

# Assessing emissions from pharmaceutical manufacturing based on temporal high resolution mass spectrometry data

Sabine Anliker<sup>a,b</sup>, Martin Loos<sup>c</sup>, Rahel Comte<sup>a,b,§</sup>, Matthias Ruff<sup>a,#</sup>, Kathrin Fenner<sup>a,b,d</sup>, Heinz Singer<sup>\*,a</sup>

<sup>a</sup> Eawag, Swiss Federal Institute of Aquatic Science and Technology, Dübendorf, Switzerland

<sup>b</sup> Institute of Biogeochemistry and Pollutant Dynamics, ETH Zürich, Zürich, Switzerland

<sup>c</sup> envibee, Dübendorf, Switzerland

<sup>d</sup> Department of Chemistry, University of Zürich, Zürich, Switzerland

<sup>§</sup> Current address: Bachema AG, Analytische Laboratorien, Schlieren, Switzerland

<sup>#</sup> Current address: Gewässer- und Bodenschutzlabor des Kantons Bern, Bern, Switzerland

\* Corresponding author: [heinz.singer@eawag.ch](mailto:heinz.singer@eawag.ch), tel.: +41 58 765 55 77

Word count: 5976

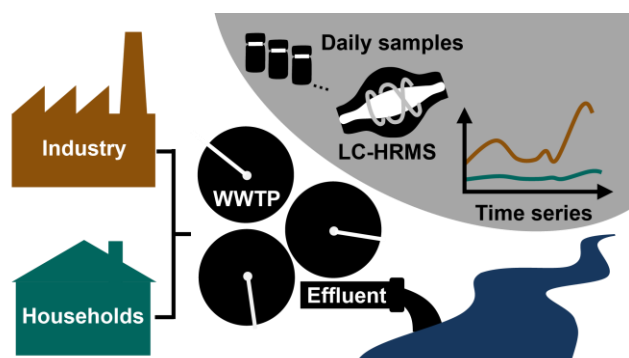
1 small table + 5 small figures: 1800

Total word count: 7776

## Abstract

This study presents a non-target approach to detect discharges from pharmaceutical production in municipal wastewater treatment plant (WWTP) effluents and to estimate their relevance on the total emissions. Daily composite samples were collected for three months at two WWTPs in Switzerland, measured using liquid chromatography high resolution mass spectrometry and time series were generated of all features detected. The extent of intensity variation in the time series was used to differentiate

relatively constant domestic inputs from highly fluctuating industrial emissions. We show that an intensity variation threshold of 10 correctly classifies compounds of known origin and reveals clear differences between the two WWTPs. At the WWTP receiving wastewater from a pharmaceutical manufacturing site: (i) 10 times as many potential industrial emissions were detected as compared to the WWTP receiving purely domestic wastewater; (ii) for 11 pharmaceuticals peak concentrations  $> 10 \mu\text{g/L}$  and up to  $214 \mu\text{g/L}$  were quantified, which is clearly above typical municipal wastewater concentrations; and (iii) a pharmaceutical not authorized in Switzerland was identified. Signatures of potential industrial emissions were even traceable at the downstream Rhine monitoring station at  $> 4000$ -fold dilution. Several of them occurred repeatedly, suggesting that they were linked to regular production, not to accidents. Our results demonstrate that small wastewater volumes from a single industry not only left a clear signature in the effluents of the respective WWTP, but also influenced the water quality of one of Europe's most important river systems. Overall, these findings indicate that pharmaceutical production is a relevant emission source even in highly developed countries with a strong focus on water quality, such as Switzerland.



## Introduction

Effluents from wastewater treatment plants (WWTPs) are an important input pathway of pharmaceuticals and other polar organic compounds to surface waters and contributions from domestic wastewater have been intensely studied.<sup>1-3</sup> However, municipal WWTPs often treat both domestic and industrial wastewater. Wastewater from pharmaceutical companies may contain a particularly large number of biologically active and potentially toxic and persistent compounds.<sup>4-6</sup> Substantial contaminant emissions from pharmaceutical manufacturing have been previously reported,<sup>7-9</sup> but the environmental relevance of these industrial discharges is still poorly understood.<sup>10</sup>

The lack of knowledge regarding the importance of organic contaminant emissions from pharmaceutical manufacturing may be explained by the challenges associated with their investigation. First, batch production predominates in the pharmaceutical industry and production cycles lead to short-term emissions,<sup>8, 11</sup> which are unlikely to be captured by conventional grab sampling or composite sampling conducted during short monitoring periods<sup>12</sup>. Second, manufacturing activities are undisclosed due to production data confidentiality. Thus, substance information is rarely available, and compounds present in production wastewater remain mostly unknown. Under these circumstances target screening approaches are likely to overlook many relevant substances, including intermediates, catalysts and synthesis byproducts.<sup>13</sup> Third, even for known compounds their origin is not easily identified, since compounds detected in municipal WWTP effluents can originate either from domestic or industrial wastewater or both. However, to propose efficient mitigation measures it is crucial to attribute emissions to their sources.

Different strategies have been applied to detect industrial emissions of organic contaminants, albeit mostly in surface water. For example, tracking parent/metabolite ratios revealed significant contributions from pharmaceutical manufacturing discharges to the concentrations of an antiviral drug in the river

Rhine.<sup>14</sup> Several studies detected industrial emissions by spatial sampling, which locally revealed very high concentrations of specific compounds.<sup>15-18</sup> Moreover, by comparing effluent concentrations at different WWTPs, discharges from pharmaceutical industry could be identified based on extraordinarily high concentrations.<sup>19</sup> While most of these approaches focused on a few known compounds, liquid chromatography high resolution mass spectrometry (LC-HRMS) in combination with time series analysis has proven to be of great value to detect industrial discharges of unknown compounds based on temporally fluctuating signal intensities. For example, industrial emissions of previously unknown compounds have been detected in long-term LC-HRMS monitoring data of the river Rhine<sup>20, 21</sup>. More recently, time pattern analysis has been used to identify industrial spills in time series of WWTP influent samples.<sup>22</sup> These studies applied time pattern analysis as a prioritization tool to focus on the structural identification of a few top prioritized compounds, thereby disregarding a large majority of the data. Only a few studies have exploited the wealth of information contained in unidentified HRMS signals. For example, Chiaia-Hernandez, et al. <sup>23</sup> applied non-target time pattern analysis to identify profiles of anthropogenic origin in lake sediments, and Peter, et al. <sup>24</sup> used non-target HRMS signatures to differentiate samples from different sources. However, to the best of our knowledge, so far no study has attempted to assess the contribution of industrial-type time profiles on the total detected emission profiles, nor have long time course data of WWTP effluent samples been systematically investigated for industrial discharges.

The main objectives of this study were to propose and evaluate an approach to (i) attribute unknown substances to their potential emission sources based on their temporal dynamics in daily composite samples of WWTP effluent, and (ii) to estimate the contribution of all likely industrial emissions to the overall emissions of municipal WWTPs to surface waters, without focusing only on the most intense peaks or on identification. To achieve these objectives, we conducted a temporally highly resolved sampling campaign, collecting daily effluent samples for three months at two municipal WWTPs, with one of which

81 is known to treat wastewater from a pharmaceutical production site. Samples were analyzed using a non-  
82 target LC-HRMS method. We then present and evaluated an efficient method suitable for long LC-HRMS  
83 time series to systematically detect fluctuating industrial discharges based on intensity variation and to  
84 distinguish them from more constant domestic emissions. Finally, we assessed the environmental  
85 relevance of the detected industrial discharges by studying their persistence downstream of the WWTPs  
86 in the river Rhine.

## 87 **Material and methods**

### 88 **Chemicals**

89 Details on the preparation of the reference standard solutions and lists of the chemicals used are given in  
90 the Supporting Information (SI) 1-3.

### 91 **Sampling sites**

92 Two Swiss WWTPs were selected as sampling sites. They were chosen such that (i) they are located in the  
93 river Rhine catchment upstream of the international monitoring station (RUES), (ii) one receives input from  
94 pharmaceutical manufacturing and one does not, and (iii) they are similar in terms of treatment processes  
95 and treatment efficiency. WWTP\_dom treats mainly domestic wastewater and serves 146'000 inhabitants.  
96 WWTP\_ind serves 33'000 inhabitants and 3% (by volume, 60% by TOC load) of the wastewater comes from  
97 a pharmaceutical production site. Both plants are equipped with mechanical treatment, a biological  
98 activated sludge treatment including nitrification and denitrification steps and phosphate precipitation.  
99 Detailed information on the WWTPs is in SI 4.

100 In this study, long-term daily LC-HRMS data from the river Rhine measured at and provided by the RUES  
101 close to Basel were also considered and processed with the same method as the WWTP data. The  
102 analytical method used at the RUES is described elsewhere.<sup>17</sup>

### 103 **Sampling**

104 For three months, from November 2014 to January 2015, daily flow-proportional 24-h composite samples  
105 of wastewater effluent were collected at WWTP\_ind and WWTP\_dom (i.e., 87 samples per sampling site).  
106 Sample aliquots (20 mL) were filled in 50 mL amber glass vials and immediately stored at -20°C. Once a

month samples were shipped on ice to the analysis laboratory and again stored at -20°C until chemical analysis.

## **Sample preparation**

Samples were thawed and 5 mL aliquots were centrifuged (Megafuge 1.0 R, Heraeus Sepatech) for 30 min at a relative centrifugal force of 3 g in glass vials. 1 mL of supernatant was transferred to 2 mL measurement vials and 10 µL of a solution containing 135 isotope-labeled internal standards (ISTDs, concentration of each ISTD was 0.5 mg/L, SI 3) was spiked into the samples. Laboratory blank samples were included in the measurement sequence to assess contamination and instrument carry-over.

## **LC-HRMS measurement**

Large volume direct injection LC-HRMS measurement was performed to cover the broadest possible range of substances with one method. An injection volume of 100 µL was used and chromatographic separation was performed on a reversed-phase C18 column (Atlantis T3, 3 µm particle size, 3.0 x 150 mm inner diameter, Waters) applying a water-methanol gradient (both containing 0.1% formic acid). All measurements were performed on a hybrid quadrupole-orbitrap mass spectrometer (QExactive Plus, Thermo Scientific). Full scan spectra at a resolution (R) of 280'000 (at  $m/z$  200) were acquired in positive and negative electrospray ionization (ESI) in separate runs over the mass range of  $m/z$  100-1000, resulting in mass errors generally < 2 ppm. Each full scan was followed by five data-dependent MS2 scans (R=17'500 at  $m/z$  200). MS2 measurements were triggered for the top five most abundant ions, using an exclusion list to prevent recording of MS2 data for the ISTDs and background ions. It should be noted that, using this method, very highly abundant features might have impeded the detection of low abundant co-eluting signals.

For confirmation of the three non-target compounds, samples with the highest intensities were reinjected to acquire MS2 spectra at different collision energies. The full description of the analytical instrumentation and measurement method is in SI 5.

## **Data processing**

### **Time profile building**

The enviMass workflow (version 4.0<sup>25</sup>) was used for the automated detection of intensity time profiles of all features (i.e., chromatographic peaks, defined by  $m/z$  and retention time (RT)) in the LC-HRMS MS1 full scan data. The analysis was based on intensities, as, in contrast to areas, they do not depend on correct peak integration. Mass recalibration and intensity normalization based on ISTDs, as well as RT alignment, were included in the workflow to correct for matrix effects and instrumental drift across measurement sequences. For intensity normalization specifically, intensities of picked peaks in each sample were normalized by the median deviation of all internal standards in the same sample from their individual median intensity across all samples. The enviMass processing steps and parameter settings are provided in SI 6 and SI 7 and the results of the intensity normalization are shown in SI 8.

### **Time profile filtering**

Analysis of the resulting time profiles was performed with the R statistical software v.3.3.3<sup>26</sup>. The function *profiles\_to\_matrix* of the enviMass R package was used to convert time profiles into an R matrix object for further analysis and to aggregate time profiles that originate from the same chemical compound (e.g., related to adducts or isotopes) based on their intensity correlation. Within a set of aggregated features, only the time series with the maximum intensity was further analyzed. Nevertheless, for some compounds more than one time series may be included in the reduced matrix, e.g., compounds that ionized in both positive and negative ESI or artifacts from peak picking (for details, see SI 9). To enhance data quality, the results matrix was filtered using an average sample/blank ratio > 10. Furthermore, only time profiles of



features that eluted after the dead volume of the analytical system ( $RT \geq 3$  min for the WWTP samples and  $\geq 1$  min for the river samples) were considered. Finally, results were restricted to features that were detected on at least three consecutive days. Further information on data reduction and the individual filtering steps is provided in SI 10.

## **Data analysis**

### **Time pattern analysis**

The intensity variations observed in the time series were used to systematically screen for potential industrial emissions and to differentiate them from more constant domestic emissions. In contrast to domestic emissions, industrial discharges were assumed to have intensity variations that exceeded variations in WWTP discharge. Temporal variations were assessed by the fold change in intensities observed in the time series. This “intensity spread” was calculated as the ratio of the 0.95-quantile to the 0.05-quantile of the intensity values using the R *quantile* function. This definition of intensity spread is less sensitive to outliers and therefore more robust than the ratio between maximum and minimum values. Moreover, the applied default algorithm of the R *quantile* function does not rely on any assumption of the underlying distribution of the data, which, particularly in the case of intermittent industrial emissions, is not easily accessible and expected to differ for different substances.

Considering that discharge varied by about a factor of 3 at both WWTPs (see **Figures 2 & 3**), an intensity spread of  $> 10$  was set as the criterion for selecting time profiles that may represent industrial emissions. This assumption is in line with the results of previous studies that have rarely reported temporal variations above a factor of 10 for pharmaceuticals in domestic WWTP effluents.<sup>27-29</sup> Compared to the multivariate empirical Bayes statistic for replicated time course data<sup>30</sup> applied by Alygizakis, et al.<sup>22</sup> to detect industrial emissions, our approach has the advantage that it is suitable for long time series for which replicate measurements would be extremely time consuming. Moreover, no gap-filling is required. Missing values

are common in this data set for various reasons; hence, adequate gap-filling would be very challenging. The intensity spread was calculated for each time series based the available data points, i.e., missing data were treated as NAs (see SI 9 for details on missing data points).

The similarity of temporal patterns from the different sampling locations was investigated using correlation analysis. In this case, for computational reasons, non-detects were set to zero to obtain complete time series. The mass window and RT tolerance for defining time profile pairs from different sampling locations was chosen based on the deviations observed for the ISTDs (see SI 11 for details). The R *ccf* function was used to identify the time shift between the WWTP profiles and the profiles detected downstream in the river Rhine, i.e., the travel time between the WWTPs and the RUES. The time lag that yielded the best correlations for the majority of the profiles was applied to calculate Spearman's rank-order correlation coefficients ( $\rho$ ) between the time profile of the WWTPs and the time-shifted profiles measured in the Rhine using the R *cor.test* function. Time profile pairs with  $\rho \geq 0.8$  were regarded as well-correlated and only profile pairs that met this threshold were considered in further evaluation.

#### **Target quantification**

For quantification five calibration standards, containing 5  $\mu\text{g/L}$  of the ISTD solution and 0.1, 0.5, 1, 5 and 10  $\mu\text{g/L}$  of a mix of 535 target substances, were prepared in ultra-pure water. Quantification of selected compounds was performed with the TraceFinder software 4.1 (Thermo Scientific). Details on the quantification are given in SI 14, Table S7. Concentrations of the non-target compounds that were identified were retrospectively estimated based on a one-point calibration.

#### **Non-target structure elucidation**

Structural identification and confirmation of non-target compounds was performed as described in Ruff et al.<sup>17</sup>. Briefly, molecular formulas were calculated using the Xcalibur formula calculator (Thermo Scientific). Xcalibur Isotope Simulator (Thermo Scientific) was used to simulate isotopic patterns, which

197 were then compared to the measured data. For molecular structures, the ChemSpider database<sup>31</sup> was  
198 queried. MetFrag<sup>32</sup> was used for *in silico* fragmentation of possible structures and to compare the  
199 calculated MS2 spectra with the measured spectra. Furthermore, the number of references found in  
200 ChemSpider was considered to rank most likely candidate structures, which were, if available, compared  
201 to mass spectra found in the two spectral libraries MassBank<sup>33</sup> and Metlin<sup>34</sup>. Finally, for unequivocal  
202 identification authentic reference standards were purchased and RTs and MS2 spectra of sample, spiked  
203 sample and reference standard were compared.

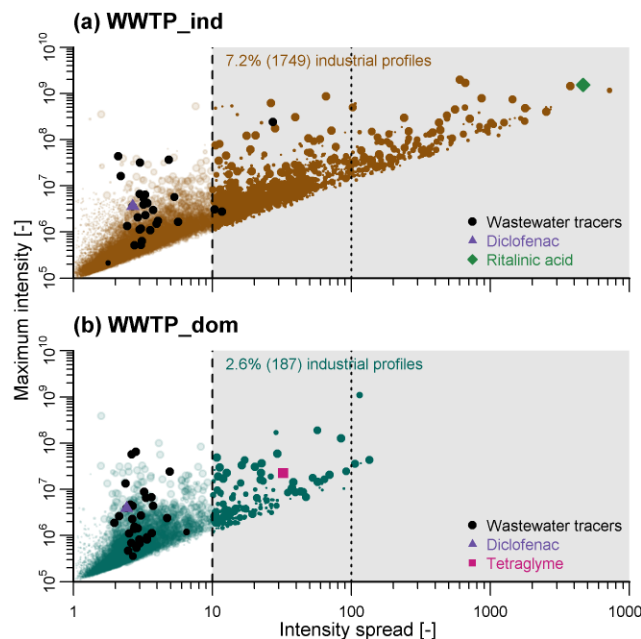
## Results and discussion

### Classification of time profiles

The outcome of data processing was time profiles of intensities for all features detected in the samples, ideally each representing one compound. The time profile inventory (i.e., the full set of time profiles detected in positive and negative ESI) of WWTP\_dom consisted of 7'200 time profiles, whereas more than three times as many profiles, i.e., 24'245, were detected for WWTP\_ind. This result suggests a larger variety of substances was emitted by WWTP\_ind compared to WWTP\_dom.

A primary goal of this study was to assess the contribution of intermittent industrial discharges on the total emission profiles. Therefore, all time profiles were classified into domestic or potential industrial emissions using the intensity spread threshold of  $> 10$ . The result of the classification is illustrated in **Figure 1**. At both WWTPs, a large majority of the profiles (i.e., 92.8% at WWTP\_ind and 97.4% at WWTP\_dom) were attributed to domestic wastewater. The predominance of domestic emission patterns was not surprising, since even in WWTP\_ind households contributed more than 90% of the total wastewater volume. However, the proportion of time profiles classified as potential industrial emissions differed substantially between the two WWTPs and was significantly higher for WWTP\_ind (Pearson's Chi-squared test,  $p < 0.01$ ). For WWTP\_ind, 7.2% (1'749) of the time profiles were classified as potential industrial emissions, compared to 2.6% (187) for WWTP\_dom. While at first, it might be surprising that potential industrial emission profiles were also found at WWTP\_dom, there might still be some small industries in the catchment. However, disproportionately more profiles with very high intensity spreads were observed at WWTP\_ind compared to WWTP\_dom. Specifically, 200 time profiles had an intensity spread greater than 100 at WWTP\_ind, whereas this was only the case for 4 time profiles at WWTP\_dom (profiles right of the dotted lines in **Figure 1**). Moreover, at WWTP\_ind potential industrial time profiles accounted for 96% (compared to 67% at WWTP\_dom) of the profiles of very high intensities (i.e.,  $> 1E8$ ),

suggesting that they were responsible for most peak concentrations. The observed differences between the two WWTPs are likely a consequence of the larger share of industrial wastewater treated by WWTP\_ind and indicate that, despite its relatively small proportion on the total wastewater volume (3%), the industrial inputs are clearly observable in the effluents of WWTP\_ind.



**Figure 1:** Categorization of all detected time profiles at (a) WWTP\_ind and (b) WWTP\_dom. Each bubble represents one time profile. Bubble size represents the number of data points in the respective time profile. Time profiles on the grey background were classified as potential industrial emissions based on their intensity spread > 10. The intensity spread was defined as the ratio of the 0.95-quantile to the 0.05-quantile of the intensity values of a time profile. Please note the logarithmic scale of the axes. Intensity spreads of 10 and 100 are marked with dashed and dotted lines, respectively. The black dots indicate the profiles of selected domestic wastewater compounds (see section “Target and suspect screening”). Examples of domestic (diclofenac) and industrial (ritalinic acid, tetraglyme) time profiles are shown in Figure 3.

We recognize that any continuous industrial discharges are overlooked by this approach, which is further biased towards features with high intensities. Therefore, the estimate of the share of industrial profiles is expected to be rather conservative. On the other hand, temporally fluctuating time profiles might not only be caused by industrial processes but also by storm water run-off from urban areas<sup>35, 36</sup>, irregular use patterns of compounds, e.g., X-ray contrast media or cytostatics in hospital wastewater,<sup>37</sup> or pesticides,

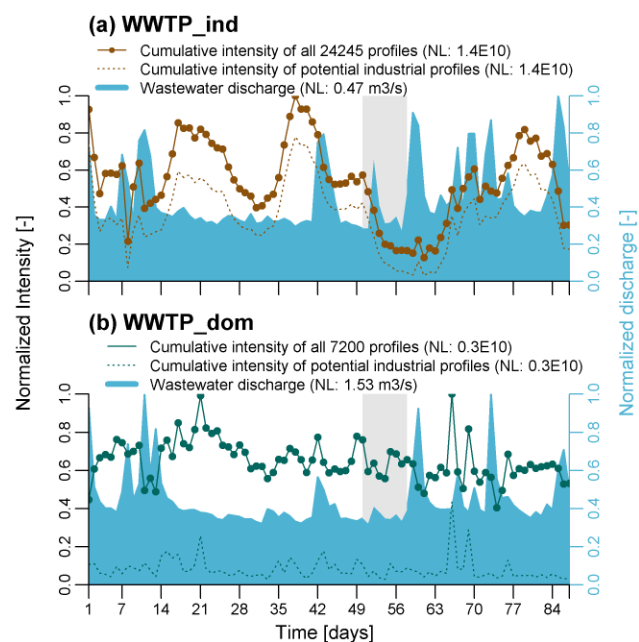
biocides, drugs of abuse<sup>38</sup> and inappropriately disposed medications in domestic wastewater. Moreover, short time peak concentrations of pesticides due to improper handling, washing activities or direct disposal have been reported in WWTP effluent.<sup>35</sup> However, it should be noted that most of these sources only lead to peak emissions of individual compounds.

In the following, we evaluate different lines of evidence to confirm that those profiles classified as potential industrial emissions, are indeed plausibly related to industrial activities.

## **Evaluation of classification**

### **Cumulative profile intensities**

As a first plausibility check of the profile classification, the cumulative profile intensities (i.e., the sum of all intensities across the profiles) were examined and compared between the two WWTPs (**Figure 2**). At WWTP\_ind, a distinct pattern emerged in the cumulative profile intensities, with periods of high intensities alternating with periods of low intensities, which could not be explained by dilution effects resulting from discharge variation. At WWTP\_dom fluctuations in cumulative profile intensities were generally smaller and occurred at a higher frequency. Moreover, on average the potential industrial emission profiles explained 63% of the cumulative profile intensity measured at WWTP\_ind, whereas they accounted only for 12% at WWTP\_dom (dotted lines in **Figure 2**). Interestingly, at WWTP\_ind, the period of lowest cumulative intensities coincided the holiday period between Christmas and New Year, suggesting lower inputs from industrial wastewater due to a production break. Reduced industrial discharges during vacation periods have been previously observed.<sup>20</sup> Overall, these findings provide evidence that production cycle dependent emissions dominate the time profile inventory of WWTP\_ind and thus support the results of the profile classification.



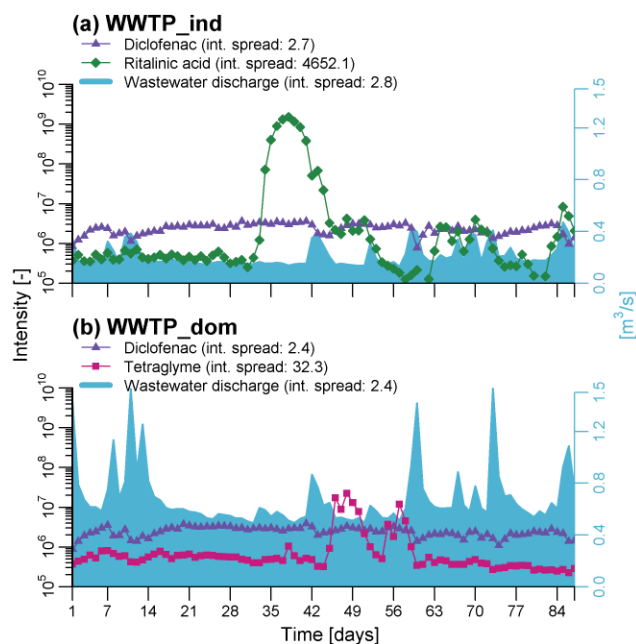
**Figure 2:** Time pattern of the cumulative profile intensities and WWTP discharge in **(a)** WWTP\_ind and **(b)** WWTP\_dom. The cumulative intensities, as obtained by adding up the intensities of all detected features per day, were normalized to the overall maximal value. The respective normalization levels (NL) are indicated in the figure legends. Dashed lines indicate the cumulative intensities of all time profiles classified as potential industrial emissions (see section “Classification of time profiles”). The grey area marks the holiday period between Christmas and New Year.

### Target and suspect screening

To evaluate the time profile classification further, 30 target compounds (26 pharmaceuticals, 2 artificial sweeteners and 2 household chemicals) were investigated that are commonly measured at high concentrations in domestic WWTP effluents due to low removal efficiencies<sup>1, 3, 39</sup>. As expected, based on regular consumption by the population in the catchment, the large majority of these target compounds showed uninterrupted time series with moderate intensity fluctuations. As an example the time patterns of the pharmaceutical diclofenac are shown in **Figure 3**. The time profiles of diclofenac are also marked in **Figure 1**. The time profiles of all investigated domestic wastewater tracer compounds are in SI 12. Within a given WWTP, time profiles of most target compounds displayed very similar time patterns. As a consequence of dilution, minimum values of time profile intensities coincided with periods of maximal

WWTP hydraulic discharge. The variability of the time profile intensities was generally in the same range as the fluctuations of the WWTP discharge (throughout the monitoring period the WWTP discharge varied by a factor of 2.8 and 2.4 at WWTP\_ind and WWTP\_dom, respectively), confirming that discharge dynamics were the main factor influencing time patterns of compounds constantly emitted in domestic wastewater. This finding is in line with previous studies that found a strong association between hydrological parameters and compounds originating from domestic wastewater, e.g., Carpenter, et al.<sup>40</sup>. Our results showed that at WWTP\_dom the time profiles of all domestic wastewater tracer compounds were classified as domestic emissions (marked by the black dots in **Figure 1b**), whereas at WWTP\_ind three pharmaceuticals showed large intensity spreads and were therefore classified as potential industrial emissions (black dots in **Figure 1a**). In the absence of replicate measurements, we concluded from the low variation observed in the time series of the domestic wastewater tracers that also the variability of the measurements must have been low.



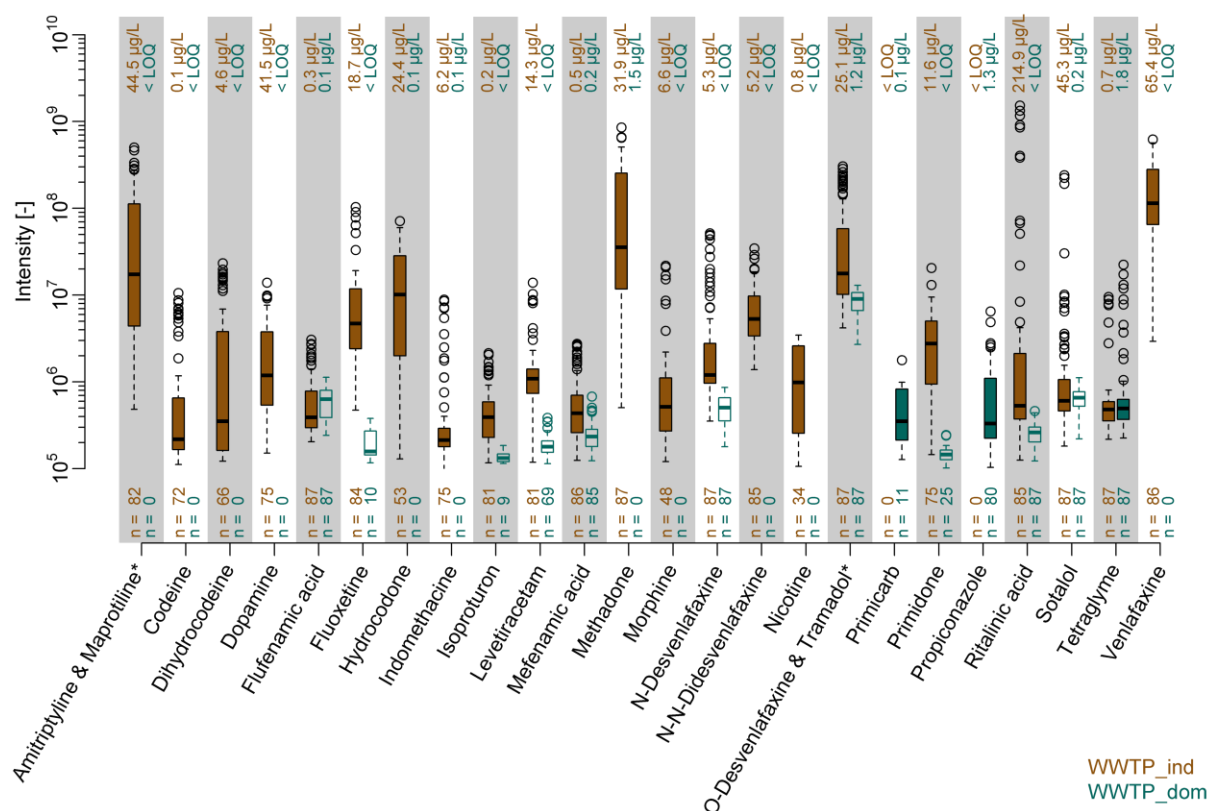


**Figure 3:** Time series of two selected features and WWTP discharge time pattern, in (a) WWTP\_ind and in (b) WWTP\_dom. Diclofenac is shown as an example for a domestic emission pattern. As examples of industrial emission patterns the target compound with the largest intensity spread at the respective WWTP is shown, i.e. ritalinic acid at WWTP\_ind and tetraglyme at WWTP\_dom. The intensity spread was defined as the 95%-quantile/ 5%-quantile ratio of the intensity values of a time profile. Please note the logarithmic scale of the y-axis.

Next, potential industrial emission profiles were screened for matches with one of the 535 spiked reference standards, i.e., target compounds. For WWTP\_ind 22 target compounds and for WWTP\_dom 3 target compounds were detected among the potential industrial emissions (the corresponding time series are given in SI 13). As an example, the time pattern and the intensity distribution of the target compound with the largest intensity spread at WWTP\_ind (ritalinic acid) and WWTP\_dom (tetraglyme) are shown in **Figures 3a** and **3b**, respectively. The time profiles of the latter compounds are also marked in **Figure 1**. The intensity distributions of all target compounds classified as industrial emission are visualized in **Figure 4**. Multiple lines of evidence indicate that most of the emissions of these target compounds originated from an industrial source: (i) their time patterns did not follow WWTP discharge dynamics, in contrast to the time patterns of the domestic wastewater tracer compounds (see SI 12-13 and example profiles in **Figure 3**); (ii) tetraglyme, a compound known to be used in different industrial applications,<sup>15, 41</sup> was

312 correctly classified as industrial emission at both WWTPs; (iii) 20 of 22 target compounds classified as  
313 potential industrial emissions at WWTP\_ind, were pharmaceuticals or pharmaceutical transformation  
314 products (TPs), which is in accordance to the fact that the respective WWTP treated wastewater from a  
315 pharmaceutical production site; and (iv) the classification of the antidepressant venlafaxine and its TPs as  
316 industrial emissions is consistent with the previous observation of point source inputs from venlafaxine  
317 synthesis, namely of the demethylated TP N,N-didesvenlafaxine, in the Swiss section of the river Rhine  
318 downstream of WWTP\_ind.<sup>17</sup> As mentioned, occasional peak emissions of agrochemicals in WWTP effluent  
319 are not uncommon and may lead to false positives of industrial emissions. However, here, due to sampling  
320 in winter outside of the growing season, the observed emissions of isoproturon (at WWTP\_ind) and  
321 pirimicarb and propiconazole (at WWTP\_dom) are more likely related to their use in industrial applications  
322 or as biocides in the urban area.

323 Moreover, time profile inventories were screened for 30 pharmaceuticals found on the production list  
324 (publicly available on the company's website) of the industry connected to WWTP\_ind. Time patterns  
325 classified as industrial emissions were detected for 14 of the pharmaceuticals at WWTP\_ind, with eight  
326 compounds corresponding to target compounds discussed previously and six compounds were detected  
327 based on exact mass screening, i.e., suspects screening (the time profiles of the suspect compounds are  
328 provided in SI 15). All of these compounds were either not detected or categorized as domestic emissions  
329 at WWTP\_dom.

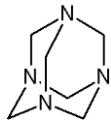
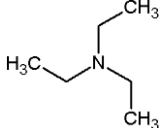
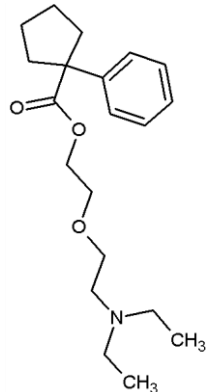


**Figure 4:** Box-whisker plots of the intensity distribution of the quantified target compounds. Maximum concentrations are indicated above each box. Filled boxes indicate time profiles of compounds classified as industrial emissions, whereas empty boxes indicate domestic emissions. If a compound was only detected at one of the two WWTPs only one boxplot appears in the figure, otherwise box-whisker plots for both WWTP\_ind (brown) and WWTP\_dom (blue-green) are shown. The upper and the lower limit of the box indicates the third quartile and first quartile, respectively, the line represents the median, the whiskers extend to 1.5 times the interquartile range from the bottom and the top of the box and any data beyond that range are represented as points. Please note the logarithmic scale of the y-axis. Below each box the number of data points (n) in the time profile is indicated. Stars denote isobaric co-eluting substance pairs that were quantified as the sum of two compounds. Discrepancies between intensity data and quantification results originate from different detection limits of the workflows used for time profile extraction (enviMass) and quantification (TraceFinder).

## Non-target identification

After evaluating the profile classification based on target and suspect compounds, we were interested if also non-target compounds support the classification. Identification efforts were focused on features classified as potential industrial emissions that displayed high signal intensities in the effluent of WWTP\_ind and for which clear candidates were found in the structure elucidation workflow. Finally, the structure of three non-target compounds could be elucidated (**Table 1**). All three compounds were confirmed with an authentic reference standard, i.e., level 1 confirmation confidence according to Schymanski, et al.<sup>42</sup> The time profiles of the structurally identified compounds as well as information on their confirmation can be found in SI 16. One compound was identified as methenamine. This substance was also detected in the effluent of WWTP\_dom, where it also displayed an industrial emission pattern. Methenamine is a high production volume chemical that is widely used in various industrial processes and everyday products<sup>43</sup>, which may explain its occurrence in the effluents of both WWTPs. The other two identified compounds were only detected in the effluents of WWTP\_ind. Triethylamine is also a high production volume chemical that is frequently used in organic synthesis<sup>44-46</sup> and pentoxyverine is an active pharmaceutical ingredient not registered in Switzerland<sup>47</sup>, thereby strongly suggesting an industrial source. This result is in agreement with previous work from Singer, et al.<sup>48</sup>, which also reported discharges of unauthorized pharmaceuticals into Swiss surface waters via WWTP effluent from manufacturing sites.

**Table 1:** Identified and confirmed non-target compounds

Name (CAS)	Detected <i>m/z</i> [Da]	Adduct	RT [min]	Neutral formula	main HCD fragments (NCE)	Structure
Methenamine (100-97-0)	141.1134	[M+H] <sup>+</sup>	3.3	C <sub>6</sub> H <sub>12</sub> N <sub>4</sub>	112.0870; 85.0761 (60)	
Triethylamine (121-44-8)	102.1277	[M+H] <sup>+</sup>	4.7	C <sub>6</sub> H <sub>15</sub> N	74.0965; 58.0655 (75)	
Pentoxifyverine (77-23-6)	334.2376	[M+H] <sup>+</sup>	15.9	C <sub>20</sub> H <sub>31</sub> O <sub>3</sub> N	100.1120; 145.1012 (50)	

CAS: Chemical Abstract Service number; Da: Dalton; HCD: higher-energy collisional dissociation; NCE: normalized collision energy; RT: retention time

Overall, non-target structure identification along with the examination of time patterns of target and suspect compounds of known origin showed that time series classified as potential industrial emissions indeed mostly originated from industrial sources, mainly from wastewater of the pharmaceutical manufacturing site in the catchment of WWTP\_ind. Therefore, we conclude that our approach to systematically screen LC-HRMS time course data of 24-h composite samples for features with high intensity variations was suitable to detect fluctuating industrial discharges of unknown compounds and to estimate their contribution on the total contaminant emissions via WWTP effluent. The intensity spread threshold > 10 worked well to detect time patterns that represent industrial discharges, but would need to be adapted for other WWTPs with different hydraulics. Here, fluctuating discharges from pharmaceutical batch production were examined; however, as they are mostly linked to cleaning activities,

intermittent emissions are characteristic for other industrial sectors as well<sup>49, 50</sup>. Future studies that utilize this approach should provide further insights into its general applicability.

## **Relevance of industrial emissions**

### **Quantification**

Next, to assess the impacts of industrial emissions, the target compounds classified as potential industrial emissions (see section “Target and suspect screening”) were quantified. Quantification revealed maximal concentrations in the µg/L range for 18 of 25 compounds (**Figure 4**, more details on the quantification results in SI 14, **Table S7**). At WWTP\_dom the highest concentration of 1.8 µg/L was measured for the industrial chemical tetraglyme (corresponding to a load of 0.081 kg/day, considering the WWTP discharge on the respective day). Much higher maximal concentrations and loads were found in the effluent of WWTP\_ind. The highest concentration was measured for ritalinic acid (214.9 µg/L or 2.6 kg/day), but also for the pharmaceuticals venlafaxine (65.4 µg/L or 2.0 kg/day) and sotalol (45.3 µg/L or 1.5 kg/day) very high concentrations were measured in the effluent of WWTP\_ind. It should be noted that concentrations > 10 µg/L are estimates since they exceeded the calibration range. Nevertheless, the concentrations found for potential industrial emissions in the effluent of WWTP\_ind differ substantially from the pharmaceutical concentrations commonly measured in WWTP effluent. Effluent concentrations from regular domestic consumption are typically in the ng/L range for most pharmaceuticals with only a few compounds exceeding concentrations of 1 µg/L.<sup>1, 3</sup> Pharmaceutical concentrations in the µg/L to mg/L range in WWTP effluents that receive wastewater from pharmaceutical factories have previously been reported, e.g., in India<sup>51</sup>, Korea<sup>52</sup> and the U.S.A.<sup>8, 19</sup>. However, in these cases the percentage of industrial wastewater of the total wastewater flow was in most cases much higher than the 3% of WWTP\_ind investigated here.

Estimated concentrations were calculated for the identified non-target compounds. This analysis revealed extremely high concentrations for triethylamine, ranging up to 2500 µg/L (43 kg/day) and maximal

concentrations of methenamine and pentoxyverine of 290 µg/L (3.8 kg/day) and 75 µg/L (1.2 kg/day), respectively. These findings clearly demonstrate that the quantification of only known target compounds misses very relevant emissions. The non-target time profile classification presented here is therefore a valuable addition to target screening methods and can provide a more comprehensive and realistic overview of inputs originating from pharmaceutical manufacturing.

#### **Signal preservation in the downstream river system**

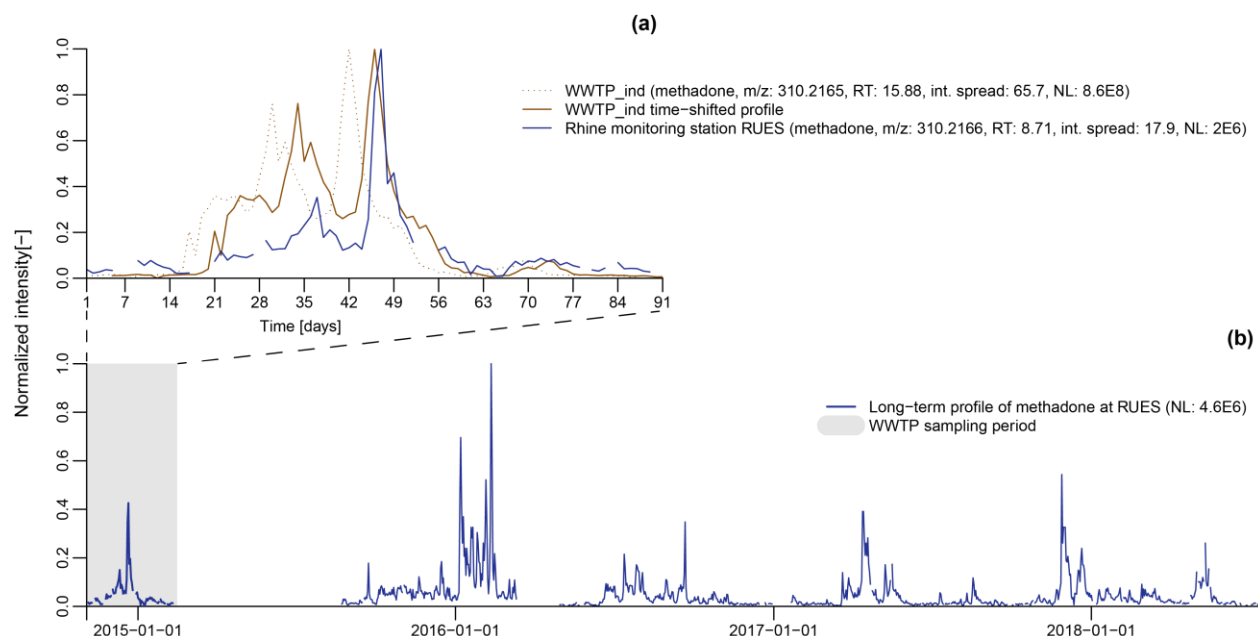
To further assess the significance of our findings with respect to the actual exposure of the aquatic environment, we investigated if the potential industrial emissions could be detected in the downstream river system, at the international Rhine monitoring station (RUES), located more than 100 km downstream of WWTP\_ind and WWTP\_dom. Both WWTPs did not directly discharge the treated wastewater into the river Rhine, but into small streams that eventually enter the Rhine via tributaries. After shifting the WWTP time profiles by 4 days to account for the travel time, 13 emissions from WWTP\_ind were highly correlated (i.e.,  $p \geq 0.8$ ) with time profiles at the RUES, whereas for WWTP\_dom no matching time patterns were found at any time lag tested (i.e., 0 to 14 days). Correlating time profiles between WWTP\_ind and RUES corresponded to the identified non-target compound pentoxyverine, as well as two to target compounds from the product list of the pharmaceutical company connected to WWTP\_ind. One target compound was the opioid drug methadone, whose time profiles at WWTP\_ind and RUES are overlaid in **Figure 5a**. The load of methadone emitted during the sampling period at WWTP\_ind amounted to 12 kg (max. load of 0.9 kg/day). During six weeks methadone concentrations were elevated reaching up to 31.9 µg/L in the WWTP effluent, leading to a calculated maximal concentration in the receiving stream of 3.4 µg/L and 0.01 µg/L in the river Rhine. The plots of all the other correlating time profile pairs are provided in SI 17.

The example of methadone illustrates that, due to the high dilution of the WWTP effluent in the Rhine (average dilution during the sampling period was a factor of 4400 for WWTP\_ind and a factor of 1400 for WWTP\_dom), only signals with very high maximal intensities and clear peak emission patterns were

observable downstream in the catchment. The conservation of emission profiles from one individual WWTP is even more remarkable, given the fact that a total of 540 WWTPs, serving 6.6 million inhabitants, discharge their effluent into the river Rhine upstream of the RUES. Hence, for signal conservation to be observable at the RUES, WWTP\_ind was the dominating, if not the only source of the respective compounds in the catchment. In contrast, none of the time profiles of WWTP\_dom had any influence on the time profiles detected at RUES, despite its larger discharge volume and therefore lower dilution as compared to WWTP\_ind.

Finally, mass traces for the correlated features were extracted from the RUES daily LC-HRMS dataset for another three years after the WWTP sampling period, from July 2015 until July 2018, thereby taking advantage of the strength of HRMS data, which allows for retrospective analysis. For 6 of 13 features, we found clearly recurring phases of high intensity emissions. As an example the time profile of methadone in the long-term monitoring data of RUES is shown in **Figure 5b**; the other profiles are provided in SI 18. The recurring phases of high signal intensities demonstrate that several of the observed industrial emissions were not unique discharge events, but occurred repeatedly for several years. The recurring nature of the emissions indicates that they were not accidental spills, but caused by routine production operations. This result is in accordance with a previous study that also detected recurring emissions from chemical industries<sup>20</sup>.





**Figure 5:** Time profiles of the opioid drug methadone. Time profiles were normalized by dividing all intensities by the maximum intensity of the respective time profile, the respective normalization level (NL) is indicated in the legend. **(a)** Correlating profiles between WWTP\_ind and the international Rhine monitoring station (RUES), located more than 100 km downstream of WWTP\_ind. For WWTP\_ind the measured profile (dotted brown line), as well as the 4-day time-shifted profile (solid brown line) is shown, which overlaps with the time profile measured at the RUES (blue line). **(b)** Recurring phases of high methadone signal intensities observed in the long-term daily LC-HRMS monitoring data of the RUES.

## Environmental implications

The approach presented in this study enables systematic detection of potential industrial emissions of unknown compounds based on temporal variability in 24h-composite samples and distinction of these emissions from domestic emissions. Monitoring at the origin, ideally with high temporal resolution for extended time periods (i.e., several months), is critical to identify contamination hotspots, especially on vulnerable small streams, and to assess the relevance of different sources on overall emissions. Attributing emissions to their source is necessary for the implementation of efficient mitigation measures and to transfer the arising costs to the polluter, according to the polluter pays principle<sup>53</sup>.

By investigating and comparing the effluents of two municipal WWTPs, this study showed that the industrial wastewater left a very clear signature in the effluents of the WWTP that treats wastewater from a pharmaceutical manufacturing site. The large number of potential industrial emissions indicated that a wide variety of compounds was emitted into the industrial wastewater and for the majority of emissions the underlying chemicals remained unknown. Quantification of a few detected target compounds - including highly potent substances, such as opioids and antidepressants - revealed much higher concentrations and larger loads of active pharmaceutical ingredients emitted in industrial wastewater compared to domestic wastewater. Fluctuating industrial discharges caused short-term high peak concentrations, which might be relevant in the receiving waters from an ecotoxicological point of view, especially in small receiving waters with a low dilution of the wastewater effluent. Moreover, applying target and suspect screening as well as non-target identification, the present study showed that industrial emissions contained compounds that are usually not found in domestic wastewater, i.e., compounds related to chemical synthesis or finished products not authorized in Switzerland. Thus, industrial discharges expand the spectrum of compounds released to the aquatic environment compared to emissions from domestic wastewater.

The persistence of industrial emission profiles downstream in the river Rhine, which is one of the most important rivers in Europe, demonstrated that substance inputs from one source can have an effect, even in a large catchment area. In the present study, for some compounds, a few hundred m<sup>3</sup>/d of industrial wastewater were the dominating source in the catchment. These findings indicate that a single company can substantially influence the water quality and the drinking water resource of several million people. Reducing the emissions at the respective point sources seems to be an efficient and necessary way to mitigate water pollution, e.g., by improving the on-site pretreatment of the industrial wastewater. Moreover, minimizing the impacts on water quality is in accordance with and puts into practice the

principles of environmental sustainability to which pharmaceutical companies in Switzerland have committed themselves<sup>54</sup>.

These results, together with the results of previous studies,<sup>8, 19, 51</sup> support the perception that discharges from pharmaceutical production sites via WWTPs might be an underestimated source of surface water contamination, even in countries such as Switzerland with high environmental standards. Further investigations are underway to assess how widespread factory discharges are on a national level. There are more than 250 pharmaceutical premises in Switzerland alone,<sup>55</sup> many of them with production or formulating activities and the relative contribution of these different activities to contaminant emissions is still unknown. To address this knowledge gap, comprehensive production information is needed from industrial partners. This information is essential to better understand the importance of emissions from pharmaceutical manufacturing on the total contaminants emitted via municipal WWTPs to surface waters.

## **Supporting information**

Details on chemicals, sampling sites, analytical method, data processing, quantification and identification of non-target compounds, as well as additional time profiles (PDF)

List of target analytes (XLSX)

List of isotope-labeled internal standards (TXT)

## **Acknowledgments**

This work was funded by the Swiss Federal Office for the Environment (FOEN). The authors thank the personnel of the two WWTPs for the sampling. We gratefully acknowledge the cantonal authorities for their support and their efforts in providing data on the WWTPs. We also thank Steffen Ruppe from the

495 “Amt für Umwelt und Energie, Kanton Basel Stadt” (AUE) for extracting and providing time series from the  
496 long-term LC-HRMS of international Rhine monitoring station (RUES) and for valuable comments on the  
497 manuscript. Furthermore, Jennifer Schollée is acknowledged for her helpful discussions and proofreading  
498 the manuscript.

- (1) Loos, R.; Carvalho, R.; Antonio, D. C.; Cornero, S.; Locoro, G.; Tavazzi, S.; Paracchini, B.; Ghiani, M.; Lettieri, T.; Blaha, L.; Jarosova, B.; Voorspoels, S.; Servaes, K.; Haglund, P.; Fick, J.; Lindberg, R. H.; Schwesig, D.; Gawlik, B. M., EU-wide monitoring survey on emerging polar organic contaminants in wastewater treatment plant effluents. *Water Research* **2013**, *47*, (17), 6475-6487.
- (2) Verlicchi, P.; Al Aukidy, M.; Zambello, E., Occurrence of pharmaceutical compounds in urban wastewater: Removal, mass load and environmental risk after a secondary treatment-A review. *Science of the Total Environment* **2012**, *429*, 123-155.
- (3) Luo, Y.; Guo, W.; Ngo, H. H.; Nghiem, L. D.; Hai, F. I.; Zhang, J.; Liang, S.; Wang, X. C., A review on the occurrence of micropollutants in the aquatic environment and their fate and removal during wastewater treatment. *Science of The Total Environment* **2014**, *473-474*, 619-641.
- (4) Sanchez, W.; Sremski, W.; Piccini, B.; Palluel, O.; Maillot-Marechal, E.; Betoulle, S.; Jaffal, A.; Ait-Aissa, S.; Brion, F.; Thybaud, E.; Hinfrey, N.; Porcher, J.-M., Adverse effects in wild fish living downstream from pharmaceutical manufacture discharges. *Environment International* **2011**, *37*, (8), 1342-1348.
- (5) Carlsson, G.; Orn, S.; Larsson, D. G. J., Effluent from bulk drug production is toxic to aquatic vertebrates. *Environmental Toxicology and Chemistry* **2009**, *28*, (12), 2656-2662.
- (6) Gunnarsson, L.; Kristiansson, E.; Rutgersson, C.; Sturve, J.; Fick, J.; Forlin, L.; Larsson, D. G. J., Pharmaceutical industry effluent diluted 1:500 effects global gene expression, cytochrome P450 1a activity, and plasma phosphatase in fish. *Environmental Toxicology and Chemistry* **2009**, *28*, (12), 2639-2647.
- (7) Larsson, D. G. J., Pollution from drug manufacturing: review and perspectives. *Philosophical Transactions of the Royal Society B-Biological Sciences* **2014**, *369*, (1656).
- (8) Scott, T.-M.; Phillips, P. J.; Kolpin, D. W.; Colella, K. M.; Furlong, E. T.; Foreman, W. T.; Gray, J. L., Pharmaceutical manufacturing facility discharges can substantially increase the pharmaceutical load to U.S. wastewaters. *Science of The Total Environment* **2018**, *636*, 69-79.
- (9) Kleywegt, S.; Payne, M.; Ng, F.; Fletcher, T., Environmental loadings of active pharmaceutical ingredients from manufacturing facilities in Canada. *Science of The Total Environment* **2019**, *646*, 257-264.
- (10) Cardoso, O.; Porcher, J.-M.; Sanchez, W., Factory-discharged pharmaceuticals could be a relevant source of aquatic environment contamination: Review of evidence and need for knowledge. *Chemosphere* **2014**, *115*, 20-30.
- (11) Hoerger, C. C.; Doerr, B.; Schlienger, C.; Straub, J. O., Environmental risk Assessment for the galenic formulation of solid medicinal products at Roche Basel, Switzerland. *Integrated Environmental Assessment and Management* **2009**, *5*, (2), 331-337.
- (12) Ort, C.; Lawrence, M. G.; Rieckermann, J.; Joss, A., Sampling for pharmaceuticals and personal care products (PPCPs) and illicit drugs in wastewater systems: Are your conclusions valid? A critical review. *Environmental Science & Technology* **2010**, *44*, (16), 6024-6035.
- (13) Howard, P. H.; Muir, D. C. G., Identifying new persistent and bioaccumulative organics among chemicals in commerce. III: Byproducts, impurities, and transformation products. *Environmental Science & Technology* **2013**, *47*, (10), 5259-5266.
- (14) Prasse, C.; Schluesener, M. P.; Schulz, R.; Ternes, T. A., Antiviral drugs in wastewater and surface waters: A new pharmaceutical class of environmental relevance? *Environmental Science & Technology* **2010**, *44*, (5), 1728-1735.
- (15) Stepien, D. K.; Puettmann, W., Source identification of high glyme concentrations in the Oder River. *Water Research* **2014**, *54*, 307-317.

- (16) Dsikowitzky, L.; Schwarzbauer, J., Hexa(methoxymethyl)melamine: An emerging contaminant in German rivers. *Water Environment Research* **2015**, *87*, (5), 461-469.
- (17) Ruff, M.; Mueller, M. S.; Loos, M.; Singer, H. P., Quantitative target and systematic non-target analysis of polar organic micro-pollutants along the river Rhine using high-resolution mass-spectrometry – Identification of unknown sources and compounds. *Water Research* **2015**, *87*, 145-154.
- (18) Krauss, M.; Hug, C.; Bloch, R.; Schulze, T.; Brack, W., Prioritising site-specific micropollutants in surface water from LC-HRMS non-target screening data using a rarity score. *Environmental Sciences Europe* **2019**, *31*.
- (19) Phillips, P. J.; Smith, S. G.; Kolpin, D. W.; Zaugg, S. D.; Buxton, H. T.; Furlong, E. T.; Esposito, K.; Stinson, B., Pharmaceutical formulation facilities as sources of opioids and other pharmaceuticals to wastewater treatment plant effluents. *Environmental Science & Technology* **2010**, *44*, (13), 4910-4916.
- (20) Schlusener, M. P.; Kunkel, U.; Ternes, T. A., Quaternary triphenylphosphonium compounds: A new class of environmental pollutants. *Environ Sci Technol* **2015**, *49*, (24), 14282-91.
- (21) Ruppe, S.; Griesshaber, D. S.; Langlois, I.; Singer, H. P.; Mazacek, J., Detective work on the Rhine river in Basel – Finding pollutants and polluters. *CHIMIA International Journal for Chemistry* **2018**, *72*, (7-8), 547-547.
- (22) Alygizakis, N. A.; Gago-Ferrero, P.; Hollender, J.; Thomaidis, N. S., Untargeted time-pattern analysis of LC-HRMS data to detect spills and compounds with high fluctuation in influent wastewater. *Journal of Hazardous Materials* **2019**, *361*, 19-29.
- (23) Chiaia-Hernandez, A. C.; Gunthardt, B. F.; Frey, M. P.; Hollender, J., Unravelling contaminants in the anthropocene using statistical analysis of liquid chromatography-high-resolution mass spectrometry nontarget screening data recorded in lake sediments. *Environmental Science & Technology* **2017**, *51*, (21), 12547-12556.
- (24) Peter, K. T.; Wu, C.; Tian, Z.; Kolodziej, E. P., Application of nontarget high resolution mass spectrometry data to quantitative source apportionment. *Environmental Science & Technology* **2019**, *53*, (21), 12257-12268.
- (25) Loos, M., enviMass version 4.0. Zenodo, 2018.
- (26) R Core Team, R: A language and environment for statistical computing. R Foundation for Statistical Computing: Vienna, Austria, 2017.
- (27) Gerrity, D.; Trenholm, R. A.; Snyder, S. A., Temporal variability of pharmaceuticals and illicit drugs in wastewater and the effects of a major sporting event. *Water Research* **2011**, *45*, (17), 5399-5411.
- (28) Nelson, E. D.; Do, H.; Lewis, R. S.; Carr, S. A., Diurnal Variability of Pharmaceutical, Personal Care Product, Estrogen and Alkylphenol Concentrations in Effluent from a Tertiary Wastewater Treatment Facility. *Environmental Science & Technology* **2011**, *45*, (4), 1228-1234.
- (29) Kot-Wasik, A.; Jakimska, A.; Śliwka-Kaszyńska, M., Occurrence and seasonal variations of 25 pharmaceutical residues in wastewater and drinking water treatment plants. *Environmental Monitoring and Assessment* **2016**, *188*, (12), 661.
- (30) Tai, Y. C.; Speed, T. P., A multivariate empirical Bayes statistic for replicated microarray time course data. *Annals of Statistics* **2006**, *34*, (5), 2387-2412.
- (31) Pence, H. E.; Williams, A., ChemSpider: An online chemical information resource. *J. Chem. Educ.* **2010**, *87*, (11), 1123-1124.
- (32) Ruttkies, C.; Schymanski, E. L.; Wolf, S.; Hollender, J.; Neumann, S., MetFrag relaunched: incorporating strategies beyond in silico fragmentation. *Journal of Cheminformatics* **2016**, *8*.
- (33) Horai, H.; Arita, M.; Kanaya, S.; Nihei, Y.; Ikeda, T.; Suwa, K.; Ojima, Y.; Tanaka, K.; Tanaka, S.; Aoshima, K.; Oda, Y.; Kakazu, Y.; Kusano, M.; Tohge, T.; Matsuda, F.; Sawada, Y.; Hirai, M. Y.; Nakanishi, H.; Ikeda, K.; Akimoto, N.; Maoka, T.; Takahashi, H.; Ara, T.; Sakurai, N.; Suzuki, H.;

- Shibata, D.; Neumann, S.; Iida, T.; Tanaka, K.; Funatsu, K.; Matsuura, F.; Soga, T.; Taguchi, R.; Saito, K.; Nishioka, T., MassBank: a public repository for sharing mass spectral data for life sciences. *Journal of Mass Spectrometry* **2010**, *45*, (7), 703-714.
- (34) Guijas, C.; Montenegro-Burke, J. R.; Domingo-Almenara, X.; Palermo, A.; Warth, B.; Hermann, G.; Koellensperger, G.; Huan, T.; Uritboonthai, W.; Aisporna, A. E.; Wolan, D. W.; Spilker, M. E.; Benton, H. P.; Siuzdak, G., METLIN: A technology platform for identifying knowns and unknowns. *Analytical Chemistry* **2018**, *90*, (5), 3156-3164.
- (35) Wittmer, I. K.; Bader, H. P.; Scheidegger, R.; Singer, H.; Lueck, A.; Hanke, I.; Carlsson, C.; Stamm, C., Significance of urban and agricultural land use for biocide and pesticide dynamics in surface waters. *Water Research* **2010**, *44*, (9), 2850-2862.
- (36) Masoner, J. R.; Kolpin, D. W.; Cozzarelli, I. M.; Barber, L. B.; Burden, D. S.; Foreman, W. T.; Forshay, K. J.; Furlong, E. T.; Groves, J. F.; Hladik, M. L.; Hopton, M. E.; Jaeschke, J. B.; Keefe, S. H.; Krabbenhoft, D. P.; Lowrance, R.; Romanok, K. M.; Rus, D. L.; Selbig, W. R.; Williams, B. H.; Bradley, P. M., Urban Stormwater: An Overlooked Pathway of Extensive Mixed Contaminants to Surface and Groundwaters in the United States. *Environ Sci Technol* **2019**, *53*, (17), 10070-10081.
- (37) Weissbrodt, D.; Kovalova, L.; Ort, C.; Pazhepurackel, V.; Moser, R.; Hollender, J.; Siegrist, H.; McArdell, C. S., Mass flows of X-ray contrast media and cytostatics in hospital wastewater. *Environmental Science & Technology* **2009**, *43*, (13), 4810-4817.
- (38) Ort, C.; van Nuijs, A. L. N.; Berset, J. D.; Bijlsma, L.; Castiglioni, S.; Covaci, A.; de Voogt, P.; Emke, E.; Fatta-Kassinos, D.; Griffiths, P.; Hernandez, F.; Gonzalez-Marino, I.; Grabic, R.; Kasprzyk-Hordern, B.; Mastroianni, N.; Meierjohann, A.; Nefau, T.; Ostman, M.; Pico, Y.; Racamonde, I.; Reid, M.; Slobodnik, J.; Terzic, S.; Thomaidis, N.; Thomas, K. V., Spatial differences and temporal changes in illicit drug use in Europe quantified by wastewater analysis. *Addiction* **2014**, *109*, (8), 1338-1352.
- (39) Bourgin, M.; Beck, B.; Boehler, M.; Borowska, E.; Fleiner, J.; Salhi, E.; Teichler, R.; von Gunten, U.; Siegrist, H.; McArdell, C. S., Evaluation of a full-scale wastewater treatment plant upgraded with ozonation and biological post-treatments: Abatement of micropollutants, formation of transformation products and oxidation by-products. *Water Research* **2018**, *129*, 486-498.
- (40) Carpenter, C. M. G.; Wong, L. Y. J.; Gutema, D. L.; Helbling, D. E., Fall Creek monitoring station: Using environmental covariates to predict micropollutant dynamics and peak events in surface water systems. *Environmental Science & Technology* **2019**, *53*, (15), 8599-8610.
- (41) European Chemicals Agency ECHA Substance information - Bis(2-(2-methoxyethoxy)ethyl) ether. <https://echa.europa.eu/substance-information/-/substanceinfo/100.005.086>, (November 12, 2019).
- (42) Schymanski, E. L.; Jeon, J.; Gulde, R.; Fenner, K.; Ruff, M.; Singer, H. P.; Hollender, J., Identifying small molecules via high resolution mass spectrometry: Communicating confidence. *Environmental Science & Technology* **2014**, *48*, (4), 2097-2098.
- (43) European Chemicals Agency ECHA Substance information - Methenamine. <https://echa.europa.eu/substance-information/-/substanceinfo/100.002.642>, (November 11, 2019).
- (44) Sorgi, K. L., Triethylamine. In *Encyclopedia of reagents for organic synthesis*, Paquette, L., Ed. John Wiley & Sons: 2001.
- (45) Brahmachari, G.; Nayek, N.; Nurjamal, K.; Karmakar, I.; Begam, S., Triethylamine a versatile organocatalyst in organic transformations: A decade update. *Synthesis* **2018**, *50*, (21), 4145-4164.
- (46) European Chemicals Agency ECHA Substance information - Triethylamine. <https://echa.europa.eu/substance-information/-/substanceinfo/100.004.064>, (November 11, 2019).
- (47) Swissmedic Product information. <http://www.swissmedicinfo.ch>, (November 11, 2019).

- 641 (48) Singer, H. P.; Wossner, A. E.; McArdell, C. S.; Fenner, K., Rapid screening for exposure to "non-  
642 target" pharmaceuticals from wastewater effluents by combining HRMS-based suspect screening  
643 and exposure modeling. *Environmental Science & Technology* **2016**, *50*, (13), 6698-6707.
- 644 (49) Hoppe, H.; Messmann, S.; Giga, A.; Gruning, H., Options and limits of quantitative and qualitative  
645 online-monitoring of industrial discharges into municipal sewage systems. *Water Science and  
646 Technology* **2009**, *60*, (4), 859-867.
- 647 (50) Dsikowitzky, L.; Schwarzbauer, J., Organic contaminants from industrial wastewaters:  
648 Identification, toxicity and fate in the environment. In *Pollutant Diseases, Remediation and  
649 Recycling*, Lichtfouse, E.; Schwarzbauer, J.; Robert, D., Eds. 2013; Vol. 4, pp 45-101.
- 650 (51) Larsson, D. G. J.; de Pedro, C.; Paxeus, N., Effluent from drug manufactures contains extremely  
651 high levels of pharmaceuticals. *Journal of Hazardous Materials* **2007**, *148*, (3), 751-755.
- 652 (52) Sim, W.-J.; Lee, J.-W.; Lee, E.-S.; Shin, S.-K.; Hwang, S.-R.; Oh, J.-E., Occurrence and distribution of  
653 pharmaceuticals in wastewater from households, livestock farms, hospitals and pharmaceutical  
654 manufactures. *Chemosphere* **2011**, *82*, (2), 179-186.
- 655 (53) United Nations (UN), Rio declaration on environment and development. *Principle 16*, 1992.
- 656 (54) scienceindustries Switzerland The Responsible Care Programme: our commitment to  
657 sustainability. <https://en.scienceindustries.ch/involvement/responsible-care>, (August 15, 2019).
- 658 (55) Swiss Federal Statistical Office GEOSTAT, Statistik der Unternehmensstruktur STATENT 2015. CH-  
659 2010 Neuchatel.