Research Article

Benthic diatoms of an Alpine stream/lake network in Switzerland

Christopher T. Robinson^{1,*} and Barbara Kawecka²

¹ Swiss Federal Institute of Aquatic Science and Technology (Eawag), Department of Limnology, Ueberlandstrasse 133, CH-8600 Dübendorf, Switzerland

² Institute of Nature Conservation, Polish Academy of Sciences, Al. A. Mickiewicza 33, PL-31-120 Krakow, Poland; e-mail: kawecka@iop.krakow.pl

Received: 3 November 2004; revised manuscript accepted: 11 March 2005

Abstract. We compared the benthic diatom composition of lakes, and lake inlet and outlet streams in a high elevation catchment (~2600 m a.s.l.) in the Swiss Alps. The catchment consisted of a southern basin mostly fed by glacial-melt water and a northern basin fed by snowmelt and groundwater. Streams in both basins flowed through a series of small lakes before converging into a lake with a primary outlet channel. The south basin had on average 4°C cooler water temperatures and 2× higher nitrate-N levels (up to 300 µg/L) than the north basin. In contrast, the north basin had higher levels (2–4×) of particulate-P, particulate-N, and particulate organic matter than the south basin. A total of 109 and 143 diatom species was identified in lakes and streams, respectively, with a similar number of species found in each basin. *Aulacoseira alpigena* and *Achnanthes minutissimum* were common benthic algae in north basin lakes, whereas *Achnanthes subatomoides*, *Achnanthes marginulata*, *Pinnularia microstauron*, and *Psammothidium helveticum* were most common in south basin lakes. One disconnected lake in the north basin had an assemblage dominated by *Tabellaria flocculosa* (66%) and *Eunotia tenella* (14%). Principal components analysis

showed a clear separation between the north and south basins in lotic diatoms. Of the 10 most common species, streams in the south basin had greater abundances of *Psammothidium helveticum*, *Achnanthes helvetica* var. *minor*, *Achnanthes marginulata*, *Achnanthes subatomoides*, and *Diatoma mesodon* than the north basin, whereas north basin streams had higher abundances of *Achnanthes minutissimum*, *Aulacoseira alpigena*, and *Luticola goeppertiana*. Lake outlet assemblages were similar to respective downstream lake inlet assemblages, and assemblages changed in composition along each basins longitudinal flow path. However, *Aulacoseira alpigena* had higher average abundances in north basin outlets than inlets, and *Achnanthes minutissimum*, *Psammothidium helveticum*, and *Achnanthes helvetica* var. *minor* had higher average abundances in south basin outlets than inlets. In contrast, *Diatoma mesodon*, *Fragilaria capucina*, and *Gomphonema parvulum* had higher average abundances in south basin inlets than outlets. The spatial patterns in species composition reflected the hierarchical interaction of landscape features (geology, hydrology) on longitudinal gradients (lake position) in the stream/lake network.

Key words. Swiss National Park; lake outlet; acidophilus algae; lake order; fragmentation.

* Corresponding author e-mail: robinson@eawag.ch
Published Online First: August 22, 2005

Introduction

Freshwater systems in alpine environments comprise a variety of lotic and lentic waters. Sources of stream discharge range from glacial and snowmelt to groundwater (Ward, 1994) and mixtures of each (Tockner et al., 1997; Brown et al., 2003). Ephemeral and intermittent streams also are quite common in alpine environments (Robinson et al., 2003; Rüegg and Robinson, 2004). Alpine lakes range in size and degree of connectedness; some being isolated water bodies while others are inter-connected by streams forming lake chains. Many alpine lakes have outlet streams (Hieber et al., 2001) that also can be temporary (Donath and Robinson, 2001). The juxtaposition of lakes and streams in alpine catchments enhances overall habitat heterogeneity and potential biodiversity. Likewise, the position of a lake(s) in the stream network has important implications regarding the structure and functioning of downstream receiving waters (Cattaneo, 1996; Soranno et al., 1999; Kling et al., 2000) as well as instream flora and fauna (Schanz, 1984; Hershey et al., 1999; Hieber et al., 2001). Knowledge of the complex linkages in lake/stream networks is needed to better understand the spatial and temporal dynamics in the distribution and abundance of organisms in the context of ecological constructs such as, for example, the River Continuum Concept (Vannote et al., 1980). This issue is particularly important in alpine systems where the present retreat of glaciers can cause the formation of new lakes.

The presence of lakes in a stream network causes fragmentation in the longitudinal continuity of streams. Lake chains, in fact, can have a major influence on longitudinal patterns in physical and chemical properties of streams and among lakes (Soranno et al., 1999; Kling et al., 2000; Quinlan et al., 2003). A clear example of a lake influence for stream systems in lake chains is the resulting formation of lake inlets and lake outlets. Lake outlets, in particular, are well-known for their characteristic benthic assemblages, especially macroinvertebrates (Robinson and Minshall, 1990; Richardson and Mackay, 1991). Algal assemblages, although rarely studied in lake outlets, are quite sensitive to environmental change (McCormick and Cairns, 1994) and would be expected to show distinct shifts in community composition associated with physical/chemical variation in the lake water that feeds them (Kawecka et al., 1971; Cattaneo, 1996; Kawecka and Galas, 2003). An additional consideration in alpine environments is the effect of water source or stream type (*sensu* Brown et al., 2003) on algal assemblages in streams and lakes of different lake chains. For example, how does water source (*i.e.*, chemical and physical differences) influence the development of algal assemblages along a stream/lake network in high elevation alpine environments?

The present study examined diatom assemblages along two lake chains in the same alpine catchment, each having a different water source (glacial and snowmelt). Our objectives were threefold: 1) to test for differences in diatom assemblages in lakes and streams between the two basins; 2) to test for differences between lake inlet and lake outlet diatom assemblages in the two basins; and 3) to examine for longitudinal patterns in diatom assemblages in the two lake chains. We expected that water source would have a significant effect on spatial distribution patterns of diatoms between the two basins.

Study area description

The Macun Lakes region (46°44' N, 10°08' E) is a high alpine cirque (>2600 m a.s.l.) in the Canton Graubünden, Switzerland (Fig. 1). The 3.6 km² region was annexed with the Swiss National Park in 2000, and currently is an area designated for long-term monitoring of Alpine streams and lakes. The region comprises 26 small lakes with the 5 largest lakes interconnected by stream segments, each <500 m in length. The surrounding peaks reach elevations between 2800 and 3000 m a.s.l., and the outlet stream (Zeznina) drains north to the Inn River near the village of Lavin, Switzerland, in the lower Engadine. Water sources in the basin originate from precipitation (mostly as snow in winter) and a number of rock glaciers, especially in the southern part of the basin. Precipitation is low, around 850 mm/y. Air temperatures range from >20°C in summer to <-15°C in winter based on records from the nearby climate station (Buffalora) ~14 km southeast of Macun. Bedrock geology is slow-weathering crystalline (ortho-gneiss) rock (Bortolami, 1998). The area is above treeline, thus most terrestrial vegetation is typical Alpine grasses and low lying herbs (Körner, 2001); areas of bare rock are common.

The drainage network of Macun is represented by a northern and southern basin (Fig. 1). The north basin is primarily precipitation (mostly as snow) and groundwater fed, whereas the south basin is fed mostly by glacial melt from rock glaciers. Consequently, the annual flow regime differs between basins with the south basin experiencing more extreme channel contraction from the freezing of glacial water in autumn. The stream network, as a whole, contracts by up to 60% in winter (S. Matthaei, unpublished data). The water source in each basin also causes differences in water temperature, being warmer in the north basin, and water chemistry (as described below). We selected 10 stream sites and 7 lakes for assessment of diatom assemblage structure (Fig. 1). Four stream sites were located in the north basin, 4 sites in the south basin, and 2 sites in the outlet stream (called outlet basin in the remaining text). The sites were situated at the inlets and outlets of the prominent lakes in

Figure 1. Schematic map of the Macun lakes region in eastern Switzerland. Numbers show the sample locations for diatoms and correspond to site numbers in Table 1 and the appendix. Lake names correspond to lakes sampled in late August 2003. The Macun catchment can be separated into two distinct hydrologic basins (north and south) and an outlet basin that drains both upper basins into the Inn River in the lower Engadine valley. Lake Dragun is hydrologically disconnected from the other lakes in the north basin with a separate outlet stream. The sites are situated between 2600 and 2670 m a.s.l.

each basin along a longitudinal gradient. Melt-water from a large rock glacier enters the outlet stream between stream sites 9 and 10. Of the sampled lakes, Dragun is disconnected from the other lakes in the north basin with a separate outlet channel that goes dry in summer. Lake Immez is located at the confluence of the north and south drainage channels.

Methods

Water samples were collected monthly (June to September) from each stream site from September 2001 to September 2003 ($n = 9$). Lake chemistry was assumed to be similar to respective lake outlet samples. Samples were taken to the laboratory, filtered (Whatman GF/F), and analyzed for constituents of nitrogen (NO_2 , $\text{NO}_2 + \text{NO}_3$, PN), phosphorus ($\text{PO}_4\text{-P}$, PP), and silica (SiO_2) using methods detailed in Tockner et al. (1997). Measures of total inorganic (TIC) and particulate organic carbon (POC) were derived from the filters using standard methods. On each visit, we measured water temperature and conductivity (WTW LF 323, Germany), turbidity (Cosmos, Züllig AG, Switzerland), and pH (WTW pH 330, Germany) using portable field meters. Turbidity was expressed in nephelometric turbidity units (NTU).

Diatoms were sampled from each stream site on 7 September 2001, 16 July 2002 and 18 September 2002. Diatoms were sampled from each lake on 22 August 2003. From each stream site, five rocks were randomly collected along a 10 m reach. From each lake, 15–20 rocks were collected from the littoral zone around each lake. The periphyton layer on the surface of each rock was scrubbed with a brass brush and the slurry rinsed into a plastic container. A subsample (~ 4 ml) from each slurry was then combined in a 20 ml plastic vial, preserved with 2% formalin, and taken to the laboratory. In the laboratory, each sample was macerated in a chromic acid cleaning solution (3:1 v/v of sulphuric acid and potassium dichromate) for 1–2 days, and then washed by centrifugation. Diatoms from each sample were embedded permanently on glass slides with the synthetic resin Pleurax, having a reflective index of 1.75. Slides for diatoms were examined using a Nikon microscope (Eclipse 600) and, when necessary, by scanning electron microscopy (SEM; JEOL JSM 35 CF). A minimum of 250 diatom valves were identified and counted along random transects from each slide. Identifications were based on Krammer and Lange-Bertalot (1986; 1988; 1991a,b), Flower and Jones (1989), Round et al. (1990), Krammer (1992), Lange-Bertalot and Moser (1994), and Lange-Bertalot and Metzeltin (1996). A diatom species list with authorship is provided in Appendix A.

Table 1. Average (SD) physical-chemical characteristics of the study sites and for streams in each of the three basins. Site numbers correspond to numbers in figure 1. N = 9 from samples collected monthly from September 2001 to September 2003, excluding winter (October to May).

Site	Num- ber	Type	Eleva- tion (m)	Con- ductivity ($\mu\text{S}/\text{cm}$)	Turbidity (NTU)	Tem- perature ($^{\circ}\text{C}$)	pH	$\text{NO}_2\text{-N}$ ($\mu\text{g}/\text{l}$)	NO_2+NO_3 ($\mu\text{g}/\text{l}$)	PN ($\mu\text{g}/\text{l}$)	$\text{PO}_4\text{-P}$ ($\mu\text{g}/\text{l}$)	PP ($\mu\text{g}/\text{l}$)	TIC (mg/l)	POC (mg/l)	SiO_2 (mg/l)
Grond	1	outlet	2637	6.7 (0.5)	1.3 (0.7)	10.7 (3.0)	7.4 (0.4)	0.7 (0.5)	<100	94 (56)	<5	9.1 (3.5)	0.7 (0.2)	1.2 (0.8)	2.2 (0.2)
Mezza-glüna	2	inlet	2634	6.7 (0.3)	1.4 (0.6)	11.4 (3.9)	6.8 (0.3)	0.6 (0.5)	<100	104 (69)	<5	8.9 (4.4)	0.5 (0.2)	1.0 (0.6)	2.3 (0.2)
Mezza-glüna	3	outlet	2624	5.7 (0.4)	1.8 (0.8)	12.4 (4.2)	6.7 (0.4)	0.7 (0.5)	<100	102 (44)	<5	9.6 (4.3)	0.5 (0.2)	1.4 (0.7)	1.8 (0.2)
Immez-north	4	inlet	2620	6.2 (0.5)	1.5 (0.6)	13.4 (4.5)	6.6 (0.3)	1.0 (0.0)	<100	113 (46)	<5	10.7 (3.8)	0.6 (0.3)	1.2 (0.6)	1.8 (0.3)
Sura	5	outlet	2668	8.9 (4.5)	0.9 (0.4)	7.3 (3.0)	6.7 (0.7)	2.0 (1.3)	301 (82)	25 (17)	<5	3.2 (3.0)	0.3 (0.1)	0.2 (0.1)	2.1 (0.5)
Sura	6	inlet	2650	11.2 (6.0)	0.7 (0.5)	7.4 (3.4)	6.4 (0.3)	0.7 (0.8)	246 (69)	13 (4)	<5	2.3 (1.3)	0.4 (0.0)	0.2 (0.1)	2.2 (0.6)
Sura	7	outlet	2631	11.0 (5.0)	1.5 (0.9)	7.9 (3.1)	6.9 (0.6)	1.6 (0.7)	169 (61)	33 (11)	<5	4.4 (1.8)	0.4 (0.1)	0.3 (0.2)	2.1 (0.5)
Immez-south	8	inlet	2620	14.0 (9.8)	1.5 (0.8)	11.6 (3.5)	6.9 (0.5)	1.6 (0.7)	200 (90)	33 (11)	<5	4.7 (1.8)	0.7 (0.2)	0.3 (0.2)	2.1 (0.5)
Immez	9	outlet	2616	9.1 (1.7)	1.3 (0.4)	9.0 (3.0)	6.8 (0.4)	1.1 (0.4)	138 (34)	49 (22)	<5	9.7 (14.5)	0.5 (0.2)	0.4 (0.2)	2.1 (0.5)
Zeznina	10	basin outlet	2552	26.9 (21.7)	4.4 (3.3)	6.3 (2.1)	7.2 (0.1)	0.9 (0.7)	241 (78)	29 (17)	<5	5.8 (3.0)	0.8 (0.3)	0.3 (0.1)	2.5 (0.6)
North basin	1-4	all	2630	6.3	1.5	12.0	6.9	0.8	<100	103	<5	9.6	0.6	1.2	2.0
South basin	5-8	all	2642	11.3	1.2	8.6	6.7	1.5	229	26	<5	3.7	0.4	0.3	2.1
Outlet basin	9-10	all	2584	18.0	2.9	7.7	7.0	1.0	190	39	<5	9.7	0.7	0.4	2.3

Data analysis

Physical and chemical measures collected from each stream site were summarized as means and standard deviations (Table 1). Stream sites also were grouped and summarized by respective study basin, resulting in 4 sites in the north basin, 4 sites in the south basin, and 2 sites in the outlet basin (Fig. 1). Lake diatom samples were compared between basins using a paired t-test, excluding the lakes Dragun and Immez. For stream diatoms, two-way ANOVA was used to test for differences in species richness between inlets and outlets among dates, and among basins and dates (Zar, 1984). Two-way ANOVA also was used to test for differences between inlets/outlets and basins for the 10 most abundant species after arcsine square-root transformation. In addition, principal components analysis (PCA, varimax rotation) was used to illustrate assemblage differences among basins, and between inlets and outlets based on the relative abundances (arcsine square-root transformed) of the 10 most abundant species. These PCA's also were used to indicate any longitudinal or gradient patterns in the lotic diatom assemblages from the upper most sites to the most downstream site (Zeznina).

Results

Physical and chemical characteristics between basins

The north and south basin differed considerably in physical and chemical characteristics of the water, clearly reflecting the source of water in each basin (Table 1). Although overall conductivity was generally low because of bedrock geology, the south basin had two-fold higher conductivity values than the north basin. The input from the rock glacier below site 9 also caused another two-fold increase in conductivity at site 10 (Zeznina). Water clarity was high, being <2 NTU at all sites except site 10. Water pH ranged between 6.4 and 7.4 for all sites. Water temperatures were around 4 $^{\circ}\text{C}$ cooler in the south basin because of the input of glacier water. Temperatures typically increased along the longitudinal gradient within each basin, increasing from 10.7 to 13.4 $^{\circ}\text{C}$ in the north basin and from 7.3 to 11.6 $^{\circ}\text{C}$ in the south basin (Table 1). Dissolved nitrogen constituents were around twice as high in the south basin than the north basin with highest values at site 5, averaging >300 $\mu\text{g}/\text{L}$ ($\text{NO}_2+\text{NO}_3\text{-N}$). In contrast, particulate N was 3 $\times$ greater in the north basin than the south basin and outlet basin. Phosphate-P ($\text{PO}_4\text{-P}$) was always below analytical detection limits at all sites, whereas particulate-P was about two-fold higher in the north basin than the south basin. TIC was generally low, usually <0.8 mg/L , and silica was sufficient for algal growth. Lastly, POC was 4 $\times$ higher in the north than the south and outlet basins.

Figure 2. Average relative abundance (as %, +1 SE) of the top 8 most abundant diatom species in the major lakes of Macun with lakes from each basin combined (north basin, $n = 3$; south basin, $n = 2$) and Immez and Dragun presented as separate lakes. List of diatom species found in the lakes is given in Appendix A.

Between-basin differences in lentic diatoms

A total of 109 diatom species were identified from the lakes. Of the more common species, north basin lakes had higher abundances of *Aulacoseira alpigena*, *Achnantheidium minutissimum* and *Aulacoseira distans* than the south basin (t-test, $p < 0.05$) and other lakes (Fig. 2, App. A). *Achnanthes subatomoides*, *Achnanthes marginulata*, *Pinnularia microstauron*, and *Psammothidium helveticum* were more common in the south basin lakes and lake Immez than in north basin lakes (t test, $p < 0.05$). Lake Dragun had an assemblage dominated by *Tabellaria flocculosa* (66%) and *Eunotia tenella* (14%) (Fig. 2). Lastly, *Pinnularia sinistra* (~2%) was common in most lakes, and *Fragilaria capucina* was found common (13%) in Mezza Glüna (data not shown).

Between-basin differences in lotic diatoms

A total of 143 diatom species were identified from the stream samples, 109 species in the north basin, 108 in the

Figure 3. Average (+1 SE) diatom species richness in streams in each basin on each collection date and for all dates combined ($n = 3$). North basin, $n = 4$ sites; South basin, $n = 4$ sites; Outlet basin, $n = 2$ sites.

Figure 4. Scatterplot of the PCA factor scores for the 10 stream sites using the top 10 most abundant diatoms (as %) with all dates combined. Circles are north basin sites, triangles are south basin sites, and diamonds are outlet basin sites. Numbers inside symbols refer to site numbers from Figure 1. Some dates were not included for some sites in the analysis due to low numbers of diatoms in those samples (see methods). The first two axes explained 67% of the variation among sites. PSHE = *Psammothidium helveticum*, ACHM = *Achnanthes helvetica* var. *minor*, ACMA = *Achnanthes marginulata*, ACSU = *Achnanthes subatomoides*, GOPA = *Gomphonema parvulum*, PISI = *Pinnularia sinistra*, and FRCA = *Fragilaria capucina*. Species were related positively with each respective axis.

south, and 71 in the outlet basin (App. A). There was no significant difference in the average species richness of diatoms among basins ($p = 0.61$), although average richness was lowest in the outlet basin in the September samples (Fig. 3). ANOVA results indicated a significant difference in diatom richness among dates ($p = 0.005$),

Figure 5. Average relative abundance (as %, +1 SE) of the top 10 most abundant diatom species in the inlets (IL) and outlets (OL) in each basin with all dates combined ($n = 3$). In the outlet basin, IM is site 9 (Immez outlet) and ZE is site 10 (Zeznina).

being lowest in September 2001 and highest in July 2002. Average species richness was around 30 when all dates were combined, but ranged from <20 to ~ 40 .

The PCA results showed a distinct difference in diatom assemblage structure between the different basins based on the 10 most common species (Fig. 4). The south basin had greater abundances of *Psammothidium hel-*

veticum, *Achnanthes helvetica* var. *minor*, *Achnanthes marginulata*, and *Achnanthes subatomoides* than the north and outlet basins, as depicted along axis-1 of the PCA scatterplot. Axis-1 factor scores of the PCA were significantly correlated with particulate organic carbon ($r = -0.64$), particulate phosphorus ($r = -0.74$), particulate nitrogen ($r = -0.63$), and nitrate-N ($r = 0.64$). A longitudi-

Figure 6. Average (± 1 SE) diatom species richness in combined inlets, outlets, and Zeznina (site 10) for each date and for all dates combined ($n = 3$). Inlets, $n = 4$; Outlets, $n = 5$; Zeznina, $n = 1$.

Figure 7. Scatterplot of PCA factor scores for 9 stream sites based on the top 10 most abundant diatom species (as %) from data collected in July 2002. Numbers represent site numbers in Figure 1 and Table 1. PCA was used to illustrate patterns between inlet and outlets in each basin and whether a longitudinal gradient in the diatom assemblage was present. The first two axes explained 59% of the variation among sites. DIHM = *Diatoma mesodon*, PSHE = *Psammothidium helveticum*, ACHM = *Achnanthes helvetica* var. *minor*, AUAL = *Aulacoseira alpigena*, GOPA = *Gomphonema parvulum*, TAFL = *Tabellaria flocculosa*, ENMI = *Encyonema minutum*. A minus sign before a diatom species in the figure indicates a negative relationship along the axis. Circles represent outlet sites and squares inlet sites. Large ellipses simply group similar sites in respect to diatoms.

nal shift in diatoms was evident along PCA axis-2, with *Fragilaria capucina*, *Gomphonema parvulum*, and *Pinnularia sinistra* increasing in abundance downstream. The outlet basin site (site 9) was placed intermediate to the north and south basins in the PCA, whereas site 10 (Zeznina) showed assemblage structure more similar to the north basin sites (Fig. 4).

Significant basin differences were found for 6 of the 10 most common species identified (2-way ANOVA, $p < 0.05$). *Aulacoseira alpigena* had greater abundances in the north basin, and those more common in the south basin were *Diatoma mesodon*, *Psammothidium helveticum*, *Achnanthes helvetica* var. *minor*, *Achnanthes marginulata*, and *Achnanthes subatomoides* (Fig. 5). Within the top 10 most abundant species common to both basins were *Achnanthidium minutissimum*, *Gomphonema parvulum*, *Fragilaria capucina* and *Pinnularia sinistra*. Three species dominated the lower outlet basin stream (site 10, Zeznina): *Diatoma mesodon*, *Gomphonema parvulum*, and *Achnanthes subatomoides* (Fig. 5).

Differences between inlets and outlets

There was no significant difference in the species richness of diatoms between inlets and outlets ($p = 0.43$), although site 10 (Zeznina, $n = 1$ thus not included in the ANOVA) clearly had fewer species of diatoms than the inlets and outlets on all sample dates (Fig. 6). There was a significant date effect ($p = 0.019$) with September 2001 having lower average richness than July and September 2002. Average richness in the inlets and outlets ranged from ~20 to 30 with all dates combined, and from ~20 to 40 among dates excluding site 10. The lowest species richness was found at site 10 with 14 species identified.

PCA results showed that adjacent inlet and outlet assemblages (i.e., outlet of upstream lake with inlet to next downstream lake) were quite similar, being plotted near each other on the PCA scatterplot (Fig. 7). This pattern was particularly evident in south basin streams. Site 3 (Mezza Glüna outlet) in the north was similar to site 1 (Grond outlet), and site 9 (Immez outlet) was similar to site 4 (Immez inlet north). Site 2 (Mezza Glüna inlet) could not be used in the PCA analysis because of an extremely depauperate sample of diatoms, although a similar taxonomic composition was evident for the species identified. Axis-1 explained 39% of the variation in the data and primarily separated sites between the two basins (south basin on the right and north basin on the left side of the plot). Species contributing to axis-1 included *Diatoma mesodon*, *Psammothidium helveticum*, *Achnanthes helvetica* var. *minor*, and *Aulacoseira alpigena*. Axis-2 explained an additional 20% of the variation among sites and highlighted the longitudinal gradient of species abundances in the two basins (see below). Species associated with axis-2 included *Gomphonema parvulum*, *Tabellaria flocculosa*, and *Encyonema minutum*. In this analysis, site 10 (Zeznina) had a distinct assemblage of diatoms from the other sites.

Only one of the 10 most abundant species showed a significantly higher abundance in outlet streams than in inlet streams, this being *Aulacoseira alpigena* in the north basin (Fig. 4). High variances precluded additional signif-

icant differences between outlets and inlets in each basin, but *Achnanthydium minutissimum*, *Psammothidium helveticum* and *Achnanthes helvetica* var. *minor* had higher average abundances in the south basin outlets than inlets. In contrast, *Diatoma mesodon*, *Fragilaria capucina*, and *Gomphonema parvulum* had higher average abundances in south basin inlets than outlets.

Longitudinal gradients in lotic diatom assemblages

PCA, using only the July 2002 data to restrict seasonality, revealed a longitudinal gradient in species composition in the south basin with sites 5 and 6 being placed in the upper right section of the PCA scatterplot, sites 7 and 8 in the upper central section of the plot, site 9 in the lower central section, and then site 10 in the lower right section of the plot (Fig. 7, also see Fig. 4). Further, sites 1 and 3 in the north basin were situated in upper left corner of the scatterplot, site 4 was near site 9 in the bottom central section, and site 10 again in the lower right of the scatterplot.

Discussion

General features of the diatom assemblages in the catchment

Many of the identified 170 species inhabiting the streams and lakes in the Macun catchment are species ubiquitous in high elevation streams in the Alps (Kawecka, 1981; Kawecka and Galas, 2003). A similar number of species (>100) were identified in each basin, although substantially fewer species were found in the outlet basin streams (see Appendix A). Fewer species were found in lake samples (~110) than in stream samples (~150), and 76 species were found in both kinds of habitats. Species assemblages contained a few (~15–20) common species and a high number of rare species. The lower-most stream site had the fewest number of species in the catchment with only 14 species identified in the September 2001 samples. In streams, on average more species were identified in July samples than in samples from either September collection. Average diatom richness across dates ranged from around 20 to 40 among the different sites. These values lie in the range of other studies on high elevation streams in the Swiss Alps (Wuthrich, 1975; Niederhauser, 1993) and throughout Europe (Kawecka, 1980; Tolotti, 2001; Kawecka and Galas, 2003; Negro et al., 2003).

Acidophilus species of *Achnanthes* (e.g., *A. helvetica* var. *minor*, *A. subatomoides* and *A. marginulata*) and *Aulacoseira alpigena* were found in most samples in lakes and streams (following Kawecka, 1981; Kawecka and Galas, 2003). However, more circumneutral species also were found and in relatively high abundances, e.g., *Diatoma mesodon*, *Fragilaria capucina*, *Achnanthydium*

minutissimum, and *Psammothidium helveticum*. However, both *A. subatomoides* (south basin) and *A. alpigena* (north basin) were still quite common in the lakes, indicating acidophilus conditions are still present. The higher abundance of *A. minutissimum* in north basin lakes suggests basin differences in buffering capacities to acidification, and perhaps the effects of different water sources in each basin on diatom composition (discussed below).

Between-basin differences in lotic and lentic diatoms

We found clear differences in stream diatoms between the north and south basin that reflected basin differences in water chemistry. For example, there was significant correlations of the PCA axis-1 scores with particulates (N, P, and organic C), and basins were separated along this axis. There were also apparent differences in stream assemblages between the two basins and the outlet basin streams (sites 9 and 10). Outlet basin streams (sites 9 and 10) were dominated by *Diatoma mesodon*, *Achnanthydium minutissimum*, and *Gomphonema parvulum*. Site 10 was lowest in elevation and had the highest conductivity of all the sites. This site also was more characteristically stream-like than sites in the north and south basin where sites were either lake inlets or lake outlets. Interestingly, the PCA also showed site 10 to be quite similar to north basin sites. North basin stream sites were dominated by *A. minutissimum*, *G. parvulum*, *Aulacoseira alpigena*, and to a lesser extent *Achnanthes subatomoides*. As mentioned, most of these species are common in waters with circumneutral pH (Kawecka, 1981), and the north basin was snowmelt and groundwater-fed. In contrast, south basin streams were dominated by some acidophilus *Achnanthes* species, but also circumneutral *Fragilaria capucina*, and alkalophilus *Psammothidium helveticum*. The south basin was mostly fed by glacial meltwater from rock glaciers and tended to have lower conductivities and pH than the north basin.

Basin differences also were found for lake diatoms. Here, north basin lakes had higher abundances of *Aulacoseira alpigena*, *Aulacoseira distans* and *Fragilaria capucina* than south basin lakes, whereas south basin lakes were dominated by *Achnanthes subatomoides*, *Achnanthes marginulata*, *Achnanthes helvetica* var. *minor*, and *Psammothidium helveticum*. *Pinnularia* spp. (*P. microstauran* and *P. sinistra*) also seemed more common in south basin lake samples than north basin lakes. Lake Dragun had an assemblage completely dominated by *Tabelaria flocculosa* (66%), and is isolated from other lakes in the catchment. Last, the confluence lake (Lake Immez) assemblage was more similar to assemblages in south basin lakes than north basin lakes, perhaps reflecting the greater amount of its water being derived from the south system (S. Matthaei, unpublished data).

Diatom differences between lake inlets and lake outlets

PCA revealed that lake outlet assemblages were quite similar to adjacent downstream inlet assemblages. This finding may simply be caused by the close proximity of adjacent outlet and inlet locations, as most outlets were within ~100–200 m of a downstream inlet. However, a number of species differed between outlet and inlet assemblages when respective stream types were combined, suggesting a strong influence of the lake on outlet assemblage structure. For example, *Gomphonema parvulum* and *Fragilaria capucina* were more abundant in inlets than outlets in both basins. In contrast, *Aulacoseira alpigena* was more common in outlets than inlets in the north basin, and *A. alpigena* also was common in north basin lakes. In the south basin, *Achnanthes subatomoides*, *Achnanthes marginulata* and *Psammothidium helveticum* were on average more common in outlets than inlets, and both species were found in south basin lakes. Lakes are well-known to influence habitat characteristics and macrozoobenthos of outlet streams (Robinson and Minshall, 1990; Richardson and Mackay, 1991; Wotton, 1995). Further, zooplankton abundances are typically high in outlet streams, declining exponentially downstream (Robinson and Minshall, 1990), and many outlet streams contain high abundances of filter feeders that consume high quality seston coming from the lake (Wotton, 1979). Our data suggest that diatoms in the respective lakes may colonize or be a continuous source of colonists to outlet streams, thereby influencing observed assemblage differences between inlets and outlets in each basin.

Longitudinal patterns along lake chains

Both basins appeared to have a lake chain influence on the longitudinal gradient in diatom assemblage structure. For example, the abundances of *Gomphonema parvulum*, *Fragilaria capucina*, and *Pinnularia sinistra* increased, and *Encyonema minutum* decreased, along the longitudinal gradient in each basin. Physical and chemical characteristics of streams and lakes have been found to change longitudinally along lake chains (Soranno et al., 1999; Kling et al., 2000). In the Macun lake chains, streams experienced increased temperatures and increased levels of particulate-N and -P along each lake chain. We suspect in-lake processing, as found by Kling et al. (2000), also occurred in the Macun lakes, thereby influencing downstream changes in physical and chemical properties of streams. These longitudinal changes in environmental conditions probably affected the diatom assemblage structure, as shown in many studies examining diatom-environment relationships (e.g., Moss, 1973; Chessman, 1986; Passy, 2001; Potapova and Charles, 2003; Rott et al., 2003). In closing, our results demonstrated a lake chain effect on diatom assemblages colonizing the inlets and

outlets of lakes in this high elevation Alpine catchment. However, water source, by influencing instream physical and chemical properties, resulted in major differences in assemblage composition in the lakes and streams of each basin. These findings suggest that lake chains interact with landscape features in a hierarchical sense in dictating the distribution patterns of diatom assemblages in high alpine stream/lake networks.

Acknowledgments

We gratefully thank M. Hieber, C. Jolidon, J. B. Logue, S. Matthaei, J. Rüegg, L. Shama, and V. Wenzelides for assistance in the field collections of algae over the period of study. We thank R. Illi for the chemical analyses of water samples, and Prof. H. Lange-Bertalot for verification of diatom species. We appreciate the logistic support and introduction of the Macun Lakes by T. Scheurer and F. Filli from the Swiss National Park, and the community of Lavin for allowing road access to the Heidi Hut. Two anonymous reviewers provided constructive comments that improved the paper. The senior author expresses his sincere appreciation to Prof. G. Wayne Minshall for many years of guidance and friendship.

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Appendix A. Complete list of all benthic diatoms identified from the stream and lakes with relative abundances averaged across the three dates (streams) or lakes in a basin. Stream site number corresponds to numbers on map (see Fig. 1). Abundances may not sum to 100 because of depauperate samples from some sites and dates.

Site name	Grond	M-Gluna IL	M-Gluna OL	Immez IL	Sura OB	Sura IL	Sura OL	Immez IL	Immez OL	Zeznina	All	All	Immez	Dragun
Site number	1	2	3	4	5	6	7	8	9	10	NA	NA	NA	NA
Basin	North	North	North	North	South	South	South	South	Outlet	Outlet	South	North	Immez	Dragun
Stream type	Outlet	Inlet	Outlet	Inlet	Outlet	Inlet	Outlet	Inlet	Outlet	Stream	Lake	Lake	Lake	Lake
<i>Achnanthidium biasolettianum</i> (Grun.) L.-Bertalot	0.2	0.3	0.1	0.2	0.7	0.2	0.4	0.1	0.3	0.3		0.4	0.4	
<i>A. minutissimum</i> (Kütz.) Czarnecki	25.9	18.3	2.1	30.7	3.2	1.6	21.1	0.6	35.0	4.2	5.6	15.0	15.2	
<i>Achnanthes bioretii</i> Germain											1.3	0.3	1.7	
<i>Achnanthes curtissima</i> Carter	0.1	0.1				0.1	0.3		0.1	0.1	0.4	2.2	3.9	
<i>A. daonensis</i> L.-Bertalot											0.2	0.2		
<i>A. helvetica</i> var. <i>minor</i> Flower & Jones	0.1	0.5	0.0	1.0	24.7	14.3	4.0	2.0	1.4	1.1	4.5	1.5	2.2	
<i>A. lanceolata</i> (Bréb.) Grun.												0.3		
<i>A. laterostrata</i> Hust.								0.1						
<i>A. levanderi</i> Hust.											0.2			
<i>A. marginulata</i> Grun.	0.1	0.1		1.1	10.7	6.6	5.1	4.5	2.1		13.0	4.7	10.4	3.2
<i>A. subatomoides</i> L.-Bertalot & Archibald	0.4	1.3	1.9	5.3	14.4	11.3	11.7	4.1	3.2	7.7	26.7	6.4	17.7	
<i>A. scotica</i> Flower & Jones	0.1		0.1			0.4	0.4		0.2					
<i>A. valida</i> (Grun.) Krammer												0.6		
<i>Achnanthes</i> sp.				0.5			0.1				1.1	0.4		0.7
<i>Amphora ovalis</i> (Kütz) Kütz										0.1				
<i>A. pediculus</i> (Kütz.) Grun.					0.1		0.1			0.1				
<i>Anomoeonis brachysira</i> (Bréb.) Grun.				0.5										
<i>Asterionella</i> sp.				0.1										
<i>Aulacoseira alpigena</i> (Grun.) Krammer	9.6	0.8	6.1	6.6		0.2	0.3	0.1	1.9	0.5	3.0	17.7	1.7	
<i>A. distans</i> (Ehr.) Simonsen	1.3	0.1	0.7	0.5					0.1		0.2	5.1	0.4	1.8
<i>A. distans</i> var. <i>nivalis</i> (W. Smith) Haworth	0.2		0.5	0.1							0.2	1.3	0.4	
<i>A. italica</i> (Ehr.) Simonsen	0.2													
<i>A. pfaffiana</i> (Reinsch) Krammer													0.4	0.7
<i>Aulacoseira</i> sp.	0.1		0.1					0.1	0.1					
<i>Brachysira brebissonii</i> L.-Bertalot & Moser											0.4	0.2		
<i>Caloneis aerophila</i> Bock													0.4	
<i>Caloneis bacillum</i> (Grun.) Cl.	0.3			0.1		0.1	0.2	0.1				0.6	0.4	
<i>Caloneis</i> sp.											0.4	0.2	0.4	
<i>C. molaris</i> (Grun.) Krammer								0.1						
<i>C. silicula</i> (Ehr.) Cl.						0.1		0.1						
<i>Chamaepinnularia soehrensii</i> var. <i>hassiac</i> (Krasske) L.-Bertalot								0.1						
<i>Cocconeis pediculus</i> Ehr.									0.1		0.4			
<i>Cocconeis placentula</i> var. <i>euglypta</i> Ehr.	0.1	0.1	0.7	0.4			0.1		0.1	0.5	0.2	0.1	0.9	
<i>Craticula submolesta</i> (Hust.) L.-Bertalot			0.2		0.1	0.1	0.3							
<i>Cyclotella meneghiniana</i> Kütz.			0.1						0.1	0.1				
<i>Cyclotella</i> sp.										0.1				
<i>Cyclotella</i> sp. (small)				0.1										
<i>Cymbella affinis</i> Kütz.									0.8	0.1	0.2			
<i>C. descripta</i> (Hust.) Krammer & L.-Bertalot								0.1						
<i>C. gaeumannii</i> Meister	0.2			0.1		0.1	0.9	0.6			0.2	0.1		

<i>C. gracilis</i> (Ehr.) Kütz.	0.1		0.8		0.6				0.3		0.2	0.6	0.4	
<i>C. hebridica</i> (Grun.) Cl.					0.5	0.1	0.1	0.1			0.4	0.3	0.4	0.7
<i>C. helvetica</i> Kütz.										0.1				
<i>C. mesiana</i> Cholnoky												0.1		
<i>C. microcephala</i> Grun.												0.1		
<i>C. naviculiformis</i> (Auerswald) Cl.	0.2		0.1									0.1		
<i>C. perpusilla</i> Cleve-Euler											0.2	0.2	0.4	
<i>C. sinuata</i> Greg.												0.2		
<i>C. subaequalis</i> Grun.	0.1		0.1											
<i>Cymbella</i> sp.												0.1		
<i>Denticula crassula</i> Kütz.			0.1											
<i>Diademesis contenta</i> (Grun.) D.G. Mann			0.1					0.1						
<i>D. gallica</i> var. <i>perpusilla</i> (Grun.) L.-Bertalot							0.2	0.5	0.5					
<i>Diatoma ehrenbergii</i> Kütz.		0.1												0.4
<i>D. hyemalis</i> (Roth) Heiberg								0.1			1.9	0.3	2.2	
<i>D. mesodon</i> (Ehr.) K tz.	0.3	0.2	0.6	0.4	4.9	24.4	9.3	5.1	4.0	29.3				
<i>D. tenuis</i> Agardh.		0.1									0.2			
<i>D. vulgaris</i> Bory												0.1		
<i>Didymosphenia geminata</i> (Lyngbye) M. Schmidt		0.1				0.2		0.1	1.7					0.4
<i>Encyonema minutum</i> (Hilse) D.G. Mann	0.3	0.6	0.4	0.2	1.5	1.0	0.8	0.5	2.1	1.0	0.4	0.5	1.3	
<i>E. silesiacum</i> (Bleisch) D.G. Mann							0.1	0.1						
<i>Eolimna minima</i> (Grun.) L.-Bertalot		0.1												
<i>E. subminuscula</i> (Manguin) Moser, L.-Bertalot & Metzeltin	0.2	0.1												
<i>Eunotia arcus</i> Ehr.	0.4													
<i>E. bilunaris</i> (Ehr.) Mills		0.1	0.2	1.5	0.4	0.8	0.3	0.1	1.5		0.7	0.2		2.1
<i>E. bilunaris</i> var. <i>mucophila</i> L.-Bertalot & Nörpel											0.2			
<i>E. circumborealis</i> L.-Bertalot & Nörpel						0.1					0.2			
<i>E. diodon</i> Ehr.				0.1	0.5						0.4			
<i>E. diodon</i> Ehr. (sensu lato)					0.9	0.8	0.1		0.1					
<i>E. exiqua</i> (Bréb.) Rabenh.	0.3		0.2	0.5	1.2	1.7	0.5	0.4	0.4	0.3	0.5	0.4	0.9	0.4
<i>E. fallax</i> A. Cl.			0.2	1.5		0.2					0.4	0.3		
<i>E. glacialis</i> Meister												0.2	0.4	0.4
<i>E. implicata</i> Nörpel et al.										0.1				
<i>E. minor</i> (Kütz.) Grun.												0.1	0.4	
<i>E. pectinalis</i> (Dillwyn) Rabenh.					0.1									
<i>E. praerupta</i> Ehr.	0.1			0.1	0.1	0.7		0.1	0.1		0.4	0.1	0.4	
<i>E. praerupta</i> sensu lato	0.4		0.7	0.1	0.4	0.2	0.3	0.1	0.1	0.1	1.8	0.3	0.4	
<i>E. praerupta</i> var. <i>bigibba</i> (Kütz.) Grun.					0.1									
<i>E. rhomboidea</i> Hust.												0.5		1.1
<i>E. subarcuatoidea</i> Alles, Nörpel & L.-Bertalot											0.5	0.7		
<i>E. tenella</i> (Grun.) Hust.												0.1	0.4	13.9
<i>Eunotia</i> side view	0.3	0.1	0.4	1.1	0.2	1.3	0.7	0.1	0.1		0.4	0.1		
<i>Fragilaria arcus</i> (Ehr.) Cl.	0.1		0.1				0.1						0.4	
<i>F. bicapitata</i> Mayer	0.1													
<i>F. brevistriata</i> Grun.					0.1	0.1			0.5					
<i>F. capucina</i> Desm.			0.6	7.9	1.2	0.1	3.9	11.2	4.3	0.4	0.4	5.9	5.6	
<i>F. capucina</i> var. <i>amphicephala</i> (Grun.) L.-Bertalot						0.2								
<i>F. capucina</i> var. <i>austriaca</i> (Grun.) L.-Bertalot	0.4		0.6	0.1	0.3	0.1			0.3	0.4	0.7	0.1		

Site name	Grond	M-Gläna IL	M-Gläna OL	Immez IL	Sura OB	Sura IL	Sura OL	Immez IL	Immez OL	Zeznina	All	All	Immez	Dragun
Site number	1	2	3	4	5	6	7	8	9	10	NA	NA	NA	NA
Basin	North	North	North	North	South	South	South	South	Outlet	Outlet	South	North	Immez	Dragun
Stream type	Outlet	Inlet	Outlet	Inlet	Outlet	Inlet	Outlet	Inlet	Outlet	Stream	Lake	Lake	Lake	Lake
<i>F. capucina</i> cf. <i>austriaca</i>				0.1			0.5							
<i>F. capucina</i> var. <i>gracilis</i> (Oestrup) Hust.	0.1		0.1				0.1							
<i>F. capucina</i> var. <i>perminuta</i> Grun.		0.2	0.2	0.3			0.1	0.5	0.8	0.3				
<i>F. capucina</i> var. <i>rumpens</i> (Kütz.) L.-Bertalot									0.5					
<i>F. capucina</i> var. <i>vaucheriae</i> (Kütz.) L.-Bertalot	0.7	0.1	0.4		0.1	0.1	0.6		0.1	0.3	0.2	0.1		
<i>Fragilaria construens</i> var. <i>venter</i> (Ehr.) Grun.			0.0				0.1							
<i>F. exigua</i> Grun.			0.7	0.1					0.1			1.7		
<i>F. famelica</i> (Kütz.) L.-Bertalot				0.1										
<i>F. parasitica</i> (W. Smith) Grun.					0.1	0.1								
<i>F. virescens</i> Ralfs	0.1		0.6	0.8	0.6	0.1	0.4	0.1	0.3	0.5	0.2	1.0		
<i>F. virescens</i> var. <i>mesolepta</i> Schönfeld													0.4	
<i>Fragilaria</i> sp.	0.1	0.1		0.1				0.4			0.2	0.1		
<i>Frustulia rhomboides</i> (Ehr.) De Toni	0.1		0.5	0.1			0.1	0.1	0.3			0.9	0.9	1.1
<i>F. rhomboides</i> var. <i>saxonica</i> (Rabenh.) De Toni			0.1	0.1		0.2	0.3		0.9		0.5	0.2	0.4	0.7
<i>F. rhomboides</i> var. <i>crassinervia</i> (Bréb.) Ross	0.1			0.3					0.1					
<i>Gomphonema angustatum</i> (Kütz.) Rabenh.	0.4	0.2						0.1	0.1	0.1				
<i>G. gracile</i> Ehr.				0.1		0.1								
<i>G. olivaceum</i> Ehr.		0.1	0.1	0.1		0.2		0.1						
<i>G. parvulum</i> (Kütz.) Kütz.	0.5	7.7	2.5	16.4	1.6	0.7	4.2	24.6	21.0	9.0	0.4	1.1	2.6	
<i>Gomphonema</i> sp.					0.1		0.6							
<i>Hantzschia amphioxys</i> (Ehr.) Grun.	0.1		0.4			0.1		0.1			0.2	0.1		
<i>Luticola goeppertiana</i> (Bleisch) D. G. Mann	13.0			0.9					0.1	0.4				
<i>L. mutica</i> (Kütz.) D.G. Mann												0.1		
<i>L. nivalis</i> (Ehr.) D.G. Mann														0.4
<i>Mayamaea atomus</i> L.-Bertalot		0.1			0.3	0.1			0.1	0.3				
<i>Meridion circulare</i> (Greville) Ag.		0.1		0.1						0.1				
<i>Navicula</i> cf. <i>brekkaensis</i> Petersen											0.2	0.4	0.4	
<i>Navicula capitatoradiata</i> Germain			0.1		0.3									
<i>N. cincta</i> (Ehr.) Ralfs	1.2	0.4		0.1		0.1		0.1						
<i>N. cryptocephala</i> Kütz.	0.1													
<i>N. cryptotenella</i> L.-Bertalot	0.2	0.1	0.1			0.2	0.1	0.1		0.1	0.9	0.1	0.4	
<i>N. fonticola</i> Grun.											0.2			
<i>N. gallica</i> var. <i>perpusilla</i> (Grun.) L.-Bertalot											0.2	0.2	1.3	
<i>N. gregaria</i> Donkin						0.1	0.1							
<i>N. krasskei</i> Hust.			0.1	0.1	0.4	0.1					1.1			
<i>N. lanceolata</i> (Ag.) Ehrenb.			0.5	0.4		0.2				0.8	0.2	0.1		

<i>N. veneta</i> Kütz.	0.1			0.2							0.2	0.1		
<i>Navicula</i> cf. <i>splendicula</i> Van Landingham						0.1								
<i>Navicula</i> sp.	0.1	0.3		0.1		0.7	1.3	0.5	0.8	0.4	0.2	1.3	0.9	
<i>Naviculodicta schmassmannii</i> (Hust.) L.-Bertalot	0.2	0.1	0.4				0.1	0.1	0.2	0.1				
<i>Neidium affine</i> (Ehr.) Pfitzer						0.1		0.1	0.3		0.4	0.3		
<i>N. affine</i> var. <i>longiceps</i> (Greg.) Cl.			0.1	0.3		0.1								
<i>N. alpinum</i> Hust.	0.1					0.1	0.1	0.1			0.4	0.5		
<i>N. bisulcatum</i> (Lagerst.) Cl.				0.1							0.2	0.2		
<i>N. iridis</i> (Ehr.) Cl.	0.1		0.1											
<i>N. septentrionale</i> Cleve-Euler				0.3	0.1	0.1	0.2	0.1						
<i>Neidium</i> sp.					0.0	0.1				0.1				0.4
<i>Nitzschia dissipata</i> (Kütz.) Grun.	0.1	0.1		0.3			0.1		0.1			0.1		
<i>N. frustulum</i> (Kütz.) Grun.							0.1							
<i>N. gracilis</i> Hantzsch.	0.3		0.7			0.1	0.3	0.1		0.1	0.7	0.2		
<i>N. halophila</i> (Grun.) Cl.												0.2	0.4	
<i>N. hantzschiana</i> Rabenh.	0.3		2.1			0.1	0.3	0.1		0.1	0.2	0.9		
<i>N. iridis</i> (Ehr.) Cl.											0.2	0.3	0.4	
<i>N. palea</i> (Kütz.) W. Smith											0.2			
<i>N. perminuta</i> (Grun.) M. Peragallo	0.2	0.2	0.6	0.1	0.1		0.5	0.2	0.1					
<i>N. pseudoscutiformis</i> Hust.											0.2	0.3		0.4
<i>N. pseudoventralis</i> Hust.											0.2	0.1		
<i>N. schmassmannii</i> Hust.												0.5	1.7	
<i>N. sublinearis</i> Hust.				0.1					0.1					
<i>N. submolesta</i> Hust.											0.7	0.1		
<i>Nitzschia</i> sp.			0.1	0.1		0.1	0.7		0.1		0.2	0.3		
<i>Orthoseira roeseana</i> (Rabenh.) O'Meara						0.3								
<i>Pinnularia appendiculata</i> (Ag.) Cl.	0.1			0.1										
<i>P. biceps</i> Greg.	3.0	0.1	0.5						0.1					
<i>P. borealis</i> Ehr.				0.1		0.4		0.1			0.2	0.2	0.4	0.4
<i>P. gibba</i> Ehr.	0.1		0.1	0.1							0.2	0.1		
<i>P. hemiptera</i> (Kütz.) Cl.											0.2	0.1		
<i>P. ignobilis</i> (Krasske) Cleve-Euler							0.1	0.2						
<i>P. interrupta</i> W. Smith											0.2	0.3		
<i>P. microstauron</i> (Ehr.) Cl.	0.8	0.4	1.8	1.7	3.5	2.8	1.5	0.1	1.2	0.4	6.4	3.4	3.5	2.8
<i>P. molaris</i> (Grun.) Krammer						0.1	0.2							
<i>P. obscura</i> Krasske		0.1		0.2		0.2	0.1					0.2		0.4
<i>P. sinistra</i> Krammer	0.3	0.7	1.0	1.8	0.7	1.3	4.9	3.7	1.5	0.9				
<i>P. subcapitata</i> Greg.											1.6	2.3	2.6	
<i>P. sudetica</i> (Hilse) Hilse	0.1						0.1		0.1					
<i>P. viridis</i> (Nitzsch.) Ehr.				0.1							0.4	0.1		
<i>Pinnularia</i> sp.	0.1			0.3	0.1	0.2	0.1	0.1		0.3	0.5	0.6	0.4	0.7
<i>Planothidium delicatulum</i> (Kütz.) Round & Bukht.			0.5							0.1				
<i>P. lanceolatum</i> (Bréb.) Round & Bukht.	0.1	0.1	0.5	0.4		0.1	0.1			0.4				
<i>Psammothidium bioretti</i> (Germain) Bukht. & Round						0.1	0.2	0.1	0.1					
<i>P. helveticum</i> (Hust.) Bukht. & Round	0.2	0.2	0.7	1.9	24.7	17.8	13.4	2.8	5.3	0.4	10.5	4.0	2.6	
<i>Reimeria sinuata</i> (Greg.) Kociolek & Stoermer		0.1	0.2	0.1	0.1	0.2								
<i>Rhoicosphenia abbreviata</i> (Ag.) L.-Bertalot			0.1											
<i>Sellaphora pupula</i> (Kütz.) Mereschkowsky	0.2			0.1								0.2		

Appendix A. (continued)

Site name	Grond	M-Glūna IL	M-Glūna OL	Immez IL	Sura OB	Sura IL	Sura OL	Immez IL	Immez OL	Zeznina	All	All	Immez	Dragun
Site number	1	2	3	4	5	6	7	8	9	10	NA	NA	NA	NA
Basin	North	North	North	North	South	South	South	South	Outlet	Outlet	South	North	Immez	Dragun
Stream type	Outlet	Inlet	Outlet	Inlet	Outlet	Inlet	Outlet	Inlet	Outlet	Stream	Lake	Lake	Lake	Lake
<i>Stauroneis agrestis</i> Petersen						0.1								
<i>S. anceps</i> Ehr.	0.1		0.4			0.1	1.5	0.2	0.3	0.1	0.9	0.3		0.4
<i>Stauroneis</i> sp.						0.1						0.1		
<i>Staurosirella pinnata</i> (Ehr.) Williams & Round	2.1	0.2	0.1	0.1		1.5	1.5	0.2	1.2	0.4	0.2	3.1	0.9	
<i>Stephanodiscus alpinus</i> Hust.		0.1					0.1				0.2			
<i>Stenopterobia delicatissima</i> (Lewis) Bréb.												0.1		
<i>Surirella minuta</i> Bréb.				0.1										
<i>S. roba</i> Leclercq	0.4	0.1	0.4	0.8		0.4	1.2	0.1	0.1					
<i>S. terricola</i> L.-Bertalot			0.4	0.1	0.1	0.3	0.2	0.5						
<i>S. linearis</i> W. Smith											1.4	1.1	0.4	
<i>Surirella</i> sp.	0.1				0.1	0.1	0.1		0.3					
<i>Tabellaria flocculosa</i> (Roth) Kütz.	0.1	0.1	1.9	8.1	0.1	0.2	0.1		1.9	4.1	0.9	1.4	3.9	65.8