in $GAC_{loaded/OZO}$. The filter was loaded with the secondary effluent of another WWTP until 16'000 bed volumes (yellow area) and thereafter with ozonated effluent from the WWTP Neugut.

Formation and fate of AOC and fate of DOC

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Fig. S13 shows that AOC increased during ozonation at the recommended specific ozone dose (0.55 g O₃/g DOC) by a factor of 3: the AOC level in BIO-EFF was 90 µg C/L and reached about 280 µg C/L at OZO-EFF. The increase of AOC is associated with the oxidation of dissolved organic matter (DOM) in secondary effluent to form biodegradable and potentially toxic products, such as aldehydes, ketones or carboxylic acids (von Gunten 2003). These classes of compounds are generally well removable during biological post-treatments of ozonated drinking water (Hammes et al. 2006). Among the biological posttreatments studied here, the sand filter performed best in reducing the AOC by 40% (Fig. S14), similar to AOC reduction (14-46%) in other sand filters (Zimmermann et al. 2011). MB and FB reactors only slightly reduced AOC (by 17% and 13%, respectively). In FB, an intense growth of snails (*Physidae*) was observed, preventing the formation of a biofilm, which explains the low performance of this filter. The GAC filters showed the highest abatement of AOC (72% in GAC_{fresh/OZO} at 15'000 BV and 57% in GAC_{loaded/OZO} at 35'000 BV) due to additional sorption capability. Ozonation alone induced a relative abatement of DOC of about 10% (Fig. S15), which is relatively high in comparison to other studies where the DOC concentration almost did not change (Haslinger et al. submitted, Hollender et al. 2009). The DOC concentration after ozonation was poorly abated in in MB and FB (6% and 5%, respectively). In contrast, the sand filter showed a better relative abatement of DOC (20%), due to biodegradation of degradable compounds and the deposition of colloidal organic substances on the sand filter material. The GAC filters performed similar to sand filtration with a relative DOC abatement of 15-20% on GAC_{fresh/OZO} or GAC_{loaded/OZO} at 20'000-50'000 BV. The loading of DOC at the beginning (up to 15'000 BV) onto GAC_{fresh/bio} with biologically treated wastewater occurred slightly faster than onto GAC_{fresh/OZO} with the ozonated effluent (Fig. S15-S16). The same behavior was observed by Haslinger et al. (submitted). Similar to DOC, the removal of total suspended solids (TSS) was less efficient in MB or FB (4% or 17%, respectively) than in the filter systems, where 75% TSS removal was achieved in SF and 70-80%

in the GAC filters (Fig. S17). A filter cloth installed after the MB could enhance the overall TSS elimination to 50%. TSS was relatively low at around 2 mg/L in the influent and effluent of ozonation.

Formation and fate of bromate

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The concentration of bromide in the influent to the ozonation reactor (n=30) was on average 60 µg/L ranging 35-85 μ g/L (n=26), but a few exceptionally high values (up to 150 μ g/L) were measured (Table S10). These values correspond to common bromide concentrations in Swiss municipal WWTPs without discharges from waste incinerators, chemical industries or landfills (Schindler Wildhaber et al. 2015). Bromate, an ozonation product from the oxidation of bromide, was also detected in WWTP-INF and BIO-EFF, mostly below 1 μ g/L (n=20), with a maximum concentration of 17 μ g/L, concurring with a relatively high bromide concentration (85 µg/L). The presence of bromate before ozonation may be attributed to a pre-oxidation in one of the industrial sites connected to the WWTP. During ozonation, bromide was oxidized to bromate with increasing yields for increasing specific ozone doses, reaching 5.7 μg/L at 0.97 g O₃/g DOC with a bromide level of 60 μg/L (Fig. S18). The bromate yield was calculated as the ratio between the produced bromate and the bromide concentration in the influent (Fig. 4). For specific ozone doses ≤ 0.45 g O₃/g DOC, the bromate yield was low (<1%, w/w). In this range, ozone and •OH exposures are very low, because ozone is rapidly depleted due to the multiple reactions of ozone with DOM (Chon et al. 2015, Soltermann et al. 2017). For specific ozone doses >0.45 g O₃/g DOC, the lifetime of ozone increases steadily with an increasing bromate yield reaching 14% for 1.07 g O₃/g DOC. For the recommended specific ozone dose of 0.55 g O₃/g DOC, the bromate yield is approximately 3%. At this ozone dose, the bromate formation corresponds to 1.8 µg/L for the average concentration of bromide (60 µg/L), and in worse case 4.5 µg/L at the rare maximum concentration of bromide (150 µg/L). As expected, Fig. S19 shows no significant changes of bromide or bromate concentration in all post-treatments. Since bromate was mostly below 1 µg/L in BIO-EFF, its concentration in the WWTP effluent would range normally between 1 and 3.5 µg/L at bromide levels below 85 µg/L. Taking into account that, after discharge in the river, wastewater effluent is diluted by an average factor of 20 at this site, the WWTP input on bromate concentration in the receiving water is low (normally below 0.18 μg/L, with a maximal level of 0.28 μg/L at the highest bromide concentration), well below the environmental quality standard (50 μ g/L) (Ecotox Center 2017), the usual drinking water standards (10 μ g/L) (Commission Directive 2003, U.S. EPA 2006, WHO 2011), and even below the most stringent guidelines (1 μ g/L) applied for instance in the Netherlands (Knol et al. 2015). In the current study, once a bromate concentration of 17 μ g/L (Table S10) was observed in WWTP-INF, which would lead to a bromate concentration as high as 21.5 μ g/L in the effluent. In this case, the sources of bromate should be identified and measures should be taken to minimize the bromate discharge.

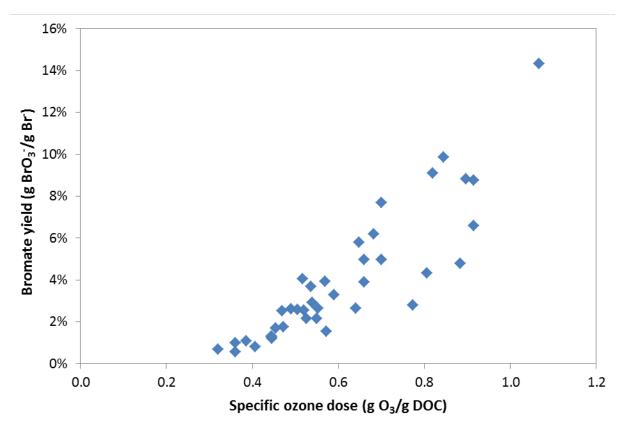


Fig. 4. Bromate yields (w/w) in the full-scale ozonation for a range of specific ozone doses from $0.32-1.07 \text{ g O}_3/\text{g DOC } (n=30)$.

Formation and fate of nitrosamines

NDMA was regularly detected in relatively high concentrations after the biological treatment (BIO-EFF), with a maximum concentration of 190 ng/L. This is in accordance with previous measurements in 20 WWTPs in Switzerland where after the biological treatment NDMA concentrations were found from <LOQ to 188 ng/L (Krauss et al. 2009). In the US, NDMA concentrations in non-chlorinated effluents were found up to 380 ng/L (Pehlivanoglu-Mantas et al. 2006, Plumlee and Reinhard 2007, Sedlak et al. 2005). A large variation was also observed from day to day and among differing WWTPs. At WWTP Neugut, the maximum NDMA formation during ozonation for specific ozone doses in the range of 0.54-

0.97 g O₃/g DOC (Fig. 5) was 30 ng/L. No correlation was observed between the formation of NDMA and the specific ozone dose, in agreement to other studies (Hollender et al. 2009, Schindler Wildhaber et al. 2015, Zimmermann et al. 2011). Thus, the secondary clarifier effluent composition with likely changing concentrations of differing NDMA precursors (not determined here) and varying NDMA formation rates affects the extent of NDMA more than the specific ozone dose. In sand filtration, NDMA was significantly abated always reaching values below 50 ng/L in the WWTP effluent (Fig. 5). Therefore, the expected concentration in the receiving waters after a 20-fold dilution is well below the WHO drinking water guideline of 100 ng/L and also below the tentative drinking water value of 10 ng/L in several countries like Germany (Selin 2011).

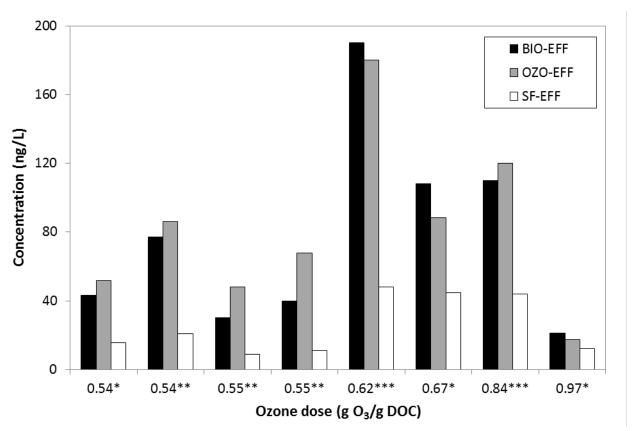


Fig. 5. Concentrations of NDMA in BIO-EFF, OZO-EFF and SF-EFF for various specific ozone doses ranging between 0.54-0.97 g O₃/g DOC. *: 72-h flow proportional samples; **: 48-h flow proportional samples; ***: 24-h flow proportional samples.

NDMA was abated in all of the investigated post-treatments (Fig. S20). Among the solely biological post-treatments, SF showed the best relative degradation of NDMA by 66%, while MB and FB showed slightly lower relative eliminations (48% and 41%, respectively). The relatively fresh filter GAC_{fresh/OZO}

- (12'500-16'500 BV) showed the highest abatement of 83%, while GAC_{loaded/OZO} at higher BV (35'000-
- 40'000 BV) exhibited a similar efficiency as SF (with 56% removal of NDMA).

Fate of transformation products

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Formation of transformation products during ozonation

581 Sixty-three ozonation transformation products from a total of 19 parents were screened for, based on 582 evidence of formation in literature (Table S6). Eighteen of them could be quantified because standards were available. However, ten out of these (3-[(4-chlorobenzoyl)amino] propanoic acid, 1-[(4-chlorobenzoyl)amino] 583 chlorophenyl) phenylmethyl] piperazine, 4-chlorobenzophenone, N-desmethyl-clarithromycin, DCPMU, 584 azacyclonol, gabapentin acid, ranitidine S-oxide, N-desmethyl-tramadol and N,N-didesmethyl-tramadol) 585 586 were detected in none of the ozonated wastewater samples. 4'-/5-hydroxydiclofenac was detected in high concentrations in most of the wastewater samples, however, its concentration dramatically 587 decreased after ozonation, independently of the specific ozone dose, and was therefore not evaluated 588 589 further. The following discussion will focus on seven OTPs frequently detected in wastewater samples 590 (Merel et al., 2017). 591 Some of these OTPs are also known to be human metabolites, biological transformation products or 592 used as active ingredients, and were therefore already detected in the wastewater after the conventional 593 biological treatment (Table S5). For instance, chlorothiazide was detected in all BIO-EFF samples 594 (n=12) at concentrations ranging 45-67 ng/L during the evaluation of the 4 specific ozone doses. 595 Tramadol N-oxide (n=5), clarithromycin N-oxide (n=3) and cetirizine N-oxide (n=1) were detected at 596 least once in samples from biological treatment, at a maximum concentration of 35 ng/L. 597 The formation of the seven OTPs and their molar yields decreased with increasing specific ozone doses 598 (Fig. S21 and Table S11). For instance, in the case of chlorothiazide, the concentration of its parent 599 compound, hydrochlorothiazide, was relatively constant in BIO-EFF over the sampling period (ca. 1000) 600 ng/L), but chlorothiazide concentrations dropped from 421-446 ng/L at 0.35 g O₃/g DOC to 163-308 ng/L at 0.97 g O₃/g DOC, respectively, corresponding to a decrease of the molar yield from 40% to 601 602 21%. At the specific ozone dose of 0.54 gO₃/gDOC, only clarithromycin N-oxide (62-74 ng/L), 603 chlorothiazide (364-385 ng/L), fexofenadine N-oxide (111-171 ng/L), tramadol N-oxide (76-82 ng/L) and venlafaxine *N*-oxide (27-30 ng/L) were >LOQ in OZO-EFF. Highest molar yields of the OTPs were observed for chlorothiazide (36%), clarithromycin *N*-oxide (34%), and fexofenadine *N*-oxide (30%) at this specific ozone dose. Amisulpride *N*-oxide and cetirizine *N*-oxide were detected only at 0.35 g O₃/g DOC (*n*=3), but their yields did not exceed 4% and 3%, respectively. The selected OTPs were formed rapidly during ozonation due to the high ozone-reactivity of the parent compounds (*k*_{O3}>10⁴ M⁻¹ s⁻¹) as recently determined in batch experiments with ozone alone for chlorothiazide, fexofenadine *N*-oxide and cetirizine *N*-oxide (Borowska et al. 2016), but were subsequently further transformed mostly by attack of hydroxyl radical.

For the 45 OTPs without available standard, normalized peak areas were determined in the samples (Fig. S22). Out of these potential OTPs, the exact masses of only 12 were detected after ozonation: four OTPs from hydrochlorothiazide (HCT_TP1-4, Table S6), two from carbamazepine (BQM and BaQD), two from phenazone (PHE_TP2 and TP6), two from sulfamethoxazole (SMX_TP1 and TP3), one from atenolol (ATE_TP1), and one from lamotrigine (LAM_TP2). These results are in agreement with the studies listed in Table S6, where these OTPs were found to be predominantly formed during the ozonation of parent compounds.

Fate of the transformation products during post-treatments

Among the seven quantified OTPs, only five OTPs (chlorothiazide, clarithromycin N-oxide, tramadol N-oxide, venlafaxine N-oxide and fexofenadine-N-oxide) were measured regularly in concentrations well above the LOQ and could be evaluated in the post-treatments. SF, MB and FB were all inefficient to biodegrade these OTPs (Fig. S23). This shows that the investigated OTPs are very stable compounds, not being degradable in the selected post-treatments.

In contrast to the three biological post-treatments, $GAC_{fresh/OZO}$ was efficient to remove the selected OTPs due to sorption (Fig. 6 and Fig. S24). Up to about 8'000 BV, the abatement was \geq 80% for N-oxides and even \geq 96% for chlorothiazide. Thereafter, the abatement of the N-oxides started to decrease, but stayed above 80% until 17'000 BV for chlorothiazide. At 23'000 BV, the abatement of N-oxides was \leq 40%, while chlorothiazide abatement was still higher (57%). In $GAC_{loaded/OZO}$, only chlorothiazide was efficiently abated by 30-40% at 50'000 BV (Fig. S23b). In general, more polar compounds are

produced in ozonation, which are expected to be less efficiently sorbed than their parent compounds, therefore exhibiting an earlier breakthrough. For instance, the sorption of venlafaxine N-oxide and tramadol N-oxide on GAC_{fresh/OZO} dropped below 80% at 8'000 BV, while for the parent compounds venlafaxine and tramadol, this occurred only after 14'000 BV (Fig. 6). The relative abatement of chlorothiazide was also smaller than for hydrochlorothiazide, but only at higher BVs. However, the abatement of fexofenadine was similar to fexofenadine N-oxide (the biggest molecule of the three Noxides, mostly uncharged), where the difference in polarity between the parent and the OTP is probably less pronounced. OTPs without available standards also showed mostly no significant changes of normalized peak areas in SF, MB and FB (Fig. S22). However, significant peak reductions were observed in GAC_{fresh/OZO} (at 14'000 BV) and rarely in GAC_{loaded/OZO} (at 37'000 BV), which may be explained by the respective residual sorption capacity. Transformation products of carbamazepine, BQM and BaQD, showed a particular behavior in SF and GAC_{loaded/OZO}: The BQM peak area was reduced concomitantly to a BaQD peak increase. The other biological systems, MB and FB, showed no change. These results are in agreement with previous observations (Hübner et al. 2014), where BQM formation was found as the major OTP of carbamazepine, and a successive conversion of BQM to BaQM (not detected in this

study) and of BaQM to BaQD was observed in the subsequent biological post-treatments.

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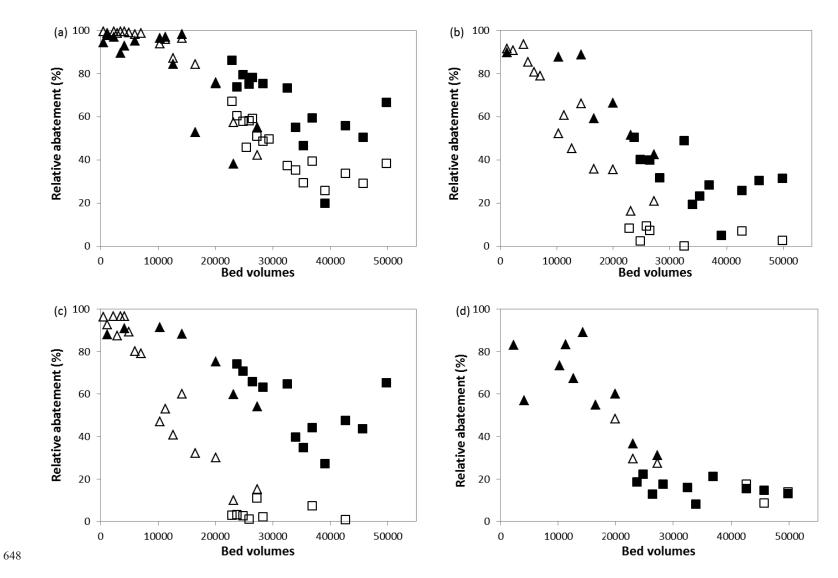


Fig. 6. Comparison of relative abatement of parent compounds (solid) and their respective primary OTP (empty) in $GAC_{fresh/OZO}$ (triangle) and $GAC_{loaded/OZO}$ (square) for (a) hydrochlorothiazide/chlorothiazide, (b) tramadol/tramadol N-oxide, (c) venlafaxine/venlafaxine N-oxide, and (d) fexofenadine/fexofenadine N-oxide. Tramadol*: sum of tramadol and N-oxide (see Text S2).

Cost of new ozone treatment

The cost estimation of the installation and operation of ozonation at WWTP Neugut is presented in detail in SI Text S6. To summarize, investments accounted to 3.27 Mio CHF, and operation costs are 240 CHF/d or 0.013 CHF/m³ (180 CHF/d for ozone and electricity and 60 CHF/d for manpower). Overall, the total cost accounts to 0.06 CHF/m³, corresponding to an increase of 10% for wastewater treatment at WWTP Neugut, what is at the lower end of general estimations (Abegglen and Siegrist 2012). According to the new Swiss Water Protection Act, the investment costs will be covered to 75% by a fund established from payments of 9 CHF/y by each Swiss inhabitant who is connected to a WWTP without installation of advanced treatment (Eggen et al. 2014).

CONCLUSIONS

- Ozonation of the effluent of the secondary clarifier in WWTP Neugut resulted in a significant reduction of the micropollutant load in the WWTP effluent. At the recommended ozone dose (0.55 g O₃/g DOC), indicator substances were abated by >80% even at potential high nitrite concentrations of 0.2 mg NO₂⁻-N/L. For a set of about 200 micropollutants detected in the influent of the WWTP, an average abatement of >79% was observed over the whole treatment chain. Nitrite could be compensated directly by using a feed-back control for ozone dosing by the delta UV_{254nm} signal from in- and outlet of the ozone reactor (Wittmer et al. 2015). In this case, a specific ozone dose of 0.40 g O₃/g DOC (1.5-2.5 mg O₃/L) would be sufficient for the required micropollutant abatement of 80%.
- For the implementation of ozonation, the formation of the oxidation by-products bromate and NDMA needed to be evaluated and were found to be formed also at WWTP Neugut. NDMA was significantly reduced in all post-treatments. As bromide was mostly in a concentration range common for Swiss municipal wastewater, the bromate level was typically < 3.5 ug/L in the WWTP effluent. The concentrations expected after dilution of discharged water were clearly below (potential) drinking water standards (10 μg/L for bromate, 10 ng/L for NDMA).
- While the micropollutants were not significantly abated in the biological post-treatment (SF, MB, FB) as expected, GAC filtration was efficient to remove the majority of micropollutants due to sorption, even after 30'000 bed volumes.
- GAC_{fresh/OZO} (operated with wastewater treated biologically and subsequently by ozonation)
 showed a later breakthrough of DOC and micropollutants than GAC_{fresh/BIO} (operated with
 biologically treated wastewater), potentially due to altered organic matter, being less
 competitive for sorption sites after ozonation.
- GAC_{loaded/OZO} was efficient to abate certain substances even up to 50'000 bed volumes (about 2 years of running time). It cannot be assessed so far whether this is due to a residual sorption capacity of the activated carbon or due to biodegradation. GAC filters operated at high bed volumes should be further investigated as they represent a promising alternative to sand filters for an improved quality of WWTP effluents.

- The investigated OTPs were not abated in the biological post-treatments, except one OTP of carbamazepine, which was degraded to another OTP. The detected OTPs appeared to be very stable compounds, and biodegradable compounds seem to be more difficult to be detected. Only the GAC filters were showing abatement of OTPs due to additional sorption capacities, even at 30'000 bed volumes. However, as demonstrated by the abatement of AOC in the biological post-treatments, degradable compounds are being removed, complying with the task of the post-treatments. Overall, the main goal of the biological post-treatment is the elimination of possible negative ecotoxicological effects generated during ozonation.
- For the evaluation of ozonation and the post-treatments, the water quality needs to be assessed with bioassays. Therefore, various ecotoxicological endpoints, using living cells, organisms and communities, were investigated. The outcomes of these tests will be presented in a follow-up study (Kienle et al. in preparation).

Acknowledgements

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Supporting information

- 714 Additional data associated with this article can be found in the Supplementary Material (Text S1-S5,
- 715 Fig. S1-S24 and Table S1-S11).

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