Research Article

Periphyton metabolism along a nutrient gradient in a desert river (Truckee River, Nevada, USA)

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Abstract. Periphyton metabolism was studied at five sites along a 70 km unshaded stretch of the Truckee River below the City of Reno, Nevada (USA). Sites differed with respect to concentrations of soluble reactive phosphorus and total dissolved inorganic nitrogen because a water pollution control facility discharged its treated wastewater through a small tributary into the river downstream of site 1 (most upstream located site). Unglazed tiles were incubated at each site from late June to August (summer) and from September to November (autumn) 1986. At the end of each incubation period, oxygen metabolism of the periphyton communities growing on tiles was measured in transparent flow-through respiration chambers for 24 h under near natural light and temperature conditions. In August, when biomass and metabolism were positively

correlated with nitrogen and phosphorus, mean chlorophyll *a* ranged from 53 (site 1) to 290 mg chl *a* m⁻² (site 2, 3 km downstream of the wastewater input). Periphyton gross primary production varied between 3.3 ± 0.8 (site 1) and 9.1 ± 1.4 g O₂ m⁻²d⁻¹ (site 2), and respiration between 2.1 ± 0.4 and 10.1 ± 1.3 g O₂ m⁻²d⁻¹. P/R ratios ranged from 0.9 at site 2 to 1.8 about 65 km downstream from the wastewater input. In November, chlorophyll *a* was significantly higher (99 ± 29 to 509 ± 155 mg chl *a* m⁻²) and gross primary production and respiration on average 50 and 65% lower, respectively, than in August. Hence, colder temperatures appeared to reduce respiration more than primary production and significantly increased P/R. Neither gross primary production nor respiration were correlated with autumn nutrient concentrations.

Key words. Primary production; community respiration; P/R; biomass; nutrients; chamber method.

Introduction

Periphyton is the primary autotrophic energy base in open-canopied streams (Minshall, 1978). Apart from light, temperature, nutrients and flow are major determinants controlling periphyton development (Biggs, 1995; Biggs, 1996; Borhardt, 1996; Fisher and Grimm, 1988; Hill, 1996; Uehlinger et al., 1996). Human activities

(urbanization, agriculture) are typically associated with prolific periphyton accrual as a result of enhanced nutrient input from runoff from fertilized crop land or sewage effluents, and increased light availability due to channel widening and removal of riparian vegetation (Biggs, 1995). Studies relating periphyton biomass and nutrients are numerous and directed to detect nutrient limitation and effects of nutrient enrichment (e.g., Bushong and Bachmann, 1989; Chessman et al., 1992; Pringle, 1987; Pringle and Bowers, 1983), or to describe nutrient-periphyton biomass relationships (e.g., Biggs, 1995; Biggs, 2000; Dodds et al., 2002; Taylor et al., 2004; Welch et

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al., 1988). Investigations on the relationship between periphyton metabolism and nutrients are less common (Guasch et al., 1995), in particular with respect to respiration or the balance between auto- and heterotrophic processes (typically characterized by the P/R ratio).

Streams in arid or semi-arid catchments (except from high latitudes or altitude) offer favorable conditions for algal growth, in particular with respect to light and temperature (Fisher et al., 1982; Minshall, 1978). In this environment, periphyton is expected to respond with rapid growth to enhanced nutrient supply unless frequent flooding, drought (zero flow), or low sediment stability limit accrual (Fisher and Grimm, 1988; Grimm and Fisher, 1989; Uehlinger et al., 2002). The focus of this paper is on periphyton metabolism along a 70-km river reach located in a semi-arid environment. This river system is influenced by a nutrient point source (sewage treatment facility) that created a distinct nutrient gradient with concentrations of dissolved inorganic nitrogen and phosphorus constituents varying by 1 to 2 orders of magnitude. Our objective was to assess changes in periphyton biomass and metabolism, including possible changes in the balance between auto- and heterotrophic processes along this nutrient gradient. To elucidate the effect of season on the manifestation of nutrient enrichment, we studied biomass and metabolism in summer and autumn, periods representing contrasting light and temperature conditions.

Methods

Study system and study design

The Truckee River flows from the oligotrophic Lake Tahoe (river km 0, 1898 m a.s.l.) to its terminus, Pyramid Lake (river km 187, 1157 m a.s.l.) in the Great Basin desert. The flow regime is characterized by high flow in spring and early summer (snowmelt in the Sierra Nevada) and a subsequent constant, managed, low discharge until spring. The base flow period during summer and autumn is maintained by releases from control structures on several lakes and reservoirs. Inter-annual variation in flow patterns is high with annual discharge varying between 2.6 and 70.8 m³s⁻¹. Most parts of the study reach (river km 97–166; hereafter, values represent distance from Lake Tahoe) are exposed to direct solar radiation because the active channel is wide (>20 m) and riparian tree vegetation is scattered or widely lacking. Bed sediments consist of sand, gravel, and at a few sites, cobble. Steamboat Creek, the recipient of the treated sewage from the cities of Reno and Sparks (combined population ~220,000 in 1986), enter the river at river km 101, and considerably increase concentrations of major nutrients. Three years after the end of this study about 75% of the inorganic nutrient load to the river was removed through

Figure 1. Location of the study sites along the Truckee River. The insert shows the study area within the state of Nevada.

an upgrade of the wastewater treatment facility that featured biological nutrient removal and denitrification.

We incubated artificial substrata at five stations between the city of Sparks and Pyramid Lake (Fig. 1), and assessed periphyton biomass and metabolism. The first station S1 (McCarran, city of Sparks at river km 97, 1335 m a.s.l.) was located about 4 km upstream of Steamboat Creek mouth near the East McCarran Boulevard Bridge. Stations S2 (Lockwood, river km 106), S3 (Clark, river km 125), S4 (S-S, river km 160), and S5 (Nixon, river km 166) were located on a nutrient gradient of the impacted reach below the confluence with Steamboat Creek. The design of the study did not allow a rigorous statistical evaluation of nutrient effects on river periphyton because of the lack of replication (only one river).

Physics and chemistry

From August to mid October 1986, incident photosynthetically active radiation (PAR) was recorded at 10-min intervals at S1 using an LI-192 SA quantum sensor connected to a LI-1000 datalogger (LI-COR Inc., Lincoln, Nebraska). Incident PAR for the period mid-October to November 24 were calculated from hourly solar radiation data recorded in Sparks (Renewable Resource Data Center database <http://rredc.nrel.gov/solar/>). On a few occasions, underwater PAR intensities were measured with a LI-192 SA underwater quantum sensor connected to a LI-1000 datalogger (LI-COR Inc.), and vertical PAR attenuation coefficients (k_{PAR}) were obtained by linear regression of $\ln[PAR(z)]$ on depth z . Water temperature was measured with thermistor probes (Yellow Springs Instrument Co., Yellow Springs, Ohio) at S1 several times a week between 6 August and 24 November 1986.

Table 1. Temperature conditions and light (PAR $\pm$ SD) in respiration chambers during metabolism measurements. avg = average temperature of all measurements, min = average minimum temperature, max = average maximum temperature.

Period	Temperature ($^{\circ}\text{C}$)			PAR	
	avg	min	max	Daily integral (E m^{-2})	Average intensity from 10 am - 2 pm ($\mu\text{E m}^{-2}\text{s}^{-1}$)
August	23	18	27	35.0 ± 2.8	1161 ± 69
November	7	5	10	15.0 ± 0.8	644 ± 23

Concentrations of soluble reactive phosphorus (SRP), ammonium ($\text{NH}_4\text{-N}$), nitrite ($\text{NO}_2\text{-N}$), and nitrate ($\text{NO}_3\text{-N}$) were determined by the Desert Research Institute, Water Resources Center (Reno, Nevada) and Truckee River Monitoring Program of the cities of Reno and Sparks. All analyses were performed according to APHA (1976) and USEPA (1985).

Periphyton

Artificial substrata (unglazed quarry tiles: $1.5 \times 7.5 \times 15$ cm) were incubated in riffle/run areas. Between 9 and 12 tiles were fastened in clusters on polypropylene netting (mesh size = 5 mm). At each site, 4 to 5 clusters were secured to the river bottom with steel rods. Current velocities and water depth were measured on several occasions during incubation. Velocity measurements were taken above the tiles at 0.6 times depth and at 2–5 cm above the tile using an electromagnetic transducer (Marsh McBirney Inc., Frederick, MD).

Two sets of tiles were collected: A summer series was retrieved in August after 7 to 8.5 wk of incubation in the river. An autumn series was collected in November after 9 to 11 wk of incubation. The longer incubation during fall partly compensated for the lower temperature sum (Rushforth and Brock, 1991). After retrieval, the tiles were transported to S1 for metabolism measurements and periphyton sampling.

The outdoor setup for measuring periphyton metabolism (see Uehlinger and Brock, 1991) consisted of 13 continuous flow-through respiration chambers immersed in a water bath cooled with water from the river. The chambers were made of acrylic glass and operated simultaneously. Polypropylene inserts allowed varying the chamber volume from 1.2 to 1.9 L. Internal circulation of water was provided by a submersible pump with variable discharge. A 16-channel peristaltic pump supplied corresponding river water (collected at each site simultaneously with tiles) to the chambers at rates varying between 0.35 and 0.90 L h^{-1} . Containers with river water were stored in ice chests and cooled with river water. The

Figure 2. Discharge (A) and incident solar radiation (B) of the Truckee River at Sparks (USGS Station 10348200) in 1986. The shaded area delimits the periods when artificial substrata were incubated.

chamber outflow and water from the containers with river water were collected in 50 ml BOD-bottles at intervals of 1 to 3 h. Concentrations of dissolved oxygen were measured using a 5905 YSI self-stirring BOD bottle probe (Yellow Springs Instrument Co., Yellow Springs, Ohio). The intensity of incident light was reduced by 32% using black greenhouse shading to approximate *in situ* light intensities during incubation in the river. Temperature and light conditions in the respiration chambers during the metabolism measurements, which typically lasted for 24 h, are summarized in Table 1. During measurements, average chamber temperatures exceeded the average river temperature (at S1) by 3°C in August but were 1°C below the average river temperature in November. Diel amplitudes were 2 (November) to 3°C (August) higher in the chambers than in the river. Net oxygen production $\beta(t)$ was calculated as:

$$\beta(t) = \frac{\Delta\text{O}_2}{\Delta t} - \frac{f}{V}(\text{O}_{2\text{in}} - \text{O}_2) \quad (1)$$

where O_2 is the oxygen concentration in each chamber ($\text{mg O}_2 \text{ L}^{-1}$) as measured in the chamber outflow, $\frac{\Delta\text{O}_2}{\Delta t}$ is the change in oxygen concentration between two subsequent O_2 measurements of the chamber outflow, $\text{O}_{2\text{in}}$ is the oxygen concentration ($\text{mg O}_2 \text{ L}^{-1}$) in the containers with river water (= O_2 concentration in the chamber inflow), f is the rate of the chamber inflow (L h^{-1}), and V is the water volume in the chamber (L). Four metabolic parameters (Bott et al., 1985) were based on net production

Figure 3. Concentration of ammonia ($\text{NH}_4^+\text{-N}$), nitrite ($\text{NO}_2^+\text{-N}$), nitrate ($\text{NO}_3^+\text{-N}$) (upper panels), and soluble reactive phosphorus (lower panels), during summer (June – August) and autumn (September – November) 1986 in the lower Truckee River.

rates (equation 1): net daily metabolism (NDM), community respiration (CR), gross primary production (GPP), and the ratio of gross primary production to community respiration (P/R). Net daily metabolism is the average net production rate of O_2 during 24 h. Average respiration during the dark period was extrapolated to 24 h to get estimates of CR. Gross primary production is sum of NDM and CR. Calculations of CR and GPP relied on the assumption that day respiration equals night respiration, but see Bott et al. (1985).

After metabolism measurements, samples of periphyton were removed from each tile by scrubbing a known area using templates and a stiff nylon brush. The resulting suspension was filtered through Whatman GF/F glass fiber filters. Filters were stored at -20°C prior to the analysis of chlorophyll *a* and ash-free dry mass (AFDM). Filters were ground using a glass homogenizer with a Teflon pestle, and chlorophyll *a* extracted with 90% acetone, and extracts cleared by centrifugation. After photometric determination of chlorophyll *a* and pheopigments (APHA, 1985), the acidified extract and centrifuged sediment were combined and dried at 60°C to constant weight, ashed at 550°C for 2 h, rewetted with distilled water, dried at 60°C , and weighed. The rewetting and redrying steps of the procedure were included as a precaution for loss on combustion of water associated with clays (Greenson et al., 1977). The autotrophic index (AI) calculated as the ratio AFDM: chlorophyll *a* is a measure to characterize the heterotrophic nature of the

periphyton community (the inverse of AI describes the chlorophyll *a* content of periphytic biofilms). High AI values (>100) have been associated with organic pollution (Weber, 1973).

Data analysis

Two-way ANOVA followed by post-hoc Tukey multiple comparison test was used to evaluate the effect of site and season (August, November) on biomass and metabolism. Data were transformed $\log(x+1)$ to improve normality (Zar, 1984). Effects were considered significant at $p < 0.05$. Regressions and correlations were assumed significant at $p < 0.05$.

Results

The physico-chemical environment

The hydrograph of 1986 (USGS gage in Sparks) was characterized by 2 large floods (February and March), and relatively high flow until mid June (Fig. 2). Daily discharge was low in July ($5\text{--}7 \text{ m}^3\text{s}^{-1}$, apart from a minor flow peak of $10 \text{ m}^3\text{s}^{-1}$ on July 26), August, and the first half of September. In the second half of September discharge began to increase, finally reaching $\sim 10 \text{ m}^3\text{s}^{-1}$ in October and November. Water diversion for irrigation, especially at Derby Dam, reduced flow in the lower river to levels as low as $2 \text{ m}^3\text{s}^{-1}$ (Rushforth and Brock, 1991).

Table 2. Two-way ANOVA results for periphyton biomass and metabolism (log-transformed data except for net daily metabolism).A) Chlorophyll-*a* (chl *a*), ash-free dry mass (AFDM) and the autotrophic index (AI).

Source	dF	chl <i>a</i>		AFDM		AI	
		SS	p	SS	p	SS	p
Site	4	17.7	<0.001	9.3	<0.001	1.2	<0.01
Season	1	42.0	<0.001	9.8	<0.001	13.8	<0.001
Site x season	4	14.2	<0.001	5.4	<0.01	4.1	<0.001

B) Net daily metabolism (NDM), gross primary production (GPP), community respiration (CR), and P/R ratio.

Source	dF	NDM		GPP		CR		P/R	
		SS	p	SS	p	SS	p	SS	p
Site	4	9.7	<0.001	5.9	<0.001	30.2	<0.001	2.8	<0.01
Season	1	19.2	<0.001	4.8	<0.001	18.1	<0.001	0.9	<0.001
Site x season	4	47.6	<0.001	1.9	<0.01	5.8	<0.01	0.4	n.s.

Water depth ranged from 0.36 to 0.67 m in summer and from 0.27 to 0.63 m in autumn. Corresponding current velocities (measured at 0.6 times depth) varied between 0.50 and 0.97 ms⁻¹ in summer and 0.48 and 0.99 ms⁻¹ in winter. Average water temperatures during the two incubations were 20.2 °C (summer) and 12.8 °C (autumn). Solar radiation (Fig. 2) averaged 7886 Wm⁻² in summer and 4157 Wm⁻² in autumn. In July, vertical PAR attenuation coefficients ranged from 0.85 (S1) to 1.93 m⁻¹ (S2).

During both incubation periods, concentrations of total dissolved inorganic nitrogen (NO₃-N, NO₂-N, NH₄-N) were relatively low at S1 (0.08 ± 0.08 mg N L⁻¹ in summer and 0.05 ± 0.01 mg N L⁻¹ in autumn) and dominated by NH₄-N (70%) (Fig. 3). Ammonium peaked at S2 in summer (1.36 ± 0.94 mg N L⁻¹) and autumn (1.21 ± 0.43 mg N L⁻¹, Fig. 3). During summer, NH₄-N rapidly declined along the 60 km reach from S2 to S5, and NO₂-N and NO₃-N reached maximum concentrations at S3 (0.20 ± 0.06 NO₂-N L⁻¹, 0.80 ± 0.24 NO₃-N L⁻¹). In autumn, the longitudinal trend of decreasing NH₄-N was less pronounced and the NO₃-N maximum was shifted downstream (presumably reflecting reduced nitrification rates at relatively low water temperatures). Concentrations of soluble reactive phosphorus (SRP) averaged 24 ± 14 (summer) and 12 ± 5 µg P L⁻¹ (autumn) at S1, peaked at S2 with 92 ± 63 µg P L⁻¹ (summer) and 39 ± 9 µg P L⁻¹ (autumn), and declined downstream (Fig. 3). SRP concentrations tended to be lower in autumn than in summer.

Periphyton

Analysis of variance suggested site and season as significant sources of variation in chlorophyll *a*, ash-free dry mass, autotrophic index (AI), and the four metabolism parameters (Table 3). The interaction of site x season was

Table 3. Correlations between concentrations of soluble reactive phosphorus (SRP) and total dissolved inorganic nitrogen (TDN) against periphyton biomass and metabolism parameters. AFDM = ash-free dry mass, AI = autotrophic index, GPP = gross primary production, CR = community respiration. n.s. = non significant.

	August		November	
	SRP	TDN	SRP	TDN
Chlorophyll <i>a</i>	0.78	0.81	0.48	0.45
AFDM	0.66	0.72	n.s.	n.s.
AI	-0.73	-0.57	n.s.	n.s.
GPP	0.65	0.55	n.s.	n.s.
CR	0.83	0.77	n.s.	n.s.
P/R	-0.63	-0.64	n.s.	n.s.

significant for both biomass parameters, AI, and three metabolism parameters (net daily metabolism, gross primary production, and community respiration). The correlation between nutrient concentration and periphyton biomass or metabolism indicates the effects of nutrient enrichment were mostly evident in August and not in November (Table 5). However, the limitation of the research to one river system precludes a rigorous statistical evaluation of nutrient effects on periphyton generally.

Biomass: Variation in chlorophyll *a* and ash-free dry mass (AFDM) along the study reach was distinct and significant at the end of both incubation periods (Fig. 4, Table 2). In August, AFDM and chlorophyll *a* were relatively low at S1 (53 ± 26 mg chl *a* m⁻², 18 ± 11 g AFDM m⁻²), peaked at S2 (290 ± 116 mg chl *a* m⁻², 49 ± 13 g AFDM m⁻²), and declined further downstream (Fig. 4). Differences in biomass between the stations S1, S4, and S5 were not significant (Tukey's test, *p* > 0.05). We observed a similar longitudinal pattern in November, except that there was a significant increase in chlorophyll *a* and AFDM between S4 and S5 for reasons that are not clear (Fig. 4). Chlorophyll *a* and AFDM were higher on tiles in

Figure 4. Chlorophyll *a* and ash-free dry mass (AFDM) grown on tiles (A) in August and (B) in November, and the autotrophic index (AI) in August and November (C).

November than August, a similar pattern to that observed on natural substrata (J. Brock, Unpublished Data). The autotrophic index decreased from 360 ± 78 (August) and 466 ± 66 (November) at S1 to 170 ± 41 (August) and 129 ± 24 (November) at S2, and increased from S3 to the end of the reach, particularly in August (Fig. 4).

Metabolism: A significant increase in gross primary production (GPP) and community respiration (CR) downstream of Steamboat Creek characterized the spatial (longitudinal) pattern in periphyton metabolism in August and November (Fig. 5A; B, Table 2). In August, gross primary production increased from 3.3 ± 0.8 g O₂ m⁻² d⁻¹ at S1 to 9.1 ± 1.4 g O₂ m⁻² d⁻¹ at S2, stayed at a high level (>7.8 g O₂ m⁻² d⁻¹) at S3 and S4, and finally declined to 4.8 ± 2.3 g O₂ m⁻² d⁻¹ at S5. Corresponding values of CR were 2.1 ± 0.4 and 10.1 ± 1.3 g O₂ m⁻² d⁻¹ at S1 and S2, respectively, and 3.0 ± 2.0 g O₂ m⁻² d⁻¹ at S5. Differences between sites were less pronounced in

November when GPP ranged from 2.2 ± 0.6 (S4) to 4.6 ± 1.0 (S3), and CR from 0.6 ± 0.2 (S4) to 1.7 ± 0.9 (S3). Net daily metabolism (NDM) in August was 1.2 ± 0.8 g O₂ m⁻² d⁻¹ at S1, decreasing to -1.0 ± 0.9 g O₂ m⁻² d⁻¹ at S2, and peaked at 2.6 ± 0.9 g O₂ m⁻² d⁻¹ at S5 (Fig. 5C). In November, NDM ranged from 1.3 ± 0.9 to 2.1 ± 0.6 g O₂ m⁻² d⁻¹. The P/R ratios varied between 0.9 ± 0.1 (S2) and 1.8 ± 0.4 (S5) in August, and between 2.0 ± 0.6 (S3) and 3.3 ± 2.3 (S5) in November (Fig. 5D). Both NDM and P/R were significantly higher in November than in August (Table 3), but post-hoc Tukey multiple comparisons indicated no significant differences between sites in November. Chlorophyll *a* and AFDM explained 43 to 67% of the variation in GPP, and 56 to 81% in CR in August, but only 26 to 35% of the variation in GPP and CR could be attributed to variation in chlorophyll *a* and ash-free dry mass (linear regression analysis) in November.

Discussion

Our study showed that in the Truckee River the response of periphyton metabolism to nutrient enrichment is strongly influenced by seasonal changes in light and temperature. During both incubation periods, flow was constant and low, longitudinal patterns in dissolved inorganic nitrogen (TDN) and soluble reactive phosphorus (SRP) were similar, while light and temperature conditions distinctly differed. The lack of hydrological disturbance during extended time periods is expected to favor biotic processes such as grazing, or the aging and subsequent sloughing of biofilms (e.g., Biggs, 1996; Fisher and Grimm, 1991). Poor water quality (e.g., high ammonia concentrations) downstream of Steamboat Creek inlet may have affected benthic grazers and, as a consequence, enhanced periphyton accrual (in addition to the stimulation of periphyton growth by the increased nutrient supply). However, the biomass of invertebrates classified as grazers correlated neither with periphyton biomass nor with ammonia and nitrite (Rushforth and Brock, 1991). Despite relatively long incubation periods, we found no evidence for autogenic sloughing.

Periphyton biomass and metabolism paralleled SRP and TDN concentrations in August. The high explanatory power of correlations between periphyton and nutrients suggests either SRP or TDN as a major stimulant of biomass and metabolism during summer. Upstream of the Steamboat Creek inlet, SRP varied between 12 and 24 µg P L⁻¹, and TDN between 0.05 and 0.08 mg N L⁻¹. The corresponding molar N: P ratios (TDN: SRP) ranged from 7.5 (summer) to 9.1 (autumn). The transition range from nutrient saturation to nutrient limitation is 0.6 to 10 µg P L⁻¹ for inorganic P and 0.05 to 0.06 mg N L⁻¹ for inorganic N (e.g., Bothwell, 1989; Grimm and Fisher, 1986; Newbold, 1992). Relatively high SRP

Figure 5. Periphyton gross primary production and community respiration (A) in August and (B) November. Net daily metabolism (C), and P/R ratio (D) of the periphyton community in August and November. The shaded areas delimit negative net daily metabolism rates and P/R values ≤ 1 .

concentrations and low N : P ratio suggest N limitation of periphyton upstream of the Steamboat Creek inlet. Steamboat Creek, carrying the treated wastewater of a large sewage treatment facility, increased TDN and SRP in the Truckee River far above limiting concentrations. After a flow distance of 65 km SRP and, especially, TDN were still elevated. Stimulation of periphyton accrual may not only result from increased concentrations of P and N. For example, flume experiments by Wuhrmann and Eichenberger (1975) showed that minor additions of sewage remarkably enhanced periphyton growth. Wuhrmann and Eichenberger (1975) hypothesized that minor nutrients occurring in wastewater may account for the growth stimulating effect.

The increase in periphyton biomass (chlorophyll *a* and ash-free dry mass) downstream of Steamboat Creek inlet was quite substantial in August. In terms of chlorophyll *a*, the river shifted from mesotrophy to eutrophy. The level considered to cause nuisance conditions ($200 \text{ mg chl } a \text{ m}^{-2}$ (Dodds et al., 1998; Welch et al., 1988) was approached ($193 \text{ mg chl } a \text{ m}^{-2}$ at S2) or exceeded ($290 \text{ mg chl } a \text{ m}^{-2}$ at S3). The increase in biomass was paralleled by a decline in the autotrophic index (AI). This change in AI primarily reflects a reduction of nutrient

stress downstream of Steamboat Creek inlet and not a reduction of organic pollution, which is expected to be highest at S2. The chlorophyll content of algae has been shown to decline with increasing N or P limitation (Healy and Hendzel, 1975; Rhee, 1978; Uehlinger, 1981).

In November, both chlorophyll *a* and ash-free dry mass were about 2 x higher than in August, and the link between nutrients and periphyton biomass was not evident (only chlorophyll *a* correlated with N and P). The increasing trend in periphyton biomass on tiles during the onset of colder autumn temperatures paralleled that observed on cobble substrata sampled from the Truckee River during the same period. Although the species composition tended to be less rich on tiles compared to natural substrata, the range of periphyton biomass overlapped with respect to chlorophyll *a* determined on tiles and cobbles, and both followed similar trends spatially (J. T. Brock, Unpublished Data). Seasonal shifts in composition of the diatom community were observed between summer and fall, with *Nitzschia dissipata* dominating the fall community in the nutrient-enriched zone downstream from Steamboat Creek (S2-S3) during autumn (Rushforth and Brock, 1991). High periphyton biomass during periods of apparently less favorable light and tem-

perature conditions has been observed elsewhere (De-long and Brusven, 1992; Shamsudin and Sleight, 1994; Uehlinger et al., 1996). Seasonally changing responses of periphyton biomass to nutrient amendments was found in 12 New Zealand gravel bed streams, where stimulation of periphyton accrual was maximum in summer, minimum in winter, and intermediate in autumn and spring (Francoeur et al., 1999). The pattern found in this study of elevated periphyton biomass during the cold season was observed in the Truckee River in several other subsequent years for which chlorophyll and AFDM data are available (J. T. Brock, Unpublished Data). These elevated biomass conditions observed for periphyton during late autumn and early winter may be associated with seasonally-reduced grazing pressure from benthic macroinvertebrates due to reduced numbers or lowered metabolic activity associated with cold temperatures.

The change in periphyton metabolism downstream of the point source was considerable, at least under the favorable light and temperature conditions in August. Daily gross primary production increased from 3.3 to 9.1 g O₂ m⁻²d⁻¹. However, these maximum rates are not exceptionally high compared with open-canopied rivers subject to similar or even minor nutrient supply and with periphyton as the dominant primary producer (e.g., Brock et al., 1999; Busch and Fisher, 1981; Cushing and Wolf, 1984; Fisher et al., 1982; Minshall, 1978; Uehlinger and Naegeli, 1998). In August, community respiration (CR) of periphyton responded to the enhanced nutrient supply with a significant increase (from 2.8 to 10.1 g O₂ m⁻²d⁻¹), and about 82% of the variation in CR could be explained by the variation in GPP (linear regression), indicating a close link between respiration and primary production. Periphyton respiration includes algal respiration, which is often assumed to be 10–50% of gross primary production (McAllister et al., 1964; Whittaker and Likens, 1975) and respiration of heterotrophic organisms located at the surface or in the biofilm matrix. In contrast to primary production, measurements of periphyton respiration provide limited information with respect to respiration at the ecosystem level since hyporheic respiration of open-canopied streams is known to contribute between 40–90% to ecosystem respiration (Grimm and Fisher, 1984; Naegeli and Uehlinger, 1997).

In November, gross primary production and respiration were significantly lower than in August (in contrast to biomass), presumably due to reduced light intensity and temperature. This finding concurs with the general annual patterns of primary production and respiration rates typically observed in many streams (e.g., Cushing and Wolf, 1984; Servais et al., 1984; Uehlinger, 1993). Periphyton metabolism in the Truckee River lacked an apparent response to the enhanced nutrient availability downstream of the point source in November. In contrast, Guasch (1995) found a distinct increase in net daily me-

tabolism and gross primary production following nutrient addition (P, N, K) to stream flumes in winter, despite low light intensity (31 E m⁻²d⁻¹) and temperature (3.6 °C).

The P/R ratio characterizes the balance between auto- and heterotrophic processes in the periphyton community. If algae respire between 10 and 50% of gross primary production, upper limits of P/R of periphyton are expected to range from 2–10 (respiration of heterotrophic microorganisms occurring in autotrophic biofilms not included). The P/R of periphyton (ratio based on daily gross primary production and respiration rates) has been reported to be >1 (Munn and Brusven, 2004; Peterson et al., 1986). The transient depression of P/R downstream of Steamboat Creek inlet in August to values <1 can be the outcome of several mechanisms, including a shift in the relative proportion of the photic and aphotic zone with increasing biofilm density (major parts of dense biofilms can be light limited) (Dodds et al., 1999), the supply of organic matter from the sewage treatment plant supporting additional heterotrophic activity (besides algal respiration), and the oxygen consumption by nitrifying bacteria. Concentrations of NH₄-N and NO₂-N indicated that the reach between river km 101 and 125 is an active zone of nitrification. We are aware that problems may arise if P/R is used to characterize the trophic status of functional compartments exhibiting substantial chemo-autotrophic activity. Metabolism of the periphyton community became more autotrophic with the seasonal change; in November P/R ratios averaged 2.6 compared to 1.4 in August. The reduction of light and temperature by 60 and 70%, respectively, from August to November apparently influenced gross primary production and community respiration to a different extent. Primary production is known to be less temperature dependent than respiration (Busch and Fisher, 1981; Phinney and McIntire, 1965). For example, Busch and Fisher (1981) reported Q₁₀ temperature coefficients of 1.5 for gross primary production and 2.5 for community respiration in Sycamore Creek. Photosynthesis of epilithic algae has been reported to be light saturated at intensities ranging from 100 to 600 μE m⁻²s⁻¹ (e.g., Boston and Hill, 1991; Dodds et al., 1999; Hill, 1996; Hill and Boston, 1991). Compared to these values, PAR intensities in November were still relatively high (>600 μE m⁻²s⁻¹ for about 4 h each day, Table 1), and may account for the relatively moderate reduction in GPP.

To summarize, periphyton responded to nutrient enrichment from a point source with a significant increase in biomass and metabolism, but only under favorable light and temperature conditions prevailing during summer in a desert river. The effect of nutrient enrichment was hardly evident in autumn, indicating a strong seasonality in the response to nutrient amendment. The balance between auto- and heterotrophic processes in the periphyton community also showed distinct seasonal

variation under conditions of constant and low flow. The conditions observed of negative net daily oxygen metabolism during summer have implications to the oxygen balance in this and other desert rivers, where low dissolved oxygen conditions may prevail seasonally and serve as a stressor for aquatic biota.

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