1 **Low Fe(II) concentrations catalyze the dissolution of various Fe(III) (hydr)oxide minerals in the**

2 **presence of diverse ligands and over a broad pH range**

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Abstract

Dissolution of Fe(III) (hydr)oxide minerals by siderophores (i.e. Fe-specific, biogenic ligands) is an important step in Fe acquisition in environments where Fe availability is low. The observed co-exudation of reductants and ligands has raised the question of how redox reactions might affect ligand-controlled (hydr)oxide dissolution and Fe acquisition. We examined this effect in batch dissolution experiments using two structurally distinct ligands (desferrioxamine B (DFOB) and N,N′-di(2-hydroxybenzyl)ethylene-diamine-N,N′-diacetic acid (HBED)) and four Fe(III) (hydr)oxide minerals (lepidocrocite, 2-line ferrihydrite, goethite and hematite) over an environmentally-relevant pH range (4 - 8.5). The experiments were conducted under anaerobic conditions with varying concentrations of (adsorbed) Fe(II) as the reductant. We observed a catalytic effect of Fe(II) on ligand-controlled dissolution even at sub-micromolar Fe(II) concentrations with up to a 13-fold increase in dissolution rate. The effect was larger for HBED than for DFOB. It was observed for all four Fe(III) (hydr)oxide minerals, but it was most pronounced for goethite in the presence of HBED. It was observed over the entire pH range with the largest effect at pH 7 and 8.5, where Fe deficiency typically occurs. The occurrence of this catalytic effect over a range of environmentally relevant conditions and at very low Fe(II) concentrations suggests that redox-catalysed, ligand-controlled dissolution may be significant in biological Fe acquisition and in redox transition zones.

Introduction

Dissolution of Fe(III) (hydr)oxide minerals is an important process in the natural Fe cycle and for biological Fe acquisition. Despite its crustal abundance, the bioavailability of Fe is often insufficient to meet the requirements of organisms due to the poor solubility and slow dissolution kinetics of Fe(III) (hydr)oxide minerals at circum-neutral pH[1]. This exerts a selective pressure favouring organisms that have evolved highly efficient Fe acquisition strategies. Secretion of protons, reductants and ligands allows proton-promoted, reductive, and ligand-controlled dissolution, all of which involve surface-controlled dissolution mechanisms [2]. Proton-promoted Fe(III) (hydr)oxide dissolution is very slow in the neutral to alkaline pH range[3]; the required acidic conditions are difficult to generate in well-buffered (e.g. carbonatic) systems. In reductive Fe(III) (hydr)oxide dissolution, structural Fe(III) is reduced to Fe(II) at the mineral surface. Because Fe(II) is kinetically more labile than Fe(III), detachment is facilitated, leading to increased dissolution rates. The greater solubility of Fe(II) minerals also allows for higher concentrations of dissolved Fe in equilibrium with the solid. However, in the presence of oxygen and particularly at circum-neutral pH, reduced Fe(II) is quickly re-oxidized to Fe(III) by a surface-catalyzed process.[4, 5] This fast re-oxidation inhibits the mobilization of Fe, unless ligands are present that either stabilize the Fe(II) redox state [6] or form Fe(III) complexes in solution. Ligand-controlled Fe(III) (hydr)oxide dissolution involves the formation of ligand-surface complexes that weaken the bond between the Fe(III) participating in the surface complex and the mineral matrix, facilitating detachment. [1, 7-12] Both biogenic ligands (e.g. desferrioxamine B (DFOB)) and synthetic ligands (e.g. *N,N*′-di(2-hydroxybenzyl)ethylene-diamine-*N,N*′-diacetic acid

61 (HBED)) can enhance Fe bioavailability [13, 14]. However, ligand-controlled dissolution is slow

62 compared to reductive dissolution.[15,16]

63 Synergistic Fe mobilization by ligands in the presence of reductants or Fe(II) at low pH was

64 shown in the 1980s, falsifying previous assumptions regarding the additivity of reductive and

65 ligand-controlled dissolution rates.[17] Observation of co-exudation of reductants and ligands by

66 microorganisms and plants [18, 19] suggested that synergistic effects of reductants and ligands

67 might also occur in the neutral and alkaline pH range. Such effects were recently demonstrated

68 at circumneutral pH in oxic and anoxic goethite suspensions[20], as well is in oxic soil

69 suspensions[21].

70 Banwart et al. suggested a mechanism for the catalysis of ligand-controlled dissolution where

71 adsorbed Fe(II)-ligand complexes served as reductant and accelerated the rate-determining

72 detachment of Fe from the surface [17, 22]. Specifically, they proposed that adsorption leads to the

73 formation of ternary surface complexes where the bridging ligand (in their case oxalate)

74 transfers an electron from the adsorbed Fe(II) to a structural Fe(III) atom at the mineral surface

75 [17, 22]. Subsequently, desorption of the adsorbed Fe(III)-ligand surface complex and rate-

76 determining detachment of Fe(II) from the surface contribute to the net dissolution reaction.

77 This mechanism does not involve electron transfer through the mineral structure. An

78 alternative mechanism has been proposed on the basis of previous observations that

79 adsorption of Fe(II) to Fe(III) (hydr)oxide surfaces induces electron transfer through the mineral

80 [23-27]. If this electron transfer destabilizes the coordination environment of Fe at the mineral

4

81 surface, the rate of ligand-controlled dissolution could be enhanced as has been suggested for

82 observations of synergistic effects of ligands and reductants on goethite dissolution[20].

83 The aim of this study was to explore the potential catalytic effect of Fe(II) in ligand-controlled

84 Fe(III) (hydr)oxide dissolution for Fe acquisition strategies under a variety of environmental

85 conditions. In this context, significant knowledge gaps exist regarding the specificity of the

86 effect with respect to mineral structure, ligand structure and pH. Furthermore, the

87 development of rate laws for the quantification of these effects is lacking. To close these gaps

88 and to provide the groundwork for further explorations of the underlying mechanisms, we

89 conducted a series of dissolution experiments under anoxic conditions in which the added Fe(II)

90 concentration, the ligand, the mineral and the pH were varied. Our results demonstrate the

91 broad relevance of this effect beyond specific mineral structures, ligand structures and in a

92 broad pH range. Our observation of significant catalytic effects even at very low transient Fe(II)

93 concentrations suggest that this mechanism may be important in environments where nutrient

94 limitation occurs, including oxic environments at circumneutral pH where Fe(II) produced by

95 thermal or photochemical redox reactions is rapidly reoxidized.

96

97 **Materials and Methods**

98 Five Fe(III) (hydr)oxide minerals were used: goethite-1 (α-FeOOH, specific surface area (SSA):

99 105 $m^2\,g^{-1}$), goethite-2 (α-FeOOH, SSA: 32 $m^2\,g^{-1}$), lepidocrocite (γ-FeOOH, SSA: 70 $m^2\,g^{-1}$),

100 hematite (α-Fe$_2$O$_3$, SSA: 22 $m^2\,g^{-1}$) and 2-line ferrihydrite. Goethite-1 was synthesized according

5

to Hiemstra et al.[28], hematite was synthesized by a sol gel method according to Sugimoto et al.[29], and the other minerals were synthesized according to the standard synthesis methods in Schwertmann and Cornell (2008)[30], as described in detail in Mørup et al.[31] and Schwertmann et al.[32], respectively. The purity and crystal structure of each Fe(III) (hydr)oxide were confirmed by X-ray powder diffraction (XRD, Huber 5042 four-circle diffractometer) (Figure S7). Mineral transformations during our experiments have not been assessed. Based on previous studies with such low examined Fe(II) concentrations, mineral transformations of Fe(III) (hydr)oxide minerals are unlikely to occur on our experimental time-scales[33, 34]. Specific surface areas were determined by a multipoint N_2-BET adsorption method (Quantachrome Nava 2000). We assume that the BET surface area is an appropriate predictor for the ligand-accessible surface area except for 2-line ferrihydrite where this assumption fails [35, 36].

Ligands and pH buffers were obtained from commercial sources and used as received. DFOB (a trishydroxamate) and HBED (containing amine, phenol and carboxyl groups) have been used as ligands. These ligands have a high affinity for Fe(III) ($log\beta_{11}$ = 33.0 and 39.0 for DFOB and HBED, respectively (I=0)). The examined ligands have been investigated intensively regarding their structure, solution speciation and reactivity [20]. Further information on the chemicals is available in the Supporting Information (Table S1). Ultrapure water (resistivity > 18.2 MΩ·cm, TOC < 2 ppb, Milli-Q, Millipore) was used to prepare all solutions and suspensions.

The pH of solutions and suspensions was monitored throughout the experiments with a pH meter (Orion 3 star, Thermo). Total dissolved Fe concentrations were measured by inductively coupled plasma mass spectrometry (ICP-MS, Agilent-7700; limit of quantification: 0.8 µg L^{-1} Fe

122 (0.014 µM)). The total DFOB and HBED solution concentrations were determined by measuring

123 light absorbance of the corresponding Fe(III) complexes using a UV-vis spectrophotometer

124 (Varian Cary 50) after addition of an Fe(III) solution to ensure complete complex formation.

125 Absorbance was measured at the respective absorbance maxima of Fe(III)-DFOB (428 nm; ε =

126 ~2820 M^{-1} cm^{-1}) and Fe(III)-HBED (493.1 nm; ε =~3440 M^{-1} cm^{-1}). Redox speciation of the

127 complexed Fe in solution was not examined.

128

129 *Batch dissolution experiments*

130 The influence of the Fe(II) concentration on Fe(II) catalysed ligand-controlled dissolution rates

131 was examined in lepidocrocite suspensions (0.1 g L^{-1}) by varying the Fe(II) concentration from 0

132 to 5 µM and subsequently adding a fixed ligand concentration. Under oxic conditions or in

133 redox transition zones where Fe(III) (hydr)oxide minerals may be found, Fe(II) concentrations

134 are typically low and often transient. Therefore we explore a low Fe(II) concentration range.

135 The effect of pH on the catalytic effect of Fe(II) was examined by fixing the pH of lepidocrocite

136 suspensions (0.1 g L^{-1}) with buffers at pH values of 4, 6, 7 and 8.5 and applying treatments

137 containing 5 µM Fe(II), 20 µM DFOB or both. The order of addition of Fe(II) and DFOB in

138 combined treatments was varied (either Fe(II) and DFOB added at the same time, or Fe(II)

139 added 20 minutes before DFOB, where the DFOB addition was considered t=0) in order to test

140 the effect of adsorption equilibration on dissolution rates. To assess whether Fe(II) was initially

141 complexed by DFOB in treatments with simultaneous Fe(II) and DFOB application, the aqueous

142 equilibrium speciation of 5 µM Fe(II) in the presence of 20 µM DFOB (in absence of surfaces)

143 was modelled as a function of pH with PhreeqC using the Minteq v4 database[37] supplemented

144 with protonation and complexation constants for DFOB presented in Kim et al[38]. The effect of

145 the type of Fe(III) (hydr)oxide mineral on the catalytic effect of Fe(II) was examined in 0.1 g L^{-1}

146 suspensions of lepidocrocite, goethite-1, goethite-2, hematite and 2-line ferrihydrite, to which a

147 fixed ligand concentration or Fe(II) or both (adding Fe(II) first) were added. 1 µM Fe(II) was

148 added in HBED treatments at pH 6 and 2 µM Fe(II) in DFOB treatments at pH 7.

149

150 Dissolution experiments were carried out in an anaerobic chamber (mBRAUN, unilab 7185)

151 under a nitrogen atmosphere at constant temperature (20 ± 1 °C). The O_2 concentration was

152 monitored and controlled to less than 1 ppm. All water used had been pre-boiled for more than

153 1 h and was purged with N_2 while cooling down before introduction into the anaerobic

154 chamber. For introduction of (N_2-purged) liquids and solid materials into the chamber, the

155 atmosphere in the antechamber was exchanged with N_2 gas at least 5 times. The batch reactors

156 (LDPE, 100 ml) were wrapped in aluminium foil to prevent potential photo-reduction. All Fe(III)

157 (hydr)oxide suspensions were thoroughly mixed with a magnetic stirrer and a Teflon-coated

158 stirring bar. NaCl was applied as background electrolyte to a final concentration of 0.01 M. The

159 effect of the electrolyte and ionic strength was not further explored in this work. The pH was

160 buffered with 0.005 M of either PIPPS (pH 4 and pH 8.5), MES (pH 6), or MOPS (pH 7). Batch

161 dissolution and adsorption experiments were conducted at the same buffer concentration so

162 that potential effects from buffers on ligand and Fe(II) adsorption[39] could be accounted for in

163 parameterizing rate law equations of surface-controlled dissolution reactions. The pH was set

164 by adding HCl or NaOH, and remained constant at the set pH values (ΔpH = ±0.05) throughout

the experiments. The Fe(III) (hydr)oxide suspensions containing buffer and electrolyte were equilibrated overnight in the anaerobic chamber prior to addition of Fe(II) and ligand at 90 % of final suspension volume. The final volume (and suspension concentration) was reached after the addition of Fe(II) and ligand stock solutions as described below. Experimental solutions were prepared under anaerobic conditions. Except for HBED, all chemicals readily dissolved in deoxygenated ultrapure water. HBED was dissolved by adding 0.1 M of NaOH solution to prepare the 700 µM stock solution. The stock solutions were prepared at a pH close to the experimental pH. Fe(II) was added to the pre-equilibrated solutions 20 minutes prior to the ligand. t=0 corresponds with the moment of ligand addition. Samples were collected periodically for a period of 24 h, were filtered through 0.1 μm PVDF syringe filters (Merck, Millipore, catalogue no. SLVV033RS), and were acidified with trace metal grade HNO_3 for analysis of the dissolved Fe concentration by ICP-MS. Dissolution experiments were carried out in duplicates. Mineral dissolution did not exceed 2% of the bulk in any experiment.

Dissolution rates were calculated from the slopes of linear regression lines of the dissolved Fe concentration over time for the data points over which the increase of dissolved Fe(III) was linear; this interval was from 0.5 to 2.0 h for lepidocrocite at pH 6 but longer intervals were used under other conditions (Table S2). The rates are usually normalized to mineral surface area except for 2-line ferrihydrite dissolution rates which are presented as mass normalized rates because ligand-accessible specific surface area cannot be determined by BET analysis.

184   For ligand-controlled dissolution far from the equilibrium solubility of the mineral, steady-state

185   dissolution can be expressed in a rate law that relates the dissolution rate to the adsorbed

186   ligand concentration[40]:

187                $R_{net} = k \, [L]_{ads}$ (1)

188   where R_{net} (pmol s^{-1} m^{-2}) is the net dissolution rate, k (s^{-1}) is a pseudo-first-order dissolution rate

189   coefficient and $[L]_{ads}$ (μmol m^{-2}) is the adsorbed ligand concentration.

190   Assuming that the presence of adsorbed Fe(II) catalyzes ligand-controlled dissolution, we

191   hypothesize that the effect on the rate coefficient can be expressed as:

192                $k = k_{L,} + k_{Fe(II)}[Fe(II)]_{ads}$ (2)

193   where k_L is the rate coefficient of ligand-controlled dissolution and $k_{Fe(II)}$ is the rate coefficient

194   associated with the effect of Fe(II) on ligand-controlled dissolution. In the absence of Fe(II), k

195   equals to k_L and in the presence of Fe(II) k equals to $k_{L,} + k_{Fe(II)}[Fe(II)]_{ads}$.The overall rate law

196   becomes:

197        $R_{net} = k_{L,} \, [L]_{ads} + k_{Fe(II)}[Fe(II)]_{ads} \, [L]_{ads} = \ R_L + R_{cat}$ (3)

198   Where R_L is the ligand-controlled dissolution rate and R_{cat} is the rate enhancement due to Fe(II)

199   catalysis. The hypothesized catalytic effect (CE) of Fe(II) would correspond to R_{net}/R_L (i.e., the

200   ratio of the net rate of dissolution in the presence of the ligand and Fe(II), R_{net}, to the rate of

201   dissolution in the presence of the ligand and the absence of Fe(II), R_L). This rate law formulation

202   is comparable to that of Suter et al., 1988[41], who proposed independent (additive) rates of

203 ligand-controlled dissolution and reductive dissolution by an Fe(II)-ligand complex at Fe oxide

204 surfaces.

205

206 *Adsorption experiments*

207 Adsorption isotherms were determined for sorption of Fe(II), Fe(III)DFOB, Fe(III)HBED, DFOB,

208 HBED to Fe(III) (hydr)oxide minerals at pH 6 with 0.01 M NaCl as electrolyte. Additionally, Fe(II),

209 adsorption to lepidocrocite was examined as a function of pH. For the ligands and Fe(III)-ligand

210 complexes adsorption isotherms were determined in the presence and absence of 1 µM Fe(II)

211 to examine the influence of Fe(II) on their adsorption behaviour. Experimental determination of

212 Fe(II) adsorption in the presence of ligands was not feasible due to the fast Fe(III) mobilization

213 by the ligands and the strong interference of the Fe-complexes in the spectrophotometric

214 determination of Fe(II) concentrations.

215 All adsorption experiments were carried out in an anaerobic tent under a N_2-atmosphere (O_2

216 concentration < 1 ppm, Coy Laboratory) at 20 ±1 °C. Reactors (Polypropylene, 15 mL) were

217 completely mixed and pre-equilibrated overnight in an end-over-end shaker. Solids

218 concentrations of 2.5 g L^{-1} at pH 4 and 6, and 0.5 g L^{-1} at pH 7 and 8.5 were used. For Fe(II)

219 adsorption experiments, total Fe(II) concentrations of 0.5, 1, 5, 10, 20, 50, 100 and 250 µM

220 were applied. After a reaction time of 0.5 h, samples were filtered (0.1 μm) and the filtrate was

221 analyzed for the dissolved Fe concentration. To investigate the adsorption of ligands and Fe(III)-

222 ligand complexes in the presence and absence of Fe(II), total ligand or complex concentrations

223   of 1, 2, 5, 10, 20 and 50 μM were added to a suspension containing no Fe(II) or 1 μM Fe(II).

224   Fe(II) was added 20 minutes before Fe(III)-ligand addition. Adsorption of complexes was

225   examined after a reaction time of 0.5 h; the remaining concentration in solution was analysed

226   by UV-vis photospectrometry. Ligand adsorption was examined after only 1 minute reaction

227   time in order to minimize the formation of Fe chelate complexes during the adsorption

228   experiments. Fe-ligand complexes accounted for less than 1.5% of the amount of ligand

229   applied, except in the treatment with HBED and lepidocrocite, in which they accounted for up

230   to 10%. For determining the adsorption isotherms of HBED, Fe(III)HBED adsorption was

231   neglected, as it was shown to be very small under the experimental conditions. After

232   interaction, the samples were filtered (0.1 μm). A subsample was analyzed by

233   photospectrometry for the total Fe concentration mobilized by the ligand. To another

234   subsample Fe(III) was added in a small excess over the ligand concentration to have all ligand

235   form Fe chelates complexes. Excess Fe was allowed to precipitate overnight. The following day

236   the subsample was filtered (0.1 μm) to remove Fe(III) precipitates and the sample was analyzed

237   by UV-vis photospectrometry. All adsorption experiments were carried out in duplicates.

238

239   **Results and Discussion**

240   *Influence of Fe(II) concentration on the rate of ligand-controlled dissolution of lepidocrocite*

241   Results from lepidocrocite dissolution experiments with 20 μM DFOB or 17.6 μM HBED and

242   various concentrations of Fe(II) at pH 6 are presented in Figure 1a and Figure 1c, respectively.

243   In a control treatment with 5 µM Fe(II), but without any ligand, the dissolved Fe concentration,

244    $[Fe]_{diss}$ remained constant at approximately 2.0 µM, which can be attributed to adsorption. In

245   experiments with added ligand but without added Fe(II), $[Fe]_{diss}$ increased linearly over time.

246   With both added ligand and Fe(II), varying patterns in $[Fe]_{diss}$ over time were observed

247   depending on the ligand and the amount of Fe(II) added.

248   With DFOB, $[Fe]_{diss}$ increased almost linearly over the entire course of the experiment (Figure

249   1a). Even with 5 µM added Fe(II), $[Fe]_{diss}$ at 9 h was still less than the expected maximum value.

250   The maximum value would correspond to Fe(III) as 1:1 DFOB complex (20 µM) plus dissolved

251   Fe(II). The dissolved Fe(II) concentration corresponds to the added Fe(II) concentration minus

252   adsorbed Fe(II) concentration. The affinity of DFOB for Fe(II) is limited and thermodynamic

253   calculations indicate that at pH 6 (in absence of reactive surfaces) less than 1% of 5 µM Fe(II)

254   will be complexed in the presence of 20 µM DFOB (Table S3). This implies that the impact of

255   DFOB on Fe(II) speciation under these experimental conditions is marginal. In the treatment

256   with 5 µM Fe(II) and DFOB, the first measurement of $[Fe]_{diss}$ (at t = 0.5 h) corresponded roughly

257   to the concentration of added Fe(II), suggesting the Fe adsorbed before addition of the ligand

258   was rapidly released back into solution, possibly as Fe(III), when the ligand was added at t = 0 h.

259   A similar observation was made in isotopic exchange experiments with a different ligand

260   (EDTA), where [57]Fe added as [57]Fe(II) to lepidocrocite at pH 6.0 was rapidly released back into

261   solution upon addition of EDTA, although release of [56]Fe also occured[42].

262   With the ligand HBED, the maximum expected value of $[Fe]_{diss}$ was observed at the first time

263   point (t = 0.5 h) with 5 µM added Fe(II) and at the second time point (t = 1 h) with 1 µM added

13

264   Fe(II). A plateau in $[Fe]_{diss}$ was also observed at the expected maximum value at longer times

265   (2, 3, and 5 h) for added Fe(II) concentrations of 0.8, 0.6 and 0.4 μM, respectively. Under these

266   conditions, the values of $[Fe]_{diss}$ at t = 0.5 h were greater than the concentration of added Fe(II),

267   suggesting that some rapid dissolution occurred before the first time point. We did not find

268   reports of measured Fe(II)HBED stability constants and could therefore not calculate Fe(II)

269   speciation under the experimental conditions. Dissolution rates (R_{net}) were extracted based on

270   measured $[Fe]_{diss}$ from t = 0.5 to 2 h for added Fe(II) from 0 to 0.6 μM.

271   As shown in Figures 1 b and d, the observed dissolution rates increased linearly with adsorbed

272   Fe(II) concentrations as measured in ligand-free systems (Fig. S1). A catalytic effect of Fe(II) was

273   observed even at sub-micromolar levels, especially in the presence of HBED.   The catalytic

274   effect (R_{net}/R_L) was substantially stronger for HBED than for DFOB. The Fe(II) effect, as derived

275   from the rate coefficients $k_{Fe(II)}$ in the presence of the two ligands, was 7 times larger for HBED

276   ($k_{Fe(II)} = 1.8 \pm 0.6 \times 10^{-2}$ μmol^{-1} m^2 s^{-1}) than for DFOB ($2.6 \pm 0.8 \times 10^{-3}$ μmol^{-1} m^2 s^{-1}).

277   Dissolution rates (reported in Table S2) were calculated on a surface-normalized basis

278   (described in Fig. S4) and rate coefficients were calculated using measured adsorbed ligand

279   concentrations (Fig. S2).   Ligand controlled dissolution in the absence of added Fe(II) was

280   measured and corresponds to the y-intercepts of Figs. 1 b and d (Eqn. 1). With $[L]_{ads}$ of 1.7 $\times$

281   10^{-1} μmol m^{-2} for both DFOB and HBED, the calculated rate coefficient for ligand controlled

282   dissolution (k_L) is 1.7 times greater for HBED (4.2×10^{-4} s^{-1}) than for DFOB (2.5×10^{-4} s^{-1}).

283   In order to calculate the rate coefficients k_L and $k_{Fe(II)}$ from the observed rates, it was necessary

284   to estimate the concentrations of the adsorbed ligands and Fe(II). This estimation assumed

285   that ligand and Fe(II) adsorption were independent from each other. To test this assumption,

286   HBED and DFOB adsorption at pH 6.0 were measured with 0 or 1 µM added Fe(II).   The

287   adsorption isotherms (Fig. S2) showed a small enhancement by Fe(II) in the case of HBED and a

288   small inhibition by Fe(II) for DFOB. These contrasting effects can be attributed to an increase in

289   positive surface charge due to Fe(II) adsorption, leading to increased electrostatic attraction

290   between HBED ligands (dominant solution species at pH 6: H_3HBED^-) and the surface, but to

291   increased repulsion between DFOB ligands (dominant solution species at pH 6: H_4DFOB^+) and

292   the surface. Thus the values calculated for $k_{Fe(II)}$ may be slightly overestimated for HBED (if its

293   adsorbed concentration was higher than the value used in the calculation) and slightly

294   underestimated for DFOB. These effects, however, are expected to be minor in comparison

295   with the effect of the ligand on the magnitude of $k_{Fe(II)}$. In the absence of detailed information

296   on surface complex formation of DFOB and HBED, which is not available at the current stage,

297   we cannot fully interpret the differences in reactivity between these ligands.

298

299   *Influence of pH on Fe(II) catalysed ligand-controlled dissolution of lepidocrocite*

300   In the absence of Fe(II), DFOB-controlled lepidocrocite dissolution rates increased by a factor of

301   2.9 over the pH range between pH 4.0 and 8.5 (Table S4). Borer et al.[43] found a somewhat

302   smaller (1.6 fold) increase of DFOB-controlled lepidocrocite dissolution rates in the pH range

303   from 3 to 8. They found that the increasing dissolution rates were correlated with increasing

304   surface concentrations of DFOB while dissolution rate coefficients for DFOB were not pH-

305   dependent, suggesting that the surface speciation of adsorbed DFOB does not change over the

306   pH range.

307   As shown in Figure 2 and Table S4, the addition of 5 µM Fe(II) had a clear effect on dissolved Fe

308   concentrations over time. After 9 h the total dissolved Fe concentration amounted 14 - 19 µM

309   Fe, depending on the pH. Comparing the results of dissolution experiments in the absence and

310   the presence of Fe(II), it is important to note that elevated Fe concentrations in solution at the

311   first time-point are partly due to the added Fe(II) concentration. For example, little Fe(II)

312   sorption is expected at pH 4 due to electrostatic repulsion between the positively charged

313   lepidocrocite surface and Fe(II) [44, 45]. Furthermore, equilibrium modelling results indicate that in

314   absence of reactive surfaces, the extent to which 5 µM Fe(II) is complexed by 20 µM DFOB

315   increases from less than 1 % at pH 6 to close to 100 % at pH 8.5 (Table S3). Therefore, sorption

316   of Fe(II) and DFOB cannot be considered independent at pH >6. The degree to which Fe(II) is

317   complexed in solution correlates with the difference in initial Fe mobilization (after 5 minutes)

318   between the treatment in which Fe(II) and DFOB were applied simultaneously and the

319   treatment in which Fe(II) was applied 20 minutes before the DFOB; at pH 6 there was no

320   difference (Figure 2b), whereas at pH 8.5 the differences corresponded with the applied 5 µM

321   of Fe(II) (Figure 2d). The order of Fe(II) and ligand addition had only a minor effect on apparent

322   initial dissolution rates (5-30 minutes) and catalytic effects (Table S4). This suggests that after 5

323   min the order of addition did not influence the surface speciation including the precurcer of the

324   rate controlling step.

325 The inhibition of Fe(II) sorption at pH 4 evidently also inhibited the catalytic effect (i.e., CE = 1.1)

326 of Fe(II) on ligand-controlled dissolution. A 70-fold increase in Fe(II)$_{ads}$ surface concentration

327 with increasing pH (measured in the absence of the ligand (Figure S1a-b)) lead to a 21-fold

328 increase in dissolution rate from pH 4 to 7, and a CE of around 7 at pH 7 (Table S4). Between pH

329 7 and 8.5, the dissolution rates decreased by 50 %, possibly in part due to formation of Fe(II)-

330 DFOB complexes in solution, inhibiting Fe(II) sorption as discussed above. Oxygen-independent

331 oxidation of Fe(II) by a metal to ligand charge transfer as observed by Kim et al. 2010[38] may

332 have also contributed to decreasing catalytic effects at elevated pH. Furthermore, the upper pH

333 values (7 and 8.5) are close to the pristine point of zero charge of lepidocrocite; in this pH range

334 changes in particle aggregation state may occur that could affect adsorption and the mineral

335 dissolution rate[46, 47].

336 In the absence of adsorbed Fe(II), DFOB adsorption gradually increases with pH[43], but

337 competitive adsorption of Fe(II) decreases DFOB adsorption (Figure S2a). The observation

338 suggests that the interplay of these two effects leads to a maximum catalytic effect at pH 7.

339

340 *Influence of the Fe(III) (hydr)oxide mineral on Fe(II) catalysed ligand-controlled dissolution*

341 The effect of Fe(II) on the dissolution rates of various Fe(III) (hydr)oxides in the presence of

342 DFOB or HBED is shown in Figure 3. Catalytic effects between 2.1 and 13 were observed for all

343 Fe(III) (hydr)oxides (Figure 3 and Table 1). They were generally larger for HBED than for DFOB

344 (except with 2-line ferrihydrite). Catalytic effects were not consistently correlated with mineral

345 structure. The largest catalytic effect was observed for HBED-controlled goethite-2 dissolution,

346 but in the presence of DFOB catalytic effects were larger for the less crystalline 2-line

347 ferrihydrite and lepidocrocite compared to the more crystalline goethite and hematite. The

348 presented rates (Table 1) are surface normalized. Application of the same total ligand

349 concentration to a mineral with a lower SSA at the same suspension density can induce as shift

350 along the adsorption isotherm potentially leading to a higher surface normalized dissolution

351 rate. However, the rate order $k_{Fe(II)}$ provides a measure, independent of the adsorbed ligand

352 concentration and specific surface area for comparing the catalytic effect among minerals.

353 Therefore, rate coefficients (k_L and $k_{Fe(II)}$) for each Fe(III) (hydr)oxide (except 2-line ferrihydrite)

354 were calculated according to rate equation 3 and 4. $HBED_{ads}$ was determined from Langmuir fits

355 of HBED adsorption in the absence of Fe(II), $Fe(II)_{ads}$ was estimated from Langmuir fits of Fe(II)

356 adsorption in the absence of a ligand. The effect of Fe(II) on HBED adsorption was shown to be

357 small (Figure S2); the effect of HBED on Fe(II) adsorption is not known and may affect $k_{Fe(II)}$

358 calculations. The rate coefficients for ligand controlled dissolution k_{HBED} were of the same order

359 of magnitude for all minerals except for hematite, where k_{HBED} was significantly smaller (Table 1

360 and Figure S6). Much larger variations were found for the rate coefficient $k_{Fe(II)}$ in the presence

361 of HBED, with a maximum rate coefficient for goethite-2 and an almost two orders of

362 magnitude lower rate coefficient for hematite. The large $k_{Fe(II)}$ for goethite-2 in the presence of

363 HBED is consistent with the largest catalytic effect of HBED-controlled goethite-2 dissolution.

364

365 **Environmental Implications of Fe-Catalysed Ligand-Controlled Dissolution**

366 Fe(II) can play a pivotal role in biological Fe acquisition and natural Fe cycling of Fe(III)

367 (hydr)oxide minerals. Our results demonstrate a catalytic effect of Fe(II) on ligand-controlled

368 dissolution of Fe(III) (hydr)oxide minerals that scales linearly with the adsorbed concentration of

369 Fe(II) under the experimental conditions. This effect was observed over the environmentally

370 relevant pH range from 6 to 8.5, for two different ligands (HBED and DFOB) and for four Fe(III)

371 (hydr)oxide minerals (lepidocrocite, 2-line ferrihydrite, hematite and goethite). Even at very low

372 (sub-micromolar) concentrations, Fe(II) can strongly enhance the ligand-controlled dissolution

373 rate of Fe(III) (hydr)oxide minerals. This is of particular relevance in the context of Fe acquisition

374 strategies. Fe deficiency generally occurs under oxic conditions, where Fe(II) has a short

375 residence time as a result of surface catalysed Fe(II) oxidation. Organisms that exude reductants

376 for Fe acquisition may only generate low transient Fe(II) concentrations. Furthermore, we

377 found that the catalytic effect of Fe(II) is not specific to a particular ligand. The ligands used in

378 this investigation, DFOB (a biogenic siderophore) and HBED (a synthetic ligand), differ in charge,

379 ligating groups and affinity for Fe(III). Therefore, it is probable that the observed catalytic effect

380 may occur with a range of naturally occurring ligands. Its size was found to differ between the

381 examined ligands. The catalytic effect of Fe(II) was observed for all examined Fe(III) (hydr)oxide

382 minerals, which commonly occur in the environment, and the effect size depended on the

383 mineral. The fact that the catalytic effect was largest around pH 7 is of particular environmental

384 relevance as this is the pH range where solubility of Fe(III) (hydr)oxide minerals is lowest, Fe

385 deficiency typically occurs and where the need for efficient Fe acquisition strategies is most

386 pronounced. Research in microbial and plant physiology and ecology is needed to explore the

387 significance of this effect for biological iron acquisition in natural systems. Similarly, further

19

geochemical investigations are needed to address open questions with regard to the mechanism of the catalytic effect of Fe(II) on ligand controlled dissolution (Biswakarma et al. 2018).

In general, our results suggest that the catalytic effect of Fe(II) on ligand-controlled dissolution may enhance dissolution rates of Fe(III) (hydr)oxide minerals under a wide range of environmental conditions and we expect that it may have broad relevance in soil and aquatic environments.

Supporting Information Available

The Supporting Information is available free of charge via the Internet at http://pubs.acs.org/ and contains adsorption isotherm data, a description of the procedure for calculating dissolution rates and adsorbed Fe(II) concentrations, an overview of the materials used, a summary of the batch dissolution experiments and dissolution rates, Fe(II) speciation modelling

407 results and an overview of dissolution rates and the catalytic effect of Fe(II) as a function of pH,

408 in 7 Figures and 4 Tables.

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Main figures and table

Figure. 1. Dissolution of lepidocrocite by HBED and DFOB in the presence of various concentrations of Fe(II) and dissolution rates as a function of adsorbed Fe(II) concentration

Figure. 2. Dissolution of lepidocrocite by DFOB in the presence and absence of Fe(II) at various pH conditions

Figure. 3. Dissolution of various types of Fe(III) (hydr) oxide mineral by ligand in the presence and absence of Fe(II)

Table. 1. Comparison of dissolution rates, the size of catalytic effect of Fe(II) and rate coefficients between various types of Fe(III) (hydr) oxide mineral

TOC figure

Figure 1. Fe mobilization from lepidocrocite (0.1 g L^{-1}, 0.01 M NaCl) by (a) 20 μM DFOB and (c) 17.6 μM HBED as a function of time for various applied Fe(II) concentrations at pH 6. Error bars indicate standard deviations. Lepidocrocite dissolution rates for (b) Fe(II) + DFOB and (d) Fe(II) + HBED as a function of the adsorbed Fe(II) concentration (uncorrected for ligand adsorption). Adsorbed Fe(II) concentrations at t=0 were calculated based on adsorption isotherm (SI. Fig. 1 (a), SI. Fig. 5). Dissolution rates were calculated from the slopes of linear regression lines over the time interval t = 0.5 -2 h for the treatments presented in panel (a) and (c). The calculation procedure is described in the SI. Fig.4.
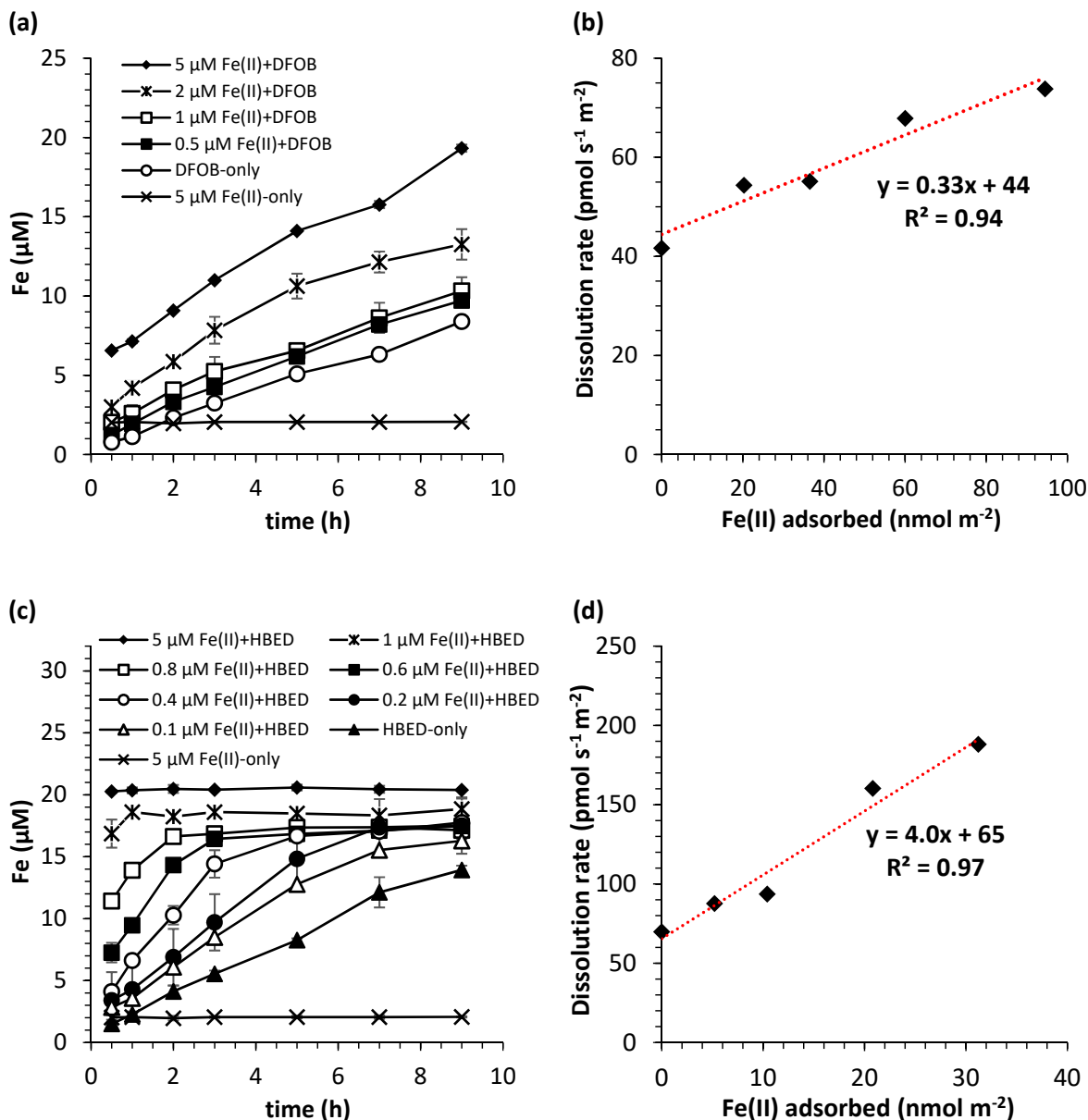
Figure 2. Fe mobilization from lepidocrocite (0.1 g L^{-1}, 0.01 M NaCl) by 20 µM DFOB as a function of time in presence and absence of 5 µM Fe(II) at (a) pH 4, (b) pH 6, (c) pH 7 and (d) pH 8.5. Error bars indicate standard deviations. 5 µM Fe(II) was added in two different ways; Fe(II)(20 min earlier)+DFOB: Fe(II) was added 20 min before the ligand was added, Fe(II)+DFOB: Fe(II) and DFOB were added at the same time by adding pre-mixed Fe(II) and DFOB stock solution.
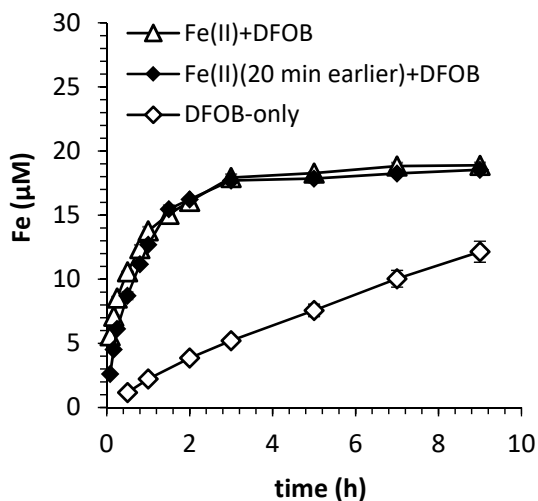
(a) - pH 4
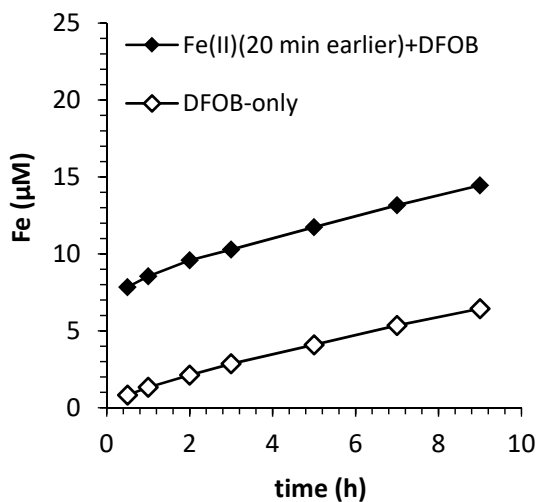
(b) - pH 6
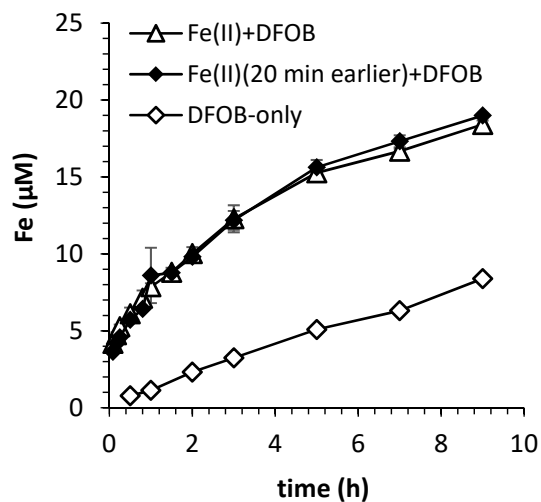
(c) - pH 7

(d) - pH 8.5
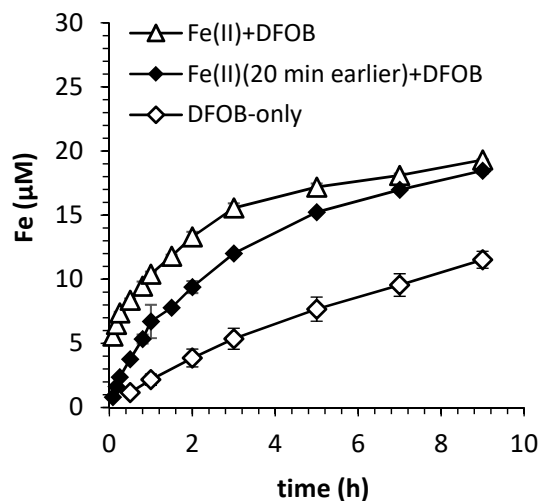
Figure 3. Fe mobilization from (a-b) hematite, (c-d) goethite-2, (e-f) goethite-1, and (g-h) 2-line ferrihydrite and by 20 µM DFOB or 17.6 µM HBED and (i) lepidocrocite (0.1 g L^{-1}, 0.01 M NaCl) by 20 µM DFOB in the presence and absence of Fe(II) (2 µM for DFOB and 1 µM for HBED) as a function of time at a fixed pH (pH 7 for DFOB and pH 6 for HBED). The unit of time on the x-axis is hours (h) for hematite and goethite (a-f) and minutes (min) for 2-line ferrihydrite and lepidocrocite (g-i). Error bars indicate standard deviations.
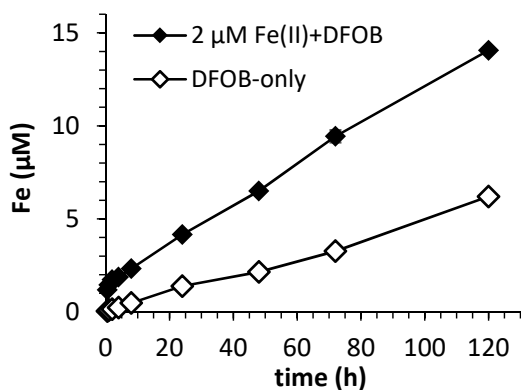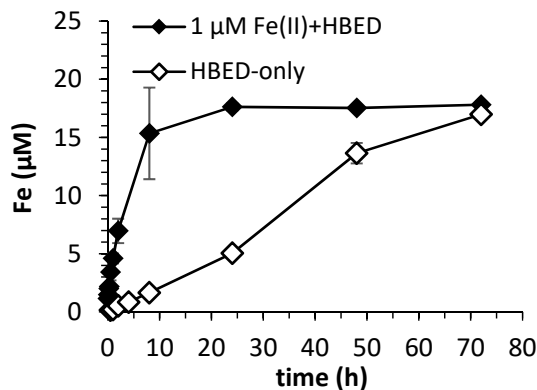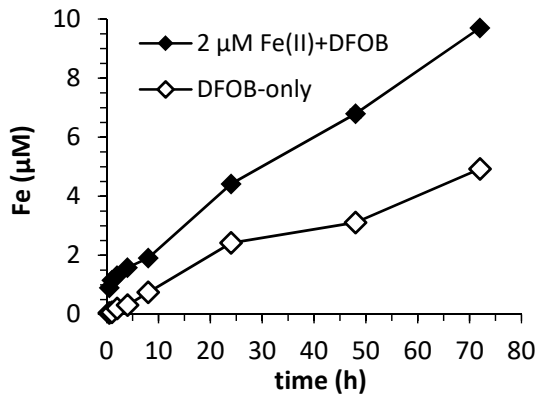

(a) - hematite, DFOB
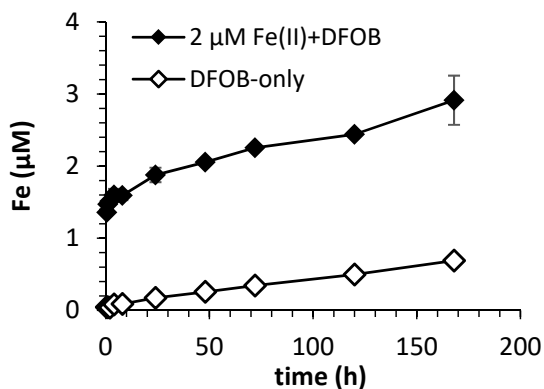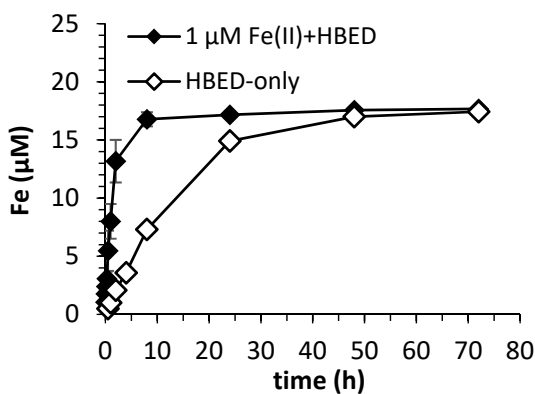
(b) - hematite, HBED
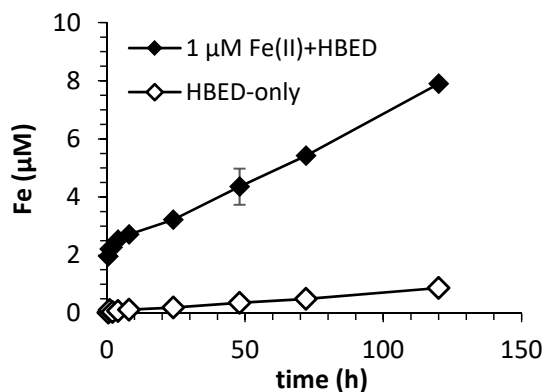
(c) - goethite-2, DFOB

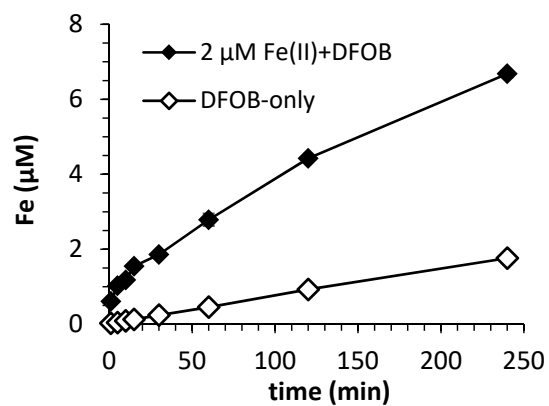**(d) - goethite-2, HBED**

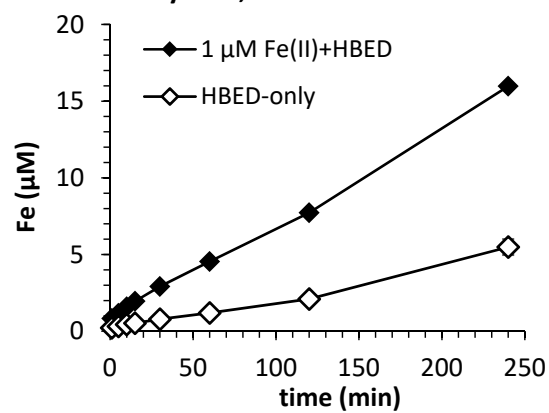**(e) - goethite-1, DFOB**

(f) - goethite-1, HBED

(g)- 2-line ferrihydrite, DFOB

(h) - 2-line ferrihydrite, HBED
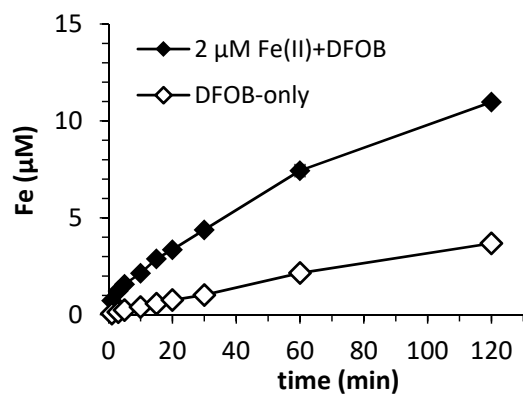

(i) - lepidocrocite, DFOB

Table 1. Dissolution rates, catalytic effect of Fe(II) (C. E. Fe(II)) and rate coefficients for various Fe(III) (hydr) oxide minerals. The catalytic effect of Fe(II) is the ratio between the dissolution rates in the treatment with both Fe(II) and ligands (R_{net}) and the treatment with ligands only (R_L). Rate coefficients of ligand-controlled dissolution (k_L) and Fe(II) catalyzed ligand-controlled dissolution ($k_{(Fe(II))}$) were determined according to rate law equations (3, 4) using the adsorbed Fe(II) ($Fe(II)_{ads}$) and HBED ($HBED_{ads}$) concentrations. No rate coefficients were determined for 2-line ferrihydrite since no surface area normalized net dissolution rates were available.

Fe(III) (hydr) oxide	treatment	R_L (pmol s^{-1} m^{-2})	R_{net} (pmol s^{-1} m^{-2})	C. E. Fe(II)	$Fe(II)_{ad}$ (µmol m^{-2})	$HBED_{ads}$ (µmol m^{-2})	k_{HBED} (s^{-1})	$k_{Fe(II)}$ (HBED) (µmol^{-1} m^2 s^{-1})
hematite	2 µM Fe(II) + DFOB		9.5×10^{-1}	2.1				
	DFOB-only	4.5×10^{-1}						
	1 µM Fe(II) + HBED		5.9	6.9	1.8×10^{-2}	1.2×10^{-1}		2.3×10^{-3}
	HBED -only	8.5×10^{-1}					7.2×10^{-6}	
goethite-2	2 µM Fe(II) + DFOB		9.2	2.1				
	DFOB-only	4.3						
	1 µM Fe(II) + HBED		2.3×10^{2}	13	2.2×10^{-2}	0.9×10^{-1}		1.1×10^{-1}
	HBED -only	18					2.0×10^{-4}	
goethite-1	2 µM Fe(II) + DFOB		3.2	1.8				
	DFOB-only	1.8						
	1 µM Fe(II) + HBED		1.9×10^{2}	7.9	3.3×10^{-2}	0.6×10^{-1}		7.9×10^{-2}
	HBED -only	24					3.8×10^{-4}	
lepidocrocite	2 µM Fe(II) + DFOB		2.9×10^{2}	4				
	DFOB-only	73						
	0.1 µM Fe(II) + HBED		88	1.3	0.5×10^{-2}			
	0.2 µM Fe(II) + HBED		94	1.3	1.0×10^{-2}	1.7×10^{-1}		[a]1.8×10^{-2} (SD: 6.0×10^{-3})
	0.4 µM Fe(II) + HBED		1.6×10^{2}	2.3	2.1×10^{-2}			
	0.6 µM Fe(II) + HBED		1.9×10^{2}	2.7	3.1×10^{-2}			
	HBED -only	70					4.2×10^{-4}	
2-line ferrihydrite	2 µM Fe(II) + DFOB		[b]4.9×10^{3}	4.1				
	DFOB-only	[b]1.2×10^{3}						
	1 µM Fe(II) + HBED		[b]1.0×10^{4}	3				
	HBED -only	[b]3.4×10^{3}						

All experiments have been performed with 0.1 g L^{-1} Fe(III) (hydr)oxide (0.01 M NaCl). The experiments involving DFOB were performed at a ligand concentration of 20 µM at pH 7 and the experiments involving HBED were performed at a ligand concentration of 17.6 µM at pH 6.

[a] Average rate coefficients and standard deviation (SD) based on observations in the presence of 0.1 to 0.6 µM of Fe(II).

[b] The unit of dissolution rates for 2-line ferrihydrite is **pmol s^{-1} g^{-1}** (normalized per mass (g), not per surface area), because the ligand-accessible specific surface area could not be determined by BET analysis for 2-line ferrihydrite.

Graphical Abstract / TOC abstract