This document is the accepted manuscript version of the following article: Takatsu, K., & Brodersen, J. (2023). Repeated elevational clines of early life-history traits and their proximate mechanisms in brown trout. Freshwater Biology. https://doi.org/10.1111/fwb.14050

2 Repeated elevational clines of early life-history traits and their proximate mechanisms in

**brown trout** 

1

4

11

15

18

- 5 Kunio Takatsu<sup>1</sup>, Jakob Brodersen<sup>1,2</sup>
- <sup>1</sup>Department of Fish Ecology and Evolution, Center for Ecology, Evolution & Biogeochemistry,
- Eawag: Swiss Federal Institute of Aquatic Science and Technology, 6047 Kastanienbaum,
- 8 Switzerland
- <sup>2</sup>Department of Aquatic Ecology & Evolution, Institute of Ecology and Evolution, University of
- Bern, 3012 Bern, Switzerland
- 12 Correspondence: Kunio Takatsu
- Email: oniku029t@gmail.com
- http://orcid.org/0000-0002-2406-1006
- 16 KEYWORDS
- climate change, common-garden experiment, maternal effects, Salmo trutta, thermal adaptation
- 19 **RUNNING HEAD**
- 20 Elevational clines in brown trout traits

#### **ABSTRACT**

22

46

1. Climate warming imposes a severe threat to freshwater ecosystems, which are dominated by 23 ectotherms such as fish and aquatic insects. To better predict the effects of climate warming on 24 thermally sensitive ecosystems, information on how temperature affects individual traits within 25 populations is fundamental. 26 2. Patterns of intraspecific variation in thermal reaction norms along geographic thermal 27 gradients provide valuable information. Immediate temperature effects on individual traits can be 28 inferred from the shape of the thermal reaction norm. The way that temperature and associated 29 environmental conditions (to which populations have been exposed over generations) affect 30 individual traits through transgenerational plasticity in, and/or natural selection on, these traits 31 can also be inferred from patterns of trait variation along a geographic thermal gradient. 32 3. Many studies have documented patterns of intraspecific variation in thermal reaction norms 33 along geographic thermal gradients. However, most previous studies cannot exclude the 34 possibility that the observed geographic patterns are solely explained by random processes, such 35 as isolation by distance, due to the lack of replication of geographic gradients. Here, we show 36 consistent patterns in intraspecific trait variation along geographic thermal gradients using Salmo 37 trutta (brown trout), which is an ecologically and economically important fish in alpine streams. 38 4. We kept trout embryos collected from 52 families from 14 populations along wide and 39 replicated elevational gradients from three Alpine drainages (Danube, Po, and Rhine) in two 40 temperature treatments (3.2 and 6.2 °C). Timing and body size at emergence from the nest, which 41 are key early life-history traits of trout affecting their early growth and survival, were measured. 42 5. Besides faster embryonic development at warmer temperatures, we found that offspring from 43 low-elevation parents took very slightly more days to reach emergence from fertilisation and 44 were larger than offspring from high-elevation parents. Importantly, this was evident for all three 45 drainages. Further analyses found that the higher number of days until- and larger body length atemergence of low-elevation trout were mainly due to the larger eggs laid by low-elevation

females, which had larger body size than high-elevation females.

6. Trout female body size, which is positively correlated with egg size, is susceptible to

temperatures and associated environmental conditions. Consequently, climate warming may not

only immediately accelerate development rate but also shift timing and size at emergence through

egg size-mediated maternal effects within a relatively short time scale.

## 1 INTRODUCTION

Climate warming can affect various individual traits by accelerating physiological processes immediately and by indirectly influencing the strength and even nature of ecological interactions (Beacham & Murray, 1990; Rudolf & Roman, 2018; Bonacina *et al.*, 2022; Lindmark, Ohlberger & Gårdmark, 2022), eventually altering whole populations (Yang & Rudolf, 2010; Ohlberger, 2013). In addition to such immediate effects, environmental conditions, including temperature, to which populations have been exposed over generations can also affect various individual traits through transgenerational plasticity in- and/or natural selection on- the traits (Endler, 1986; Angilletta, 2009; Bell & Hellmann, 2019; Leicht & Seppälä, 2019), again eventually influencing whole populations (Pelletier, Garant & Hendry, 2009; Donelan *et al.*, 2020). Therefore, to better predict the effects of climate warming on natural populations, it is necessary to investigate both immediate temperature effects and effects of temperature to which populations have been exposed over generations on individual traits. Such studies are especially urgent for freshwater ecosystems as they are dominated by ectotherms, such as fish and aquatic insects, whose physiological processes are highly dependent on ambient temperatures (Angilletta, 2009).

A practical approach to investigating immediate temperature effects and effects of temperatures and associated environmental conditions to which populations have been exposed over generations on individual traits is to examine intraspecific variation in thermal reaction norms along the geographic thermal gradient (Conover, Duffy & Hice, 2009). Representative geographic thermal gradients are elevational and latitudinal gradients. It is reasonable to assume that each geographic population has been exposed to geographically specific environmental regimes for generations. This considered, any detected patterns in intraspecific trait variations for a given rearing temperature and shape of reaction norm along geographic thermal gradients are expected to reflect the consequences of transgenerational plasticity and/or natural selection driven by local environmental conditions. Also, the immediate temperature effects on the traits are shown as thermal reaction norms.

Many previous studies have described intraspecific variations in thermal reaction norms along latitudinal, elevational, and other geographic thermal gradients (Laugen, Laurila & Merilä, 2003; Conover *et al.*, 2009; Kojima *et al.*, 2020). However, most previous studies did not test the consistency of the patterns of trait variations along geographic thermal gradients, which is most often associated with the lack of independent replicates of the geographic thermal gradients (but see Kavanagh et al., 2010). Therefore, they cannot exclude the possibility that observed patterns in trait variations along geographic thermal gradients are solely explained by random processes, such as isolation by distance (Endler, 1977; Ibrahim, Nichols & Hewitt, 1996). To obtain robust evidence for inferring the effects of temperatures and associated environmental conditions to which populations have been exposed over generations, an experiment testing the consistency of the patterns of intraspecific trait variations along geographic thermal gradients is necessary.

Here, we report results from a common-garden experiment using *Salmo trutta* (i.e., brown trout), which is a common fish species in European streams, aimed at examining the consistency of the patterns of intraspecific trait variations along elevational gradients. Importantly, their wide elevational distribution in each drainage allowed us to have replicated elevational gradients (e.g., Fig. 1a,b). Specifically, we focused on the timing of- and body size at- the emergence from the redd, i.e., the nest. In salmonid fishes, hatchlings suffer from predation after emergence and

engage in intensive resource competition among conspecifics (Jonsson & Jonsson, 2011). The timing of- and body size at- emergence affect growth and survival during this critical period (Einum & Fleming, 2000b; Skoglund, Einum & Robertsen, 2011). At the same time, similar to most ectotherms, individuals during early life-stages are highly vulnerable to ambient environmental conditions (Miller et al., 1988; Jonsson & Jonsson, 2011). All this considered, optimal early life-history tactics are expected to be associated with elevational gradients along which environmental factors, such as temperature and community composition, vary. Moreover, the embryonic development of most salmonid fishes, including brown trout, is currently facing an exaggerated increase in ambient temperatures, partly due to hydropower plants. The embryonic development occurs in winter (Beacham & Murray, 1990; Jonsson & Jonsson, 2011), where energy requirements are high and associated release of water from reservoirs above hydropower plants results in up to a 5°C increase in downstream temperatures, due to the heat storage in reservoir water (Olden & Naiman, 2010). Given the ecological and economic importance of salmonid fishes (Jonsson & Jonsson, 2011), the information on how an increase in temperature affects the early life-history traits of brown trout could provide insights to help mitigate the effects of climate warming on the fish and the whole alpine river ecosystems.

97

98

99

100

101

102

103

104

105

106

107

108

109

110

111

112

113

114

115

116

117

118

119

120

121

In the present study, we kept brown trout embryos collected from 52 families from 14 populations along wide and replicated elevational gradients from three Alpine drainages (Fig. 1a,b) in two different rearing temperatures. We examined (1) reaction norms of timing and body size at emergence from the nest, (2) how they varied along elevational gradients, and (3) whether patterns of variation in the early life-history traits are consistent across three drainages.

Moreover, we measured maternal body size, egg volume, and several traits at hatching, which are acknowledged as traits affecting timing and body size at emergence (Einum & Fleming, 2000b a; Rollinson & Hutchings, 2010; Thorn & Morbey, 2018). Then, we examined the links between maternal traits and offspring traits to explore the mechanisms underlying variations in timing and

body size at emergence along elevational gradients.

123

124

125

128

133

135

141

146

122

## 2 MATERIALS AND METHODS

### 2.1 | Study system

Salmo trutta in the Swiss canton of Graübunden provides an excellent opportunity to have 126 replicated geographic thermal gradients. Headwaters of three major Alpine drainages (i.e., 127 Danube, Po, Rhine) are located within the relatively small region (Fig. 1a), which encompasses a part of the Alps. This allowed us to collect trout from a relatively wide elevation range from each 129 drainage. Notably, previous studies demonstrated that distinct genetic characters of populations 130 from the three drainages have persisted despite intensive stocking activities using several million 131 captive-reared trout (Keller, Taverna & Seehausen, 2011). Independent evolutionary histories, 132 therefore, have not been erased completely. Also, a consistent pattern of elevational changes of genetic structure was observed across drainages, suggesting natural population structure may also 134 not have been erased completely (Keller et al., 2011). We selected 14 collection sites to cover a wide elevation range across the three drainages. The elevation ranges of collections sites in 136 Danube, Po, and Rhine drainages were 1134 - 2073 m (number of collection site [N] = 2), 343 -137 1872 m (N=6), and 771-1886 m (N=6), respectively (Fig. 1a,b, Table S1). Because of the 138 relatively narrow elevation range of the Danube drainage within the area and the small number of 139 accessible streams in the drainage, we were only able to collect trout from two sites in the 140 drainage. Water temperature at each collection site was measured from November 2018 to February 2019 using temperature loggers (Hobo Pendant data loggers, Onset Computer 142 Corporation, USA), with the exception of the 968 m elevation site in the Rhine drainage, where 143 the logger was washed away (site ID 13 in Fig. 1a-c). This confirmed that trout in higher 144 elevation sites have generally experienced colder temperatures (Fig. 1c). 145

## 2.2 Gamete collection and in vitro fertilisation

From late October to early November 2018, we collected parent fish from each of 14 selected 147 sites. Adult trout were caught by electrofishing and then brought to hatcheries in Graübunden. 148 The trout were kept until they were ripe for spawning. In early November, we conducted in vitro 149 fertilisation at canton hatcheries. Before stripping eggs or sperms, each parental trout was 150 measured for standard length (hereafter, body length). We created two to six full-sib families 151 from each site by pairing randomly selected males and females and obtained 52 families in total 152 (Fig. 1b). The eggs were transferred to an experimental room in Eawag Kastanienbaum (transport 153 time was less than 6 hours). Eggs of each family were photographed (Tough TG-5, Olympus, 154 Tokyo, Japan). The captured images were used to measure the diameter of 10 eggs randomly sub-155 sampled from each family. We measured the maximum and minimum diameter of each of 10 sub-156 sampled eggs and then calculated the family-mean egg diameter (mm) and volume (mm<sup>3</sup>). All 157 measurements in this study were conducted using Image J (National Institute of Health, USA). 158 2.3 Common-garden experiment 159 We used eight vertical incubators (MariSource Inc., WA, USA) as experimental arenas, each 160 supplied by a recirculating supply of 200 L of oxygen-saturated water at an average flow of 0.6 L 161 per second. Thirty plastic mesh tubes (4.2 cm diameter × 6 cm tall, an acrylic panel was attached 162 to the bottom) were placed in each of six trays within an incubator to keep the eggs and 163 hatchlings from each family separately (i.e., 180 mesh tubes per incubator). Thirty litres of water 164 in all incubators was exchanged every two weeks to maintain water quality. A chiller (TK-2000, 165 TECO, Ravenna, Italy) installed in each incubator enabled us to control the water temperature. 166 We established cold (constant 3.2 °C) treatment using four incubators and warm (constant 6.2 °C) 167 treatment using the remaining four incubators. The temperatures of both treatments were within 168 the range of site mean temperatures during the incubation period in the wild (Fig. 1c), and at the 169 same time, within the suitable temperature range for embryonic development (Jonsson & 170

Jonsson, 2011). Within 12 h of fertilisation, 20 randomly selected eggs from each family were

assigned into one of 180 mesh tubes in each of the eight incubators. Thus, each of the 52 families had four replicates in each of the two temperature treatments (i.e., 208 replicates for each temperature treatment in total). The day we assigned eggs into the incubator was defined as day 1 of the experiment (Table S1).

172

173

174

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

190

191

192

193

194

195

196

The developing embryos were checked daily. All dead and/or unfertilised eggs were removed to maintain the water quality. We counted the number of newly hatched individuals in each replicate daily. The newly hatched individuals in each replicate were transferred to an empty mesh tube within the same tray every two days to keep eggs and hatchlings from each replicate separately. Before transferring the newly hatched individuals, we scanned the lateral aspect of all hatchlings (CanoScan 9000F Mark II, Canon, Tokyo, Japan). We measured the body length, yolk sac length, and yolk sac height of five hatchlings randomly selected from each of 416 replicates using the scanned images. For the 16 replicates with less than five hatchlings (probably due to the low in vitro fertilisation rate of certain families), we measured all hatchlings. We estimated yolk sac volume using the following equation:  $V = (\pi/6) \times L \times H^2$  (Blaxter & Hempel, 1963), where L is the yolk sac length and H is the yolk sac height. For later analysis, we calculated replicatemean values of all measured traits. Moreover, within 48 hours after hatching, one randomly selected hatchling from each of 416 replicates was measured for oxygen consumption rate (O<sub>2</sub> μg/h) (details are described in the Supplementary Information). The oxygen consumption rate of each individual was measured at the temperature corresponding to their incubation temperature (i.e., 3.2 or 6.2 °C).

The hatchlings were kept until emergence (here, it is defined as the timing when hatchlings almost completely absorbed their yolk sac). The yolk sac absorption of hatchlings was identified visually following the approach used in Metcalfe and Thorpe (1992). We checked the size of the remaining yolk sac of each individual once per week. When the yolk sac almost disappeared, the hatchlings were removed from the mesh tube, euthanised using an overdose of MS-222, and then

scanned from the lateral side. We calculated the degree-days from day 1 to day at the yolk absorption of each hatchling by adding up the daily mean temperature during the period. Also, we measured the body length of five hatchlings randomly selected from each replicate using the scanned images. Then, we calculated the mean degree-days from fertilisation to yolk absorption (hereafter, degree-days until emergence) and mean body length at yolk absorption (hereafter, body length at emergence) in each replicate, which were subsequently used in the statistical analyses. We calculated the survival rate from hatching to emergence by dividing the number of individuals at emergence by the number of individuals at hatching. In three out of the 416 replicates, we lost hatchlings when moving them from the incubators to the scanner and back to the incubators. We, therefore, obtained complete data set from the 413 replicates and used these data for the later analyses.

## 2.4 Data analyses

197

198

199

200

201

202

203

204

205

206

207

- All statistical analyses described below were conducted using R (version 4.0.4. R Development
- 210 Core Team, 2019) within R studio interface (version 1.4.1717. R studio team 2021).
- 2.4.1 Variations in thermal reaction norms of degree-days until- and body length at-
- emergence along elevational gradients
- We performed linear mixed models (LMM) using the lmer function in the lme4 package (Bates et 213 al., 2015) to assess how degree-days until- and body length at- emergence vary depending on 214 temperature treatment, the elevation of collection site, and drainage. The interaction between 215 temperature treatment and elevation of collection site was considered in the models to examine 216 how the thermal reaction norms vary with the elevation of collection site. We also considered the 217 interaction between drainage and elevation of collection site in the models to examine how the 218 patterns of variation in early life-history traits along the elevational gradients differ among the 219 drainages. We considered family ID nested within collection site ID as a random factor to account 220 for non-independence among replicates originating from the same family caught at the same site. 221

A random slope of tray within incubator was also considered since water temperature in lower trays might have been warmer than in upper tray because of the structural issues of the incubators. Then, the initial models (Table 1) were simplified using the backward stepwise selection with the step function (Satterthwaite approximation) in the ImerTest package to select minimum adequate models. Both the degree-days until emergence and body length at emergence were log-transformed. Prior to the analyses, we examined whether the data satisfied the assumptions of linear models. The normality of data from each site in each temperature treatment was visually assessed by using QQ-plot, and we used Levene's test to examine the homoscedasticity of variance. The data generally satisfied the assumptions of linear models, although there were a few exceptions.

We performed the same LMM analysis and subsequent model selection on the survival rate from hatching to emergence. Since we kept the hatchings within the suitable temperature range for development (Jonsson & Jonsson, 2011), the survival rate was generally high (mean survival rate was 94.4 %). In the final model, no explanatory variables were included (Table S2).

## 2.4.2 Mechanisms underlying the trait variation along elevational gradients

Therefore, we did not consider the survival rate further in this study.

To explore general mechanisms underlying the consistent trait variation along elevational gradients detected in the first analyses, we examined the links between the elevation of collection site, maternal body length, egg volume, traits at hatching, and the degree-days until- and body length at- emergence in each temperature treatment by using piecewise structural equation models (piecewiseSEM) (Lefcheck, 2016). While we acknowledged that paternal traits could affect the traits of hatchlings, we did not consider paternal effects. This is because variations in the early development of salmonids are largely attributable to the female parent (Janhunen, Piironen & Peuhkuri, 2010; Thorn & Morbey, 2018). Also, while we created families by pairing randomly selected females and males within sites, their body lengths were highly correlated with

each other because both decreased with the elevation of collection site. Due to the data structure (i.e., no egg volume data for each replicate), family-mean trait values (log-transformed) were used for the analyses (number of families [N] = 52).

Prior to the second analyses, to reduce the complexity of the piecewiseSEMs as recommended by Lefcheck (2016), we screened the following six measured traits potentially explaining the variation in degree-days until- and body length at- emergence to remove those not correlated with the elevation of collection site: (1) maternal body length, (2) family-mean egg volume, and (3-6) family-mean traits at hatching (i.e., degree days from fertilisation to hatching, body length, yolk sac volume, and oxygen consumption rate) (Fig. 2). Since traits at hatching are highly correlated with egg volume (Vøllestad & Lillehammer, 2000; Régnier *et al.*, 2013; van Leeuwen *et al.*, 2017), we used residuals of each of the traits at hatching after regression against the egg volume. The residuals were calculated from linear regressions of log-transformed traits at hatching on log-transformed egg volume. We performed LMM analyses to examine whether the traits vary with elevation. We considered site ID nested within drainage as a random factor. For the traits at hatching, we also considered temperature treatment and interaction between temperature treatment and elevation of collection site as additional explanatory variables. The initial models (Table 2) were again simplified using the step function.

Then, using the elevation of collection site, the traits in which a significant effect of elevation of collection site was detected in the preliminary analyses (i.e., maternal body length and egg volume [see results]), and degree-days until- and body length at- emergence, we constructed picewiseSEMs for each temperature treatment separately. All variables were standardised (i.e., mean was 0 and SD was 1) prior to the analysis. The piecewiseSEM models were initially composed of the four component models describing all direct and indirect effects of elevation of collection site on the degree-days until- and body length at- emergence (Fig. 3a). In each component model, site ID nested within drainage was considered as a random factor to account

for non-independence among families originating from the same site within the same drainage. To avoid overparameterisation of the models, we removed random factors whose standard deviation was estimated to be zero from the models. The initial models were assessed using Shipley's test of the d-separation. When the d-separation test revealed significant unconsidered paths, the paths were added to the model if there was a plausible biological explanation for it. Then, using the variance inflation factor (VIF), we assessed the level of multicollinearity among explanatory variables in each component model. VIF never exceeded 2, indicating acceptable levels of multicollinearity (Dormann *et al.*, 2013). The models were then simplified following the backwards stepwise removal of the non-significant paths (P > 0.05). We assessed the models by using Fisher's C statistics and AICc (AIC corrected for small sample sizes) value to select among alternative models.

#### 3 RESULTS

3.1| Variations in thermal reaction norms of degree-days until- and body length atemergence along elevational gradients

The temperature treatment, elevation of collection site, and interaction between temperature treatment and elevation of collection site were included in the final models of both degree-days until emergence (i.e., degree-days from fertilisation to yolk absorption) and body length at emergence (Table 1). The fact that the interaction between the elevation of collection site and drainage was not included in the final models (Table 1) suggest that the patterns of trait variations along the elevational gradients detailed below were consistent across the drainages.

The degree-days until emergence in the warm treatment (796.5  $\pm$  30.7 degree-days [treatment-mean  $\pm$  SD]) was 1.1 times greater than in the cold treatment (708.8  $\pm$  24.1 degree-days) (estimate  $\pm$  SE = 7.2  $\times$  10<sup>-2</sup>  $\pm$  7.2  $\times$  10<sup>-3</sup>, *t*-value = 10.0, P < 0.0001, Table S3) (Fig. 1d). In terms of the number of days until emergence, hatchlings kept in the warm treatment reached the

emergence stage about three months earlier than in the cold treatment. The thermal reaction norm 297 varied with the elevation of collection site. Specifically, the degree of difference in the degree-298 days between warm and cold treatments increased by about 16 degree-days as the elevation of 299 collection site decreased by 1000 m (estimate  $\pm$  SE = -7.9  $\times$  10<sup>-6</sup>  $\pm$  1.5  $\times$  10<sup>-6</sup>, t-value = -5.4, P < 300 0.0001) (Fig. 1d). The degree-days until emergence increased with decreasing the elevation of 301 collection site. For instance, in the cold treatment, the degree-days until emergence increased by 302 about 13 degree-days (i.e., about four days) as the elevation of collection site decreased by 1000 303 m (Fig. 1d) (estimate  $\pm$  SE =  $-8.2 \times 10^{-6} \pm 3.4 \times 10^{-6}$ , t-value = -2.4, P = 0.04). 304 Although temperature treatment was included in the final model (Table 1, Fig. 1e), the body 305 length of hatchlings in the warm (21.3  $\pm$  1.3 mm [treatment-mean  $\pm$  SD]) and cold (21.3  $\pm$  1.3 306 mm) treatments were almost the same (estimate  $\pm$  SE = 3.9  $\times$  10<sup>-3</sup>  $\pm$  2.2  $\times$  10<sup>-3</sup>, t-value = 1.8, P = 307 0.08, Table S4). The hatchlings originating from low-elevation sites grew slightly larger when 308 reared in the warm treatment, while hatchlings originating from high-elevation sites grew slightly 309 larger when reared in the cold treatment (Fig. 1e) (estimate  $\pm$  SE = -2.9  $\times$  10<sup>-6</sup>  $\pm$  1.4  $\times$  10<sup>-6</sup>, t-value 310 = -2.1, P = 0.04). The body length at emergence increased with decreasing the elevation of 311 collection site (Fig. 1e). For instance, in the cold treatment, the body length of hatchlings 312 increased by 5% per 1000 m reduction in the elevation (estimate  $\pm$  SE = -2.3 × 10<sup>-5</sup>  $\pm$  6.8 × 10<sup>-6</sup>, 313 t-value = -3.4, P = 0.001) (Fig 1e). Drainage was included in the final model (Table 1). Post-hoc 314 comparison using Tukey's test found that hatchlings originating from the Danube (21.6  $\pm$  0.9 mm 315 [drainage-mean  $\pm$  SD]) were larger than from the Po (20.8  $\pm$ 1.5 mm) (t-ratio = 2.8, P = 0.02). 316 Hatchlings originating from the Rhine (21.6  $\pm$  1.2 mm) were also larger than from the Po, while 317 the difference was marginally non-significant (t-ratio = -2.4, P = 0.05). No significant difference 318 was found between Rhine and Danube (t-ratio = 0.9, P = 0.6). 319 3.2 Mechanisms underlying the consistent patterns of elevational trait variation 320

Variation in maternal body length, egg volume, and traits at hatching

Among the six traits of interest (maternal body length, egg volume, and residuals of traits at 322 hatching after regression against egg volume [body length at-, degree-days until-, yolk sac 323 volume at-, and oxygen consumption rate at- hatching]), the elevation of collection site was 324 included in the final models of maternal body length and egg volume (Fig. 2 and Table 2). 325 Specifically, maternal body length and egg volume increased by 16% (estimate  $\pm$  SE = -6.5  $\times$  10<sup>-5</sup> 326  $\pm 2.4 \times 10^{-5}$ , t-value = -2.7, P = 0.02, Table S5) and 21 % (estimate  $\pm$  SE = -8.3  $\times 10^{-5} \pm 3.2 \times 10^{-5}$ 327  $^{5}$ , t-value = -2.6, P = 0.03, Table S6) per 1000 m reduction in elevation, respectively (Fig. 2a,b). 328 Regarding the residuals of traits at hatching after regression against egg volume, only 329 temperature treatment was included in the final models (Fig. 2c-f and Tables 2, S7-10). 330 Links between the elevation of collection site, maternal body length, egg volume, and degree-331 days until- and body length at- emergence 332 All final piecewise SEM models had adequate fit to the data (Fisher's C statistic with P > 0.05, 333 Fig. 3b,c). They revealed that egg size-mediated maternal effects are the key mechanisms 334 underlying the increase in the degree-days until- and body length at- emergence with decreasing 335 the elevation of collection site. Specifically, females originating from low-elevation sites were 336 larger than from high-elevation sites, thereby producing larger eggs. The hatchlings from larger 337 eggs used more degree-days until emergence and grew larger than those from smaller eggs. On 338 the other hand, for a given egg volume, hatchlings from larger females used fewer degree-days 339 until emergence than those from smaller females. The direct negative link between the elevation 340 of collection site and degree-days until emergence was included in both temperature treatments, 341 even after accounting for the effects of maternal body length and egg volume. 342

343

344

## 4 DISCUSSION

Many studies have documented patterns of intraspecific trait variations along geographic thermal gradients and have provided valuable information for inferring how temperatures and associated

environmental conditions (to which populations have been exposed over generations) affect traits through transgenerational plasticity in- and/or natural selection on- these traits (Conover *et al.*, 2009). However, since most studies did not test the consistency of the pattern (but see Kavanagh et al., 2010), they cannot rule out the possibility that the patterns were solely explained by random processes. Here, by conducting a common-garden experiment using the brown trout from wide and replicated elevational gradients from three Alpine drainages, we showed consistent patterns of variation in key early life-history traits along elevational gradients. Specifically, degree-days until- and body length at- emergence, both affecting early growth and survival of fish (Einum & Fleming, 2000b; Skoglund *et al.*, 2011), increased with decreasing elevation of origin consistently across the three drainages.

We also found variations in thermal reaction norms of early life-history traits along the elevational gradients. Similar to previous studies (Beacham & Murray, 1990; Jonsson & Jonsson, 2011; Réalis-Doyelle *et al.*, 2016), warmer rearing temperatures markedly increased degree-days until emergence. Immediate temperature effects increased slightly with decreasing elevation of origin. Also, an increase in temperature slightly enhanced growth in size of hatchlings originating from lower elevations, but in contrast, it slightly reduced growth of hatchlings originating from higher elevations. These results suggest that the effects of climate warming on trout populations could vary with elevation, given the importance of early life-history traits in determining early growth and survival (Einum & Fleming, 2000b; Jonsson & Jonsson, 2011; Skoglund *et al.*, 2011). We have to mention, however, that the degrees of variation in thermal reaction norms are considered negligibly small, although patterns were statistically significant. The subtle differences in thermal reaction norm in timing and body length at emergence are unlikely to be reflected in early growth and survival, considering within-stream heterogeneity in temperature (Leach & Moore, 2014; Sullivan et al., 2021).

An increase in egg size with decreasing elevation of origin was a key mechanism underlying

origin. For the degree-days until emergence, the egg size-mediated maternal effects generally followed the relationship between egg size and development rate predicted by processes controlled by metabolic rate (Gillooly, Londoño & Allen, 2008). However, this may not always be the case for salmonids. Although there are studies that have shown deceleration in early development rate with increasing egg size (Einum & Fleming, 2000a; Rollinson & Hutchings, 2010), some studies showed no relationship or even the opposite relationship (Einum & Fleming, 1999; van Leeuwen et al., 2017). The discrepancy of reported patterns among studies suggests diversity in egg-size mediated maternal effects. Interestingly, for a given egg size, hatchlings from larger females developed faster than those from smaller females. Non-egg size-mediated maternal effects suggest that larger females producing larger eggs could somehow compensate for egg size-mediated maternal effects. It has been shown that, for example, hormone and carotenoid content of eggs can affect early development of hatchlings (Burton et al., 2011; Wilkins *et al.*, 2017). For a better understanding of non-egg size-mediated maternal effects, examining how egg chemical composition varies with maternal body length is an interesting next step. Even after accounting for maternal effects, the direct link between the elevation of origin and degree-days until emergence still remained. This suggests the possibility of local adaptation and grandparental effects on embryonic development. The relationship between offspring size and offspring fitness is fundamental in determining

the increase in the degree-days until- and body length at- emergence with decreasing elevation of

372

373

374

375

376

377

378

379

380

381

382

383

384

385

386

387

388

389

390

391

392

393

394

395

396

maternal investment per offspring (offspring size) (Smith & Fretwell, 1974; Venable, 1992; Rollinson & Hutchings, 2013). The present study suggests a shift in the relationship, and hence in optimal egg size, along the elevational gradient. In salmonid fishes, hatchlings engage in intensive intraspecific resource competition and suffer from predation during early free-swimming stages (Jonsson & Jonsson, 2011). During the early critical period of salmonid fishes, larger body size and faster development are generally thought to be advantageous for survival

and growth (Einum & Fleming, 2000b; Skoglund *et al.*, 2011). However, it is difficult for hatchlings to increase body size and accelerate their development rate simultaneously because of the constraints imposed by egg size. Therefore, the relationship between egg size and hatchling fitness could be determined by the relative importance of fitness benefits obtained from an increase in body size and fitness losses caused by a deceleration in development rate through an increase in egg size. The increases in degree-days until- and body length at- emergence with decreasing elevation of origin suggest that fitness benefits obtained from an increase in body size could become relatively more important at lower elevations, and thus an optimal egg size at low elevations shifts towards larger size.

The shift in the relative importance of fitness benefits obtained from an increase in body size and fitness losses caused by a deceleration in development rate through an increase in egg size along the elevational gradient seems reasonable. Given the acceleration of development rate at warmer temperatures shown in the present and many previous studies (Beacham & Murray, 1990; Jonsson & Jonsson, 2011; Réalis-Doyelle *et al.*, 2016), hatchlings may develop faster in warm, low elevations. Moreover, in our study system, an increase in body size through an increase in egg size may only slightly decelerate development rate. The slight deceleration of development rate due to an increase in egg size could be compensated easily by, for example, nest environments chosen by parents, considering the marked effects of rearing temperatures on development rate and within-stream temperature variation (Leach & Moore, 2014; Sullivan et al., 2021). Importantly, hatchlings are likely to suffer more intensive competition and predation at lower elevations due to, for example, higher conspecific and predator density (Schager, Peter & Burkhardt-Holm, 2007; Buisson, Blanc & Grenouillet, 2008; Jones *et al.*, 2019). All this considered, fitness benefits obtained from an increase in body size could become relatively more important at lower elevations.

Alternatively, there is a possibility that consistent patterns of variation in early life-history

traits along elevational gradients are just by-products reflecting a shift in female body length at reproduction along elevational gradients. For instance, in most ectotherms, an increase in size at maturation is often caused by a decrease in temperature and an improvement in food conditions (Berrigan & Charnov, 1994). While temperatures increase with decreasing elevations, food conditions can improve as elevation decreases, considering the higher temperature and nutrient availability at low elevations (Ward, 1998). If the latter is relatively more important in determining the female size at reproduction, female size should increase with decreasing elevation. In support of this, Bærum et al. (2013) showed that an increase in temperature during the growth period enhances growth of matured brown trout, perhaps by increasing prey availability. The plastic increase in female size at reproduction in response to warmer temperatures could result in an increase in egg size and, eventually, degree-days until- and body length at-emergence. Considering the possible importance of female body size plasticity in shaping variation in early-life history traits, we cannot completely rule out the possibility that consistency in the patterns of variation in early life-history traits across drainages is due to stocking activities that homogenise populations, although populations from the three drainages still possess distinct genetic characteristics (Keller et al., 2011).

Especially in alpine streams, where salmonid fishes are often dominant fish and function as keystone species, hydropower plants arguably plays a key role in recent temperature changes, while also affecting various environmental factors (Halleraker *et al.*, 2007; Olden & Naiman, 2010; Lange *et al.*, 2018). Hydropower operation caused increases of up to 5 °C in downstream temperature during winter but decreases of up to 15 °C in downstream temperature during summer (Olden & Naiman, 2010). The increase in winter temperature, when embryonic development of most salmonids occurs, could significantly accelerate emergence, as the present study and previous studies have shown (Beacham & Murray, 1990; Jonsson & Jonsson, 2011; Réalis-Doyelle *et al.*, 2016). In addition to the immediate temperature effects, given the

importance of egg size in determining timing and body size at emergence, there is a possibility that the decrease in summer temperature and associated changes in environmental conditions could affect early life-history traits by influencing maternal reproductive strategies. Moreover, we note that the actual timing of emergence is determined by spawning date and embryonic development rate, while we focused only on the latter in this study. Therefore, studying how decreased summer temperatures and associated changes in environmental conditions affecting female reproductive traits (such as size and timing at spawning and egg size and quality) is a valuable next step to better understand the effects of hydropower plant-induced temperature changes on the key early life-history traits of this economically and ecologically important fish.

#### **ACKNOWLEDGEMENTS**

We thank Livio Costa, Nicola Gaudenz, Linard Jäger, Curdin Meiler, Marcel Michel, Flavio Nollo, Roland Tomaschett, Laetitia Wilkins, and fisheries managers in Canton Graübunden for their support during the collection of parental trout in the field and keeping them and stripping eggs and sperms in hatcheries. We thank Darryl McLennan, Gaurav Baruah, Dominique Stalder, Brigitte Germann, Oliver Selz, Nicola Sperlich, Dyer Shannon, Kurt Schmid, Grégoire Saboret, Martin Schmid, Davide Vanzo, Daniel Steiner, Andreas Taverna, Pascal Reichlin, Karin Beck, Salome Mwaiko, and members of the Fish Ecology & Evolution department of Eawag for giving us constructive comments on experimental design and support during fieldwork and experiment. We are grateful to Jacques Labonne, Angus McIntosh, Belinda Robson, and two anonymous reviewers for their constructive comments that improved this manuscript. Fish sampling, in vitro fertilisation, and the laboratory experiment were conducted with permission (30513, LU05/18) by the Veterinary Office of the Swiss Canton of Luzern.

#### **FUNDING**

472	This work was supported by the Swiss Innovation Agency Innosuisse (contract 1155002546) and
473	is part of the Swiss Competence Center for Energy Research - Supply of Electricity (SCCER-
474	SoE).
475	
476	DATA AVAILABILITY STATEMENT
477	The data associated with this paper are available from the Dryad Digital Repository
478	https://doi.org/10.5061/dryad.crjdfn37r
479	
480	CONFLICT OF INTEREST
481	There is no conflict of interest.
482	
483	REFERENCES
484	Angilletta M.J. (2009). Adaptation to Anthropogenic Climate Change. In: Thermal Adaptation.
485	Bærum K.M., Haugen T.O., Kiffney P., Moland Olsen E. & Vøllestad L.A. (2013). Interacting
486	effects of temperature and density on individual growth performance in a wild population of
487	brown trout. Freshwater Biology 58, 1329–1339. https://doi.org/10.1111/fwb.12130
488	Bates D., Mächler M., Bolker B. & Walker S. (2015). Fitting Linear Mixed-Effects Models Using
489	lme4. Journal of Statistical Software 67. https://doi.org/10.18637/jss.v067.i01
490	Beacham T.D. & Murray C.B. (1990). Temperature, egg size, and development of embryos and
491	alevins of five species of Pacific salmon: A comparative analysis. Transactions of the
492	American Fisheries Society 119, 927–945. https://doi.org/10.1577/1548-
493	8659(1990)119<0927:tesado>2.3.co;2
494	Bell A.M. & Hellmann J.K. (2019). An integrative framework for understanding the mechanisms
495	and multigenerational consequences of transgenerational plasticity. Annual Review of
496	Ecology, Evolution, and Systematics 50, 97–118, https://doi.org/10.1146/annurey-ecolsys-

110218-024613 497 Berrigan D. & Charnov E.L. (1994). Reaction norms for age and size at maturity in response to 498 temperature: a puzzle for life historians. Oikos 70, 474–478 499 Blaxter J.H.S. & Hempel G. (1963). The influence of egg size on herring larvae (Clupea 500 harengus 1.). ICES Journal of Marine Science 28, 211–240. 501 https://doi.org/10.1093/icesjms/28.2.211 502 Bonacina L., Fasano F., Mezzanotte V, Fornaroli R. (2022). Effects of water temperature on 503 freshwater macroinvertebrates: a systematic review. Biological Reviews, 504 https://doi.org/10.1111/brv.12903 505 Buisson L., Blanc L. & Grenouillet G. (2008). Modelling stream fish species distribution in a 506 river network: The relative effects of temperature versus physical factors. Ecology of 507 Freshwater Fish 17, 244–257. https://doi.org/10.1111/j.1600-0633.2007.00276.x 508 Burton T., Hoogenboom M.O., Armstrong J.D., Groothuis T.G.G. & Metcalfe N.B. (2011). Egg 509 hormones in a highly fecund vertebrate: Do they influence offspring social structure in 510 competitive conditions? Functional Ecology 25, 1379–1388. https://doi.org/10.1111/j.1365-511 2435.2011.01897.x 512 Conover D.O., Duffy T.A. & Hice L.A. (2009). The covariance between genetic and 513 environmental influences across ecological gradients: Reassessing the evolutionary 514 significance of countergradient and cogradient variation. Annals of the New York Academy of 515 Sciences 1168, 100–129. https://doi.org/10.1111/j.1749-6632.2009.04575.x 516 Donelan S.C., Hellmann J.K., Bell A.M., Luttbeg B., Orrock J.L., Sheriff M.J., et al. (2020). 517 Transgenerational plasticity in human-altered environments. Trends in Ecology and 518 Evolution 35, 115–124. https://doi.org/10.1016/j.tree.2019.09.003 519 Dormann C.F., Elith J., Bacher S., Buchmann C., Carl G., Carré G., et al. (2013). Collinearity: A 520

review of methods to deal with it and a simulation study evaluating their performance.

Ecography 36, 27–46. https://doi.org/10.1111/j.1600-0587.2012.07348.x 522 Einum S. & Fleming I.A. (2000a). Highly fecund mothers sacrifice offspring survival to 523 maximize fitness. *Nature* **405**, 565–567. https://doi.org/10.1038/35014600 524 Einum S. & Fleming I.A. (1999). Maternal effects of egg size in brown trout (Salmo trutta): 525 norms of reaction to environmental quality. Proceedings of the Royal Society of London. 526 Series B 266, 2095–2100 527 Einum S. & Fleming I.A. (2000b). Selection against late emergence and small offspring in 528 Atlantic salmon (Salmo salar). Evolution 54, 628–639. https://doi.org/10.1111/j.0014-529 3820.2000.tb00064.x 530 Endler J.A. (1977). Geographic variation, speciation, and clines. Princeton University Press, 531 Princeton, New Jersey, USA. 532 Endler J.A. (1986). Natural Selection in the Wild. Princeton University Press, Princeton, New 533 Jersey, USA. 534 Gillooly J.F., Londoño G.A. & Allen A.P. (2008). Energetic constraints on an early 535 developmental stage: A comparative view. *Biology Letters* 4, 123–126. 536 https://doi.org/10.1098/rsb1.2007.0460 537 Halleraker J.H., Sundt H., Alfredsen K.T. & Dangelmaier G. (2007). Application of multiscale 538 environmental flow methodologies as tools for optimized management of a Norwegian 539 regulated national salmon watercourse. River Research and Applications 23, 493–510. 540 https://doi.org/10.1002/rra.1000 541 Ibrahim K.M., Nichols R.A. & Hewitt G.M. (1996). Spatial patterns of genetic variation 542 generated by different forms of dispersal during range expansion. Heredity 77, 282–291 543 Janhunen M., Piironen J. & Peuhkuri N. (2010). Parental effects on embryonic viability and 544 growth in Arctic charr Salvelinus alpinus at two incubation temperatures. Journal of Fish 545 Biology 76, 2558–2570. https://doi.org/10.1111/j.1095-8649.2010.02648.x 546

- Jones D.A., Akbaripasand A., Nakagawa S. & Closs G.P. (2019). Landscape features determine 547 brown trout population structure and recruitment dynamics. Ecology of Freshwater Fish 28, 548 554–562. https://doi.org/10.1111/eff.12474 549
- Jonsson B. & Jonsson N. (2011). Ecology of Atlantic Salmon and Brown Trout, 1st edn. Springer 550 Netherlands. 551
- Kavanagh K.D., Haugen T.O., Gregersen F., Jernvall J. & Vøllestad L.A. (2010). Contemporary 552 temperature-driven divergence in a Nordic freshwater fish under conditions commonly 553 thought to hinder adaptation. BMC Evolutionary Biology 10. https://doi.org/10.1186/1471-554
- Keller I., Taverna A. & Seehausen O. (2011). Evidence of neutral and adaptive genetic 556 divergence between European trout populations sampled along altitudinal gradients. 557

2148-10-350

555

- Molecular Ecology 20, 1888–1904. https://doi.org/10.1111/j.1365-294X.2011.05067.x 558
- Kojima W., Nakakura T., Fukuda A., Lin C.P., Harada M., Hashimoto Y., et al. (2020). Latitudinal 559 cline of larval growth rate and its proximate mechanisms in a rhinoceros beetle. Functional 560 Ecology 34, 1577–1588. https://doi.org/10.1111/1365-2435.13572 561
- Lange K., Meier P., Trautwein C., Schmid M., Robinson C.T., Weber C., et al. (2018). Basin-562 scale effects of small hydropower on biodiversity dynamics. Frontiers in Ecology and the 563 Environment 16, 397–404. https://doi.org/10.1002/fee.1823 564
- Laugen A.T., Laurila A. & Merilä J. (2003). Latitudinal and temperature-dependent variation in 565 embryonic development and growth in Rana temporaria. Oecologia 135, 548-554. 566 https://doi.org/10.1007/s00442-003-1229-0
- Leach J.A. & Moore R.D. (2014). Winter stream temperature in the rain-on-snow zone of the 568 Pacific Northwest: Influences of hillslope runoff and transient snow cover. Hydrology and 569 Earth System Sciences 18, 819–838. https://doi.org/10.5194/hess-18-819-2014 570
- van Leeuwen T.E., Killen S.S., Metcalfe N.B. & Adams C.E. (2017). Differences in early 571

developmental rate and yolk conversion efficiency in offspring of trout with alternative life 572 histories. Ecology of Freshwater Fish 26, 371–382. https://doi.org/10.1111/eff.12281 573 Lefcheck J.S. (2016). piecewiseSEM: Piecewise structural equation modelling in r for ecology, 574 evolution, and systematics. *Methods in Ecology and Evolution* 7. 575 https://doi.org/10.1111/2041-210X.12512 576 Leicht K. & Seppälä O. (2019). Direct and transgenerational effects of an experimental heatwave 577 on early life stages in a freshwater snail. Freshwater Biology 64, 2131–2140. 578 https://doi.org/10.1111/fwb.13401 579 Lindmark M., Ohlberger J. & Gårdmark A. (2022). Optimum growth temperature declines with 580 body size within fish species. Global Change Biology 28, 2259–2271. 581 https://doi.org/10.1111/gcb.16067 582 Metcalfe N.B. & Thorpe J.E. (1992). Early predictors of life-history events: the link between first 583 feeding date, dominance and seaward migration in Atlantic salmon, Salmo salar L. Journal 584 of Fish Biology 41, 93–99. https://doi.org/10.1111/j.1095-8649.1992.tb03871.x 585 Miller T.J., Crowder L.B., Rice J.A. & Marschall E.A. (1988). Larval size and recruitment 586 mechanisms in fishes: toward a conceptual framework. Canadian Journal of Fisheries and 587 Aquatic Sciences 45, 1657–1670. https://doi.org/10.1139/f88-197 588 Ohlberger J. (2013). Climate warming and ectotherm body size - from individual physiology to 589 community ecology. Functional Ecology 27, 991–1001. https://doi.org/10.1111/1365-590 2435.12098 591 Olden J.D. & Naiman R.J. (2010). Incorporating thermal regimes into environmental flows 592 assessments: Modifying dam operations to restore freshwater ecosystem integrity. 593 Freshwater Biology 55, 86–107. https://doi.org/10.1111/j.1365-2427.2009.02179.x 594 Pelletier F., Garant D. & Hendry A.P. (2009). Eco-evolutionary dynamics. *Philosophical* 595 *Transactions of the Royal Society B: Biological Sciences* **364**, 1483–1489. 596

https://doi.org/10.1098/rstb.2009.0027 597 R Core Team. (2019). R: A language and environment for statistical computing. R Foundation for 598 Statistical Computing. https://www.R-project.org/ 599 Réalis-Doyelle E., Pasquet A., de Charleroy D., Fontaine P. & Teletchea F. (2016). Strong effects 600 of temperature on the early life stages of a cold stenothermal fish species, brown trout 601 (Salmo trutta L.). PLoS ONE 11, 1–17. https://doi.org/10.1371/journal.pone.0155487 602 Régnier T., Bolliet V., Gaudin P. & Labonne J. (2013). Bigger is not always better: Egg size 603 influences survival throughout incubation in brown trout (Salmo trutta). Ecology of 604 Freshwater Fish 22, 169–177. https://doi.org/10.1111/eff.12018 605 Rollinson N. & Hutchings J.A. (2013). The relationship between offspring size and fitness 606 integrating theory and empiricism. *Ecology* **94**, 315–324 607 Rollinson N. & Hutchings J.A. (2010). Why does egg size increase with maternal size? Effects of 608 egg size and egg density on offspring phenotypes in Atlantic salmon (Salmo salar). 609 Evolutionary Ecology Research 12, 949–960 610 Rudolf V.H.W. & Roman A. (2018). Trophic structure alters consequences of environmental 611 warming. Oikos 127, 1646–1656. https://doi.org/10.1111/oik.05535 612 Schager E., Peter A. & Burkhardt-Holm P. (2007). Status of young-of-the-year brown trout 613 (Salmo trutta fario) in Swiss streams: Factors influencing YOY trout recruitment. Aquatic 614 Sciences 69, 41–50. https://doi.org/10.1007/s00027-007-0845-x 615 Skoglund H., Einum S. & Robertsen G. (2011). Competitive interactions shape offspring 616 performance in relation to seasonal timing of emergence in Atlantic salmon. Journal of 617 Animal Ecology 80, 365–374. https://doi.org/10.1111/j.1365-2656.2010.01783.x 618 Smith C.C. & Fretwell S.D. (1974). The optimal balance between size and number of offspring. 619 American Naturalist 108, 499–506 620

Sullivan C.J., Vokoun J.C., Helton A.M., Briggs M.A. & Kurylyk B.L. (2021). An

622	ecohydrological typology for thermal refuges in streams and rivers. <i>Ecohydrology</i> 14,
623	e2295. https://doi.org/10.1002/eco.2295
624	Thorn M.W. & Morbey Y.E. (2018). Egg size and the adaptive capacity of early life history traits
625	in Chinook salmon (Oncorhynchus tshawytscha). Evolutionary Applications 11, 205–219.
626	https://doi.org/10.1111/eva.12531
627	Venable L.D. (1992). Size-number trade-offs and the variation of seed size with plant resource
628	status. Source: The American Naturalist 140, 287–304
629	Vøllestad L.A. & Lillehammer T. (2000). Individual variation in early life-history traits in brown
630	trout. Ecology of Freshwater Fish 9, 242–247. https://doi.org/10.1111/j.1600-
631	0633.2000.eff090407.x
632	Ward J. v (1998). Riverine landscapes: biodiversity patterns, disturbance regimes, and aquatic
633	conservation. Biological Conservation 83, 269–278
634	Wilkins L.G.E., Marques da Cunha L., Menin L., Ortiz D., Vocat-Mottier V., Hobil M., et al.
635	(2017). Maternal allocation of carotenoids increases tolerance to bacterial infection in brown
636	trout. Oecologia 185, 351–363. https://doi.org/10.1007/s00442-017-3952-y
637	Yang L.H. & Rudolf V.H.W. (2010). Phenology, ontogeny and the effects of climate change on
638	the timing of species interactions. <i>Ecology Letters</i> <b>13</b> , 1–10. https://doi.org/10.1111/j.1461-
639	0248.2009.01402.x
640	
641	FIGURE LEGENDS
642	Figure 1. Overview of collection sites and variations in traits at emergence along the elevational
643	gradient. (a) Maps of collection sites. Original maps used to describe collection sites were
644	downloaded from <a href="https://d-maps.com/carte.php?num_car=24779&amp;lang=en">https://d-maps.com/carte.php?num_car=24779⟨=en</a> ,

maps.com/carte.php?num car=2251&lang=en. (b) Elevation above sea level (m) of each 647 collection site. (c) The relationship between elevation of collection site and water temperature 648 during the incubation period. (d) The relationships between the elevation of collection site and 649 degree-days until emergence in the cold and warm treatments. (e) The relationships between the 650 elevation of collection site and body length at emergence in the cold and warm treatments. In (d) 651 and (e), the circles filled with white and other colours (black, blue, and red) with error bars 652 denote population mean  $\pm$  SD in the warm and cold treatment, respectively. Solid lines in (d) and 653 (e) are predictions from the final models (Table 1), and shaded areas are 95 % CI drawn using the 654 siPlot package. Note that Y axes are log-scaled in (d) and (e). 655 Figure 2. (a) The relationship between elevation of collection site and maternal body length. (b) 656 The relationship between elevation of collection site and egg volume. In (a) and (b), the filled 657 circles with error bars denote population mean  $\pm$  SD. Solid lines in (a) and (b) are predictions 658 from the final models (Table 2), and shaded areas are 95 % CI drawn using the sjPlot package. 659 Relationships between temperature treatment and residuals of (c) body length at hatching-, (d) 660 degree-days until hatching-, (e) yolk sac volume at hatching-, and (f) oxygen consumption rate at 661 hatching- after regression against egg volume. Error bars denote SD. Note that Y axes are log-662 scaled in (a) and (b). 663 Figure 3. Initial (a) and final path models of the cold (b) and warm (c) treatments. Unidirectional 664 arrows represent supported relationship (positive [blue] and negative [orange]) between variables. 665 The thickness of lines is scaled to the magnitude of standardized correlation coefficients. 666

667

668

669

670

671

### TABLE LEGENDS

**Table 1.** List of explanatory variables and random factors included in the final models (in bold) after backwards models selection using step function (Satterthwaite approximation) and those included in the initial models explaining traits at emergence. Npar is an abbreviation for the

number of parameters in the model. ΔAIC gives the difference in AIC between the final and initial models.

Table 2. List of explanatory variables and random factors included in the final models (in bold) after backwards models selection using step function (Satterthwaite approximation) and those included in the initial models explaining maternal traits and traits at hatching. Npar is an abbreviation for the number of parameters in the model. ΔAIC gives the difference in AIC between the final and initial models.

## **FIGURES**

## Figure 1.

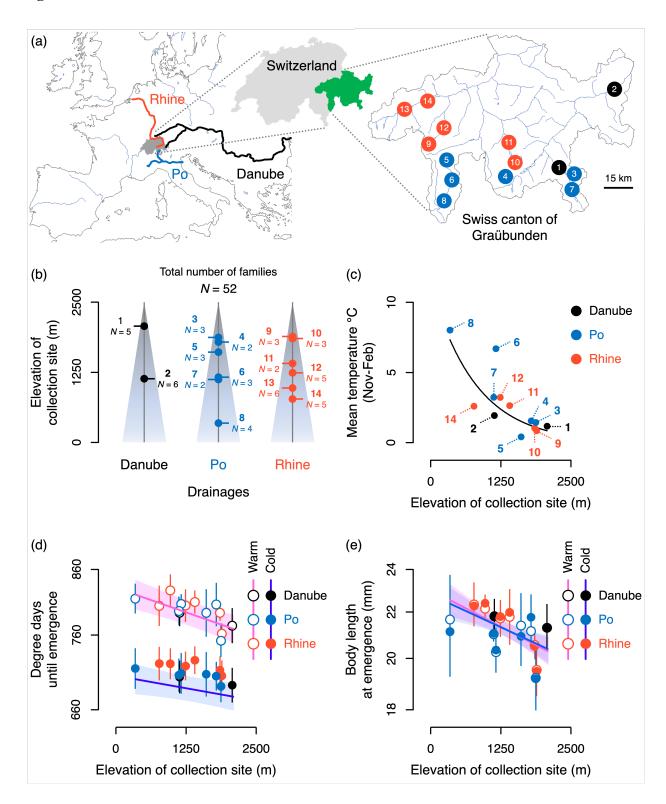
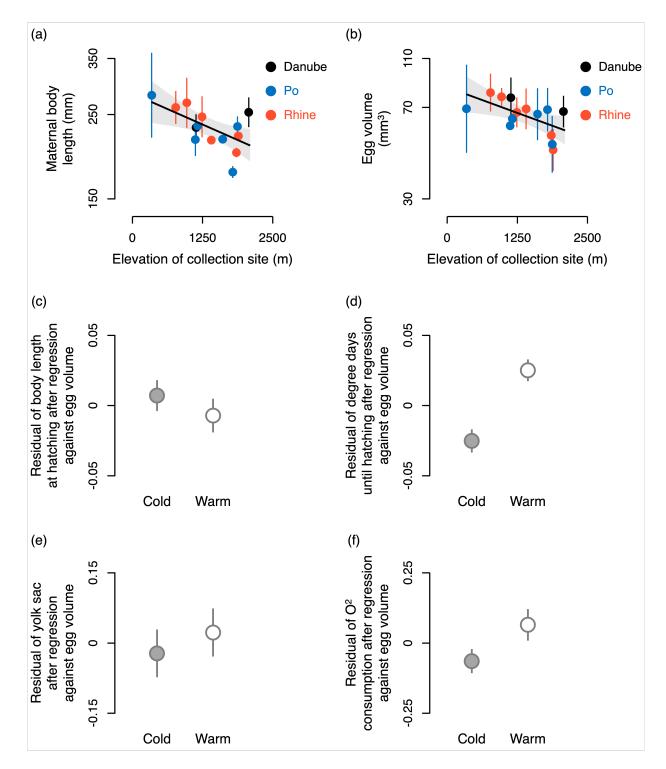
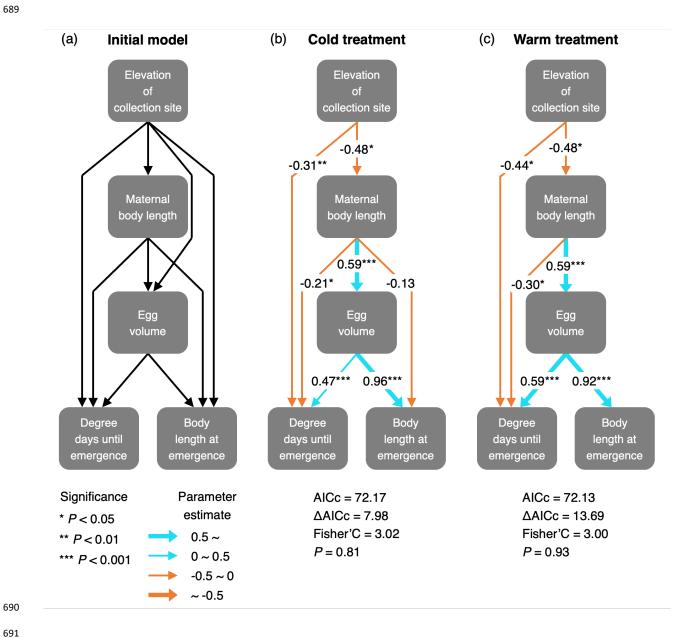


Figure 2.



## Figure 3.



# Table 1

	Npar	AIC	ΔAIC
(1) Degree-days until emergence			
Treatment + Elevation of collection site +	10	-2700.7	0
Treatment: Elevation of collection site +			
(1 site/family) + (1+tray incubator)			
Treatment + Elevation of collection site + Drainage +	14	-2697.7	3.0
Treatment: Elevation of collection site +			
Elevation of collection site: Drainage +			
(1 site/family) + (1+tray incubator)			
(2) Body length at emergence			
Treatment + Elevation of collection site + Drainage +	9	-2679.8	0
Treatment: Elevation of collection site +			
(1  family: site) + (1 incubator)			
Treatment + Elevation of collection site + Drainage +	14	-2675.5	4.3
Treatment: Elevation of collection site +			
Elevation of collection site: Drainage +			
(1 site/family) + (1+tray incubator)			

# Table 2

	Npar	AIC	ΔAIC
(1) Maternal body length			
Elevation of collection site + (1 site:drainage)	4	-314.2	0
Elevation of collection site + (1 drainage/site)	5	-312.2	2.0
(2) Egg volume			
Elevation of collection site + (1 site:drainage)	4	-213.5	0
Elevation of collection site + (1 drainage/site)	5	-211.5	2.0
(3) Degree-days until hatching			
Treatment + (1 site:drainage)	4	-723.7	0
Treatment + Elevation of collection site +	7	-719.2	4.5
Treatment: Elevation of collection site + (1 drainage/site)			
(4) Body length at hatching			
Treatment + (1 site:drainage)	4	-642.8	0
Treatment + Elevation of collection site +	7	-641.5	1.3
Treatment: Elevation of collection site + (1 drainage/site)			
(5) Yolk sac volume at hatching			
Treatment + (1 site:drainage)	4	-365.7	0
Treatment + Elevation of collection site +	7	-365.3	0.4
Treatment: Elevation of collection site + (1 drainage/site)			
(6) Oxygen consumption rate at hatching			
Treatment	3	-324.1	0
Treatment + Elevation of collection site +	7	-317.6	6.5
Treatment: Elevation of collection site + (1 drainage/site)			