## Nutrient release from moose bioturbation in aquatic ecosystems

Joseph. K. Bump<sup>1</sup>, Brenda G. Bergman<sup>1</sup>, Amy J. Schrank<sup>1</sup>, Amy M. Marcarelli<sup>2</sup>, Evan S. Kane<sup>1</sup>, Anita C. Risch<sup>3</sup> and Martin Schütz<sup>3</sup>

<sup>1</sup>School of Forest Resources and Environmental Science, Michigan Technological Univ., Houghton, MI 49931, USA

<sup>2</sup>Dept of Biological Sciences, Michigan Technological Univ., Houghton, MI, USA

<sup>3</sup>Community Ecology, Swiss Federal Inst. for Forest, Snow, and Landscape Research WSL, Birmensdorf, Switzerland

Corresponding author: J. K. Bump, School of Forest Resources and Environmental Science, Michigan Technological Univ., Houghton, MI 49931, USA. E-mail: jkbump@mtu.edu

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# (ABSTRACT)

While the ecological importance of bioturbation is well recognized and the prevalence of aquatic foraging by terrestrial ungulates is increasingly appreciated, research linking how terrestrial ungulates function as disturbance mechanisms via bioturbation in freshwater systems is lacking. The purpose of this study was to quantify potential nutrient pulses released from benthic sediments into the water column when moose Alces alces feed on aquatic plants. We also determined if we could experimentally mimic the benthic disturbance and the expected nutrient pulse created when moose feed aquatically. When moose foraged aquatically, significant releases of both total and dissolved phosphorus (P) and nitrogen (N) resulted in the waters that were disturbed in foraging areas compared to adjacent undisturbed waters. Nutrient concentrations for total P and N ranged from 42.5× and 2.7× greater in disturbed than undisturbed, respectively. Dissolved P and N were 26.8× and 1.5× greater, respectively, in disturbed versus undisturbed waters. Our experimental mimic created increases of total and dissolved P and N that were equivalent to pulses created by moose. This indicates that it is possible to experimentally test by proxy the potential impact of moose bioturbation on other ecosystem processes. This study is the first quantification of moose foraging as a consumer mechanism that influences the release of limiting nutrients in aquatic systems, thereby emphasizing the potential cascading importance for nutrient uptake and productivity of plants and microbes.

**KEYWORDS:** *Alces alces*, disturbance; ecosystem engineer; herbivory; littoral zone; macrophyte; nitrogen; phosphorus; resource pulse; environmental heterogeneity.

#### INTRODUCTION

Disturbances and consumer fluxes play an important role in shaping ecological systems. Understanding the mechanisms and consequences of disturbance and consumer fluxes in ecosystems is a fundamental focus in ecology (Walker 2012, Pickett and White 2013, Sinclair et al. 2015, Allen and Wesner 2016). Disturbances and consumer effects are important causes of spatial heterogeneity across multiple scales, shaping mosaics of functional ecosystem attributes, community dynamics, biodiversity, and dynamic feedbacks (Turner 2010, Turner and Gardner 2015). Biotic disturbances are mechanisms that create heterogeneity, especially those linked to consumer effects on ecosystems processes (Leroux and Loreau 2010, Schmitz et al. 2010). Such effects have received much recent research attention because of the reciprocal relationship between an individual organism's ability to both create and respond to spatial heterogeneity (Wisdom et al. 2006, Reichmann and Seabloom 2002, Gabet et al. 2003). Consumers respond to patterns of patchiness and through multiple feedback pathways can amplify or reduce spatial heterogeneity in ecosystem processes (McIntyre et al. 2008, Allen and Wesner 2016). For example, top predators such as wolves (Canis lupus) can strongly modulate prey carcass distribution in time and space (Bump et al. 2009a, Wilmers and Getz 2004), which influences the distribution of biogeochemical hotspots at carcass sites (Bump et al. 2009c) and the subsequent response of herbivores and scavengers. Such feedbacks that affect environmental heterogeneity are important because shifting heterogeneity is theoretically (Franklin 2005) and increasingly empirically (Pickett et al. 1997) linked to the maintenance of biodiversity.

Bioturbation is a consumer and disturbance mechanism creating feedbacks. In the broad sense, bioturbation is the biological reworking of soils and sediments (Darwin 1881, Thayer 1979, Levinton 1995, Meysman et al. 2006, Wilkinson et al. 2009, Kristensen et al. 2011). As such, bioturbation has significant effects on sediment texture, water and solute transport, and dispersal of solid particles, thereby coupling aboveground-belowground and benthic-water column interactions. 'Bioturbators' have been categorized as ecosystem engineers, as their disturbance effects can produce strong, lasting, and disproportionally large impacts on ecosystems (Moore et al. 2006). Small invertebrates are the most abundant and ubiquitous bioturbators globally (Bottinelli et al. 2015). However, some large vertebrates also create significant bioturbation effects. For example, in terrestrial systems digging mammals such as moles, gophers, and prairie dogs significantly disturb soils by creating subterranean burrow systems, thereby affecting hydrology and nutrient cycling (see e.g., Davidson et al. 2012). In marine systems, grey whales and walruses plough trenches and create conspicuous pits in the surfaces of benthic sediments. In

seagrass meadows dugongs and stingrays create similar disturbances when feeding, resulting in multiple ecosystem effects (reviewed in Meysman et al. 2006). Freshwater systems have comparatively fewer conspicuous vertebrate bioturbators, with the exception of some fish species (Vanni 2002, Vanni et al. 2002, Moore et al. 2006, Janetski et al. 2009). Fewer still are mammals that are biotubators in freshwater systems, such as North American beaver (*Castor Canadensis*; Hood and Larson 2015) and hippopotamus (*Hippopotamus amphibius*; McCarthy et al. 1998, Deocampo 2002, Moore 2006, Mosepele et al. 2009, Subalusky et al. 2015). Additionally, while the cross-ecosystem impact of cervid herbivory on aquatic macrophytes is recently recognized (Ceacero et al. 2014, Takafumi et al. 2015, Bakker et al. 2016), cervid bioturbation effects on aquatic nutrient cycling is under examined.

Here we report on the previously unquantified role of moose (Alces alces) as an important bioturbator in aquatic ecosystems. Large herbivores such as moose are generally recognized as significant sources of physical (i.e., trampling, wallows, tree toppling) and biogeochemical (i.e., dung, urine, carcasses) disturbance in terrestrial ecosystems (Hobbs 1996, Moore 2006, Wisdom et al. 2006), affecting plant community composition, nutrient cycling, and patch use among herbivores. Moose are also recognized as important aquatic-terrestrial interface specialists and resource vectors (Ballinger and Lake 2006, Doughty et al. 2015, Bakker et al. 2016), foraging on aquatic macrophytes and transporting significant amounts of aquatic-derived nitrogen (N) to terrestrial systems (Bump et al. 2009b). However, the effect of bioturbation of lake sediments by moose while foraging for aquatic plants is unknown. Moose foraging in aquatic systems likely suspends nutrient-rich sediment and interstitial waters, where concentrations of productivity limiting elements are orders of magnitude greater than in overlying water (Levine and Schindler 1992; Søndergaard et al. 1992, 2003). Even though the evolution of the muzzle anatomy in moose has been suggested as a morphological adaptation for underwater feeding (e.g., moose are able to seal their nostrils underwater; Clifford and Lawrence 2004), their functional role as disturbance agents in aquatic ecosystems is unexamined. A better understanding of the role of moose in aquatic ecosystems and development of experimental approaches for exploring their role is warranted (Bakker et al. 2016).

We focus on quantifying nutrient release because key elements [e.g., phosphorus (P) and nitrogen (N)] are generally limiting to primary production in boreal freshwater systems (Elser et al. 2007; Bergström 2010). Even low frequency, short duration pulsed episodes of increased resource availability affect community dynamics and ecosystem processes (Nowlin et al. 2008, Yang et al. 2010, Marcarelli et al. 2011, Greenville et al. 2014). Resource pulse events affect consumers at various levels, including individual level behavioral and life history traits, population level numerical responses, and community

level indirect effects (Holt 2008, Yang et al. 2008, 2010). Even when resource pulses are low in frequency and short in duration, consumers employ opportunistic strategies to capitalize on ephemeral resource increases (Yang et al 2010) and short-term responses to recurrent pulses can alter community structure by affecting coexistence dynamics and transitions between alternative states (Holt 2008). Opportunistic response strategies can reduce average resource availability over larger spatial scales (e.g., Yang et al 2010, Greenville et al. 2014). Such strategies allow consumers to tolerate prolonged inter pulse periods of decreased resource availability (Takimoto et al. 2009), which is especially important in resource limited systems such as the north-temperate lakes in this study. We expect then that moose bioturbation releases nutrient pulses at ecologically significant quantities.

In this study we quantified possible pulses of total and dissolved nutrients released into the water column when moose forage on aquatic plants. We also experimentally tested our ability to in-situ create an analogous mimic of moose-derived nutrient pulses. The ability to experimentally mimic moose aquatic disturbances is important because 1) sampling water after mimicked disturbances is logistically more feasible than sampling after opportunistic observations of moose foraging in aquatic habitats and 2) experimental approaches are likely to increase mechanistic understanding and inference (Ford and Goheen 2015). The effective mimicking of moose aquatic disturbances would support experimental research on how moose bioturbation affects aquatic systems. Such experiments are necessary to advance our understanding of feedbacks between animal populations and ecosystem processes across ecological interfaces (Polis et al. 2004). For example, if moose bioturbation creates significant nutrient pulses in freshwater systems, internal nutrient availability could hypothetically increase, producing positive effects on biotic uptake and microbial productivity. Testing such hypotheses is appreciably more feasible if we are able to experimentally mimic moose bioturbation in aquatic ecosystems (e.g., Valentine et al. 1994, Usio and Townsend 2004).

## MATERIALS AND METHODS

Study system and experimental design – We conducted field observations and collected water samples in aquatic ecosystems within Isle Royale National Park, an archipelago of islands in northwestern Lake Superior, USA (48°N, 89°W). The largest island, Isle Royale (~530 km²), is approximately 72 km long and 14 km wide, 24 km from the nearest mainland shoreline, and is surrounded by numerous small islands. Isle Royale exhibits a distinctive topography of fingered peninsulas, indented bays, and a series of parallel ridge-and-valley landforms along a northeast-southwest alignment (Huber 1975). Drainage is

poorly developed due to these topographic features. As a consequence Isle Royale exhibits numerous inland, lacustrine water bodies. Lakes and ponds ( $n = 84 \ge 1$  ha) cover ~36 km² of the island's surface; palustrine emergent wetlands cover another 8 km² (Tischler 2004). Plant communities reflect Isle Royale's geographic position between boreal and the northern hardwoods regions (Hansen et al. 1973), with forest mosaics of evergreen, mixed-evergreen and deciduous trees of two major associations: sugar maple-yellow birch and birch-balsam fir-white spruce (Hansen et al. 1973). Moose density over the past five decades ranged from ~1-5 moose per km² at the landscape level and is currently increasing (Peterson et al. 2014).

Within this system moose consume aquatic macrophytes extensively between May and September. Average daytime aquatic foraging bouts per individual are  $\sim$ 40 minutes and multiple bouts have been observed in a day (Belovsky and Jordan 1978). The mean aquatic-derived proportion of the diet during this time period is 14-37% (95% CI 9 - 54%) based on isotopic diet analysis (Tischler 2004, Bump et al. 2009b). During aquatic foraging bouts, plant consumption and feeding movements create notable disturbances that entrain and distribute presumably nutrient-rich benthic sediments and interstitial water vertically and horizontally (Figure 1). Adult moose may submerge completely during aquatic foraging (Supplementary materials Video1) and create disturbances that spread over an extended area (e.g., >100 m<sup>2</sup>). Field observations of benthic sediment clinging to moose hide indicate that moose can penetrate the benthos to a depth of  $\geq$  1 meter, depending on bottom firmness.

We used a paired design in sampling and analysis to test for nutrient pulses in lake water caused by moose foraging and to test our ability to experimentally mimic aquatic disturbances events created by moose. The moose treatment was contingent upon opportunistic sightings and accessibility, hence sampling occurred in the zone where moose were foraging and in adjacent, upwind zones of the same depth and bottom firmness that were beyond the area of foraging activity, hereafter "disturbed" and "undisturbed" respectively (Figure 1). Water sampling and analyses included two water conditions (disturbed and undisturbed), two disturbance sources (moose and investigator mimic), and an undetermined number of replicates (i.e., *n*) for each treatment. The number of replicates was contingent upon opportunistic sightings and accessibility of moose foraging in aquatic environments. Sites of moose disturbance and mimic treatment occurred in the same lakes as moose sites. Both filtered and unfiltered water samples were collected to partition dissolved fractions of nutrients from particulate-derived fractions.

Field sampling - Aquatic feeding bouts average 43 min on Isle Royale and occur on an average on 108

days seasonally (Belovsky and Jordan 1978). In spring (May-June) of 2010 and 2011 moose were opportunistically observed foraging in littoral zones of three inland lakes that were comparatively small (Daisy Farm Trail, 0.01 km<sup>2</sup>), medium (Ojibway, 0.15 km<sup>2</sup>), and large (Feldtmann, 1.86 km<sup>2</sup>). Lakes spanned both the east and west ends of Isle Royale. Littoral zone benthic sediments in foraging areas were of similar firmness and exhibited a mix of silt and sand. Once moose ceased foraging (moose were observed foraging for at least 10 minutes for all sampling) and exited the water, samples of lake water were collected ~0.25m below the water surface in paired disturbed and undisturbed waters (Figure 1). Distance between undisturbed water samples and the edge of the disturbed water sample area was at least 4 m. Subsequently, when moose were not present in the water, experimental attempts were made to mimic the moose disturbance and bioturbation effect by wading, walking, and stomping in littoral zone areas that were adjacent to, but not overlapping with, previous foraging sites (hereafter 'mimic'). Disturbance mimicking lasted 10 minutes, occurred in waters from ~0.75-1.25 m deep, and was done by the same, single investigator each time. The duration of disturbance mimicking was based on the observation that surface waters often appear clouded with sediments within  $\leq 2$  min of aquatic foraging by moose even though foraging bouts typically last longer. Paired water samples were collected from mimic disturbed and undisturbed areas respectively. To examine pulses of dissolved nutrients in the same lakes, in 2011 and 2012 duplicate water samples for each treatment, i.e., moose or mimic, were collected and filtered through a syringe filter (0.2 µm nylon membrane, Acrodisc®). Material that could pass a 0.2 µm filter was considered dissolved and more readily available for biotic uptake.

Observations of adult moose foraging during May and June 2010 and 2011 led to a total of 15 paired samples of disturbed versus undisturbed treatment for total nutrient analysis, which were associated with 13 paired mimic samples of disturbed versus undisturbed. All samples from moose foraging occurred from different individuals based on observations of distinct body characteristics (e.g. hair pattern, antler development). The mimic sample size was less than the moose sample size because on two occasions, comparable areas for the experimental mimic were unavailable due to extensive moose disturbance (e.g., foraging lasted >1hr in one instance) and significantly different littoral zone characteristics (e.g., abrupt depth increases and bottom firmness).

Laboratory and data analyses – In 2010, water samples were acidified with sulfuric acid in the field and stored in opaque, high-density polyethylene containers. Samples were refrigerated and subsequently analyzed within three weeks for total phosphorus (TP) and total nitrogen (TN) at the University of Michigan's Biological Station using a Seal Analytical AA3 segmented flow analyzer. The chemistry

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followed the Standard Methods procedures (REF methods manual,  $20^{th}$  edition): TP was digested using acidified persulfate (SM 4500-P(B.5)) followed by method SM 4500-P(F) then quantified using the ascorbic acid method (Ameel et al. 1993). TN was digested using basic persulfate followed by quantification using cadmium reduction SM 4500-NO<sub>3</sub>(F). TP detection limit was 2.5  $\mu$ g L<sup>-1</sup> with a precision of +/- 12%; TN detection limit was 44  $\mu$ g L<sup>-1</sup> a precision of +/- 18%. In 2011, samples were not acidified, but received the same storage and analysis within two weeks. Filtered water sample values indicate total dissolved nitrogen (TDN) and total dissolved phosphorus (TDP) availability.

Total and dissolved nutrient data were analyzed using nonparametric paired analysis (Wilcoxon signed rank tests) to test the expectation of no difference in total and total dissolved P or N between disturbed and undisturbed samples, separated by moose and mimic treatments (JMP Pro 10 SAS Institute); a non-parametric test was used so as to not assume the data belong to a particular distribution. The paired design accounts for across site differences among lakes, times, years in which moose were opportunistically observed foraging (Zar 2010). In sum, each paired set of samples were collected and analyzed in the same way. Estimates of effect size (difference between disturbed and undisturbed lake surface water samples) were compared to test the expectation that the effect of moose disturbance could be accurately mimicked experimentally by human disturbance as described above.

#### RESULTS

Our analysis indicates that moose cause significant (n = 15 F = 8.3, p < 0.0001) increases in both total P concentrations (mean increase = 621 µg/L) and total N concentrations (mean increase = 885 µg/L) in the waters that are disturbed in foraging areas approximately 5 minutes after foraging (Figure 2); total P and N measured in waters disturbed by moose were 42.5× and 2.7× greater than undisturbed, respectively. Similarly, moose foraging resulted in significant (n = 15, F = 4.1,  $p \le 0.008$ ) increases of both dissolved P concentrations (mean increase = 335 µg/L) and N concentrations (mean increase = 179 µg/L) relative to undisturbed waters; concentrations of dissolved P and N were 26.8× and 1.5× greater, respectively (Figure 2). The increase in nutrient concentrations for both total and dissolved nutrients created by moose disturbance was not significantly different for P nor N from the effect size created by attempts to mimic moose bioturbation (n = 13, p-values between 0.45 – 0.75; Figure 3). The experimental mimic increased total P and N 45.6× and 2.1× greater than undisturbed, respectively, and dissolved P and N 35.2× and 1.7× greater than undisturbed, respectively. Based on qualitative observations, disturbance was readily visible in water for at least three hours following moose foraging.

#### DISCUSSION

Moose bioturbation during aquatic foraging resulted in increased concentrations (total and dissolved) of P and N in the water column in the north-temperate/boreal lakes we sampled. These moose-released nutrients are then available to primary producers (Wetzel 1992, Christiansen et al. 2016), providing an ecologically significant resource pulse for pelagic food webs. Similarly, benthic, aquatic bioturbators (such as macroinvertebrates and bivalves) can have rapid effects on nutrient cycling by facilitating nutrient transport across the sediment-water interface and enhancing nutrient availability to pelagic organisms (Anschutz et al. 2012, Blankson and Klerks 2016, Remaili et al. 2016). While the significant influence that aquatic consumers have on internal P and N cycling has been recognized (reviewed by Vanni 2002, Vanni et al. 2013, Capps et al. 2015, Hölker et al. 2015), such analyses have not included moose. Moose and other large herbivores may not be as numerous as smaller consumers but larger organisms are expected to have larger per capita impacts as bioturbators (Moore 2006, Bakker et al. 2016).

The net effect of moose biotubation on water nutrient concentrations depends on a combinations of variables, e.g. moose population density, total days spent feeding on aquatic plants, length of foraging bouts, area disturbed, and littoral zone area. Based on the mean total P and N released in a single foraging bout (Figure 2), conservative estimates of the mean days spent foraging on aquatic plants (108; Belovsky and Jordan 1978), a single aquatic foraging bout per day, and a minimal volume of water disturbed per feeding bout based on field observations (25 m³), a single moose could resuspend 1.68 kg of P and 2.39 kg N per year. These estimates assume that the increased total P and N concentrations (Figure 2) result from a single release and not a continued resuspension during a foraging bout, which is likely conservative because moose are moving and disturbing the benthos frequently when feeding aquatically. Additionally, moose often feed on aquatic plants more than once per day; for example, at least 6 moose were observed feeding in the same lake during our study.

To put moose-released nutrients into context, the annual surface N deposition for our study lakes is estimated to be approximately 2 kg (Daisy Farm Trail), 25 kg (Ojibway), and 311 kg (Feldtmann). This range reflects differences in lake area, not variable atmospheric deposition rates. In small, relatively pristine lakes with low landscape inputs of nutrients such as those on Isle Royale, atmospheric deposition has been estimated at 50 – 90% of N inputs to the system (Likens 1985, Stottlemyer et al. 1998, Winter and Likens 2009). Therefore, for a small lake such as Daisy Farm Trail, a single foraging moose would nearly double the rate of N input to the system. Estimating the effects of moose on lake P budgets is less clear; for example, estimates of atmospheric deposition of P are variable, from less than 5% to 100%

(Schindler et al. 1976, Whalen and Cornwell 1985, Winter and Likens 2009). However, the importance of internal P resuspension in lake P budgets is clear (Søndergaard et al. 1992). For example, Chaffin and Kane (2010) suggested that sediment bioturbation by burrowing mayfly (*Hexagenia spp.*) in the western basin of Lake Erie was capable of releasing 1.40 mg/m² P per day, (504 mg/m² P per year) leading to a significant source of phosphorus loading. Our estimates suggest that one moose could add 168 mg/m² P per year to lake systems (e.g., Daisy Farm Trail), an amount that would likely influence nutrient limited primary producers. Understanding the net effect of aquatic nutrient releases by moose also requires more intensive analyses of time-series samples and detailed partitioning of chemical composition because both total and dissolved fractions of P and N likely contain components that vary in their availability. In general the dissolved fractions of P and N are considered more biologically available, but there is variation in the ability of microbes to directly uptake and utilize different inorganic and organic forms (Cotner and Wetzel 1992, Berman and Chava 1999), as well as the potential to access particulate-bound nutrients via extracellular enzyme activity (eg. alkaline phosphatases; Jansson et al. 1988). However, even our conservative, estimates of the effects of moose bioturbation on nutrient release suggest that moose foraging can have lake-level ecological consequences, especially for nutrient poor systems.

Pulses of dissolved P and N (Figure 2) at levels similar to those measured in this study are typically available for immediate uptake by primary producers, both algal and macrophyte species (Wetzel 1992, Christiansen et al. 2016). For example, common bladderwort (*Utricularia vulgaris*), a carnivorous aquatic macrophyte which grows in Isle Royale lakes, can reduce its investment in carnivory as more nutrients become available in the water column (Kibriya and Jones 2007), which can lead to significant increases in the populations of certain prey species, including insect larvae (Gordeev and Sibataev 1995, Harms 2002). Reductions in *U. vulgaris* carnivory occurred at water nutrient levels lower than the moose mediated release levels observed in our study. Furthermore, there are numerous studies suggesting that addition or enhanced recycling of nutrients in small, nutrient limited lake systems have transmitted effects up the food web (reviewed in Vanni et al. 2002, Meunier et al. 2016) suggesting the nutrient releases we recorded can indirectly affect community and nutrient dynamics.

Our understanding of how large herbivores influence nutrient cycling is largely dominated by a terrestrial understanding (Danell et al. 2006, but see Bakker et al. 2016). The net effect of moose foraging and excretion in forests is largely considered to decelerate (*sensu* Ritchie et al. 1998, Singer and Schoenecker 2003) N cycling as these animals selectively avoid less palatable forage that is more recalcitrant to microbial decomposition (Pastor et al. 1988, Christenson et al. 2014). The potential for ungulates to influence terrestrial P mineralization appears less than that found for N (e.g., Frank 2008,

Abbas et al. 2012). In contrast to the functional role of moose in terrestrial systems, this study indicates that aquatic herbivory by moose potentially accelerates the internal cycling and increases the availability of limiting nutrients in lakes. This role is important as the geographic range occupied by moose is extensive (e.g., ~28% of North America north of Mexico; Larsen 1980), circumpolar, characterized by a wide spectrum of freshwater ecosystems, and across these aquatic systems, both P and N are generally limiting nutrients (Elser et al. 2007).

To address the importance of moose or other ungulate bioturbation by directly studying ungulates is ideal, but can be a significant challenge to researchers. Our mimic results indicate that experimental disturbance can likely serve as a valid surrogate for moose bioturbation effects on water column dynamics to complement direct observations. These results suggest then that it is possible to begin to test the potential impact of a proxy of moose bioturbation on other ecosystem processes, such as structuring littoral zone habitat, modifying sediment mixing and oxygenation, and influencing turbidity and light penetration (Bakker et al. 2016). When impacts of moose bioturbation on such processes are experimentally estimated, we can more readily scale bioturbation effects to the lake and landscape level, identify explanatory mechanisms, and make stronger inferences (Ford and Goheen 2015). Further testing is needed to reveal whether or not an investigator mimic can successfully be used to examine benthic dynamics, e.g. mixing depth or surface area disturbance. Appreciation for ungulate effects on aquatic macrophyte communities has recently expanded (Ceacero et al. 2014, Takafumi et al. 2015, Moss 2015, Bakker et al. 2016) and similar testing of experimental mimics is recommended in other systems.

The extent to which bioturbation by moose affect other ecosystem conditions, components, and processes in addition to nutrient pulses is unexplored (Bergman and Bump 2015). For example, bioturbation has the potential to alter contaminant (e.g., mercury) transport across the sediment-water column interface and to terrestrial systems (e.g., Bergman and Bump 2014), which can affect the metal bioavailability, bioaccumulation and toxicity to organisms (Anschutz 2012, Blankson and Klerks 2016, Remalli et al. 2016). While we focused on limiting nutrients in north-temperate freshwaters, observations of bioturbation by moose indicate that sediment mixing, oxygen, and turbidity are potentially affected as well (Figure 1). Bioturbation in lake ecosystems generally causes a significant increase in oxygen at the sediment-water interface (Mermillod-Blondin and Rosenberg 2006, Mermillod-Blondin 2011). The mere movement of large mammals in water can significantly increase subsurface dissolved oxygen levels, preventing anoxia in waters with extreme nutrient loading (Gereta and Wolanski 1998, Wolanksi and Gereta 1999, Deocampo 2002). Such oxygen addition may affect P storage and the cycling of nutrients sensitive to redox state. Water clarity affects primary production of both macrophtyes and algal species

(Middleboe and Markager 1997, Karlsson et al. 2009), which in turn are important to primary (e.g., zooplankton) and secondary (e.g., fish) consumers. Hence future studies on this topic should include not only a broader analysis of the direct effects of moose bioturbation, but an examination of cascading, indirect effects as well.

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#### REFERENCES

- Abbas, F., Merlet, J., Morellet, N., Verheyden, H., Hewison, A. J. M., Cargnelutti, B., Angibault, J. M., Picot, D., Rames, J. L., Lourtet, B., Aulagnier, S. and Daufresnse, T. 2012. Roe deer may markedly alter forest nitrogen and phosphorus budgets across Europe. Oikos 121: 1271-1278.
- Allen, D.C. and Wesner, J.S. 2016 Synthesis: comparing effects of resource and consumer fluxes into recipient food webs using meta-analysis. Ecology 97:594-604.
- Ameel, J. J., Axler, R. P., and Owen, C. J. 1993. Persulfate digestion forsetermination of total nitrogen and phosphorus in low nutrient waters. Am. Environ. Lab. 10:1-11.
- Anschutz, P., Ciutat, A., Lecroart, P., Gérino, M., and Boudou, A. 2012. Effects of tubificid worm bioturbation on freshwater sediment biogeochemistry. Aquat. Geochem. 18:475-497.
- Bakker, E. S., Pagés, J. F, Arthur, R., and Alcoverro, T. 2016. Assessing the role of large herbivores in the structuring and functioning of freshwater and marine angiosperm ecosystems. – Ecography 39:162-179.
- Ballinger, A., and Lake, P. S. 2006. Energy and nutrient fluxes from rivers and streams into terrestrial food webs. Mar. Freshwater Res. 57:15-28.
- Belovsky, G.E. and Jordan, P.A. 1978. Time-energy budgets of moose. J. Theor. Biol. 14:76–104.
- Bergman, B. G., and Bump, J. K. 2014. Mercury in aquatic forage of large herbivores: Impact of environmental conditions, assessment of health threats, and implications for transfer across ecosystem compartments. Sci. Total Environ. 479:66-76.
- Bergman, B. G., and Bump, J. K. 2015. Experimental evidence that the ecosystem effects of aquatic herbivory by moose and beaver may be contingent on water body type. Freshwater Biol. 60:1635-1646.
- Bergström., A. K. 2010. The use of TN:TP and DIN:TP ratios as indicators for phytoplankton nutrient limitation in oligotrophic lakes affected by N deposition. Aquatic Sci. 72:277-281.
- Berman, T., and Chava, S. 1999. Algal growth on organic compounds as nitrogen sources. J. Plankton Res. 21:1423-1437.
- Blankson, E. R., and Klerks, P. L. 2016. The effect of bioturbation by Lumbriculus variegatus on transport and distribution of lead in a freshwater microcosm. Environ. Toxicol. Chem. *in press*.
- Bottinelli, N., Jouquet, P., Capowiez, Y., Podwojewski, P., Grimaldi, M., and Peng, X. 2015. Why is the influence of soil macrofauna on soil structure only considered by soil ecologists? Soil Tillage Res. 146:118-124.
- Bump, J.K., Peterson, R.O., and Vucetich, J.A. 2009a. Wolves modulate soil nutrient heterogeneity and 'This article is protected by copyright. All rights reserved.'

- foliar nitrogen by configuring the distribution of ungulate carcasses. Ecology 90:3159-3167.
- Bump, J. K., Tischler, K. B., Schrank, A. J., Peterson, R. O., and Vucetich, J. A. 2009b. Large herbivores and aquatic–terrestrial links in southern boreal forests. J. Anim. Ecol. 78:338-345.
- Bump, J.K., Webster, C. R., Vucetich, J.A, Peterson, R.O., Shields, J.M., and Powers, M.D. 2009c.
   Ungulate carcasses perforate ecological filters and create biogeochemical hotspots in forest herbaceous layers allowing trees a competitive advantage. Ecosystems 12:996-1007.
- Capps, K. A., Atkinson, C. L., and Rugenski, A. T. 2015. Consumer driven nutrient dynamics in freshwater ecosystems: an introduction. Freshwater Biol. 60:439-442.
- Ceacero, F., Landete-Castillejos, T., Miranda, M., García, A. J., Martínez, A., and Gallego, L. 2014. Why do cervids feed on aquatic vegetation? Behav. Process. 103:28-34.
- Chaffin, J.D. and Kane, D.D. 2010. Burrowing mayfly (Ephemeroptera: Ephemeridae: *Hexagenia spp.*) bioturbation and bioirrigation: a source of internal phosphorus loading in Lake Erie. J. Great Lakes Res. 36:57-63.
- Christenson, L. M., M. J. Mitchell, P. M. Groffman, and G. M. Lovett. 2014. Cascading effects of climate change on forest ecosystems: biogeochemical links between trees and moose in the northeast USA. Ecosystems 17:442-457.
- Christiansen, N. H., Andersen, F. Ø., and Jensen, H. S. 2016. Phosphate uptake kinetics for four species of submerged freshwater macrophytes measured by a 33 P phosphate radioisotope technique. Aquat. Bot. 128:58-67.
- Clifford, A. B. and Lawrence M. W. 2004. Case studies in novel narial anatomy: 2. The enigmatic nose of moose (Artiodactyla: Cervidae: Alces alces). J. Zool. 262:339-360.
- Cotner J.B., and Wetzel, R.G. 1992. Uptake of dissolved inorganic and organic phosphorus compounds by phytoplankton and bacterioplankton. Limnol. Oceanogr. 37:232-243.
- Darwin, C. 1881. The Formation of Vegetable Mould Through the Action of Worms With Observation of Their Habits. John Murray.
- Danell, K., Bergström, R., Duncan, P., and Pastor, J. 2006. Large herbivore ecology, ecosystem dynamics and conservation. Cambridge University Press.
- Davidson, A.D., Detling, J.K., and J.H. Brown. 2012. Ecological roles and conservation challenges of social, burrowing, herbivorous mammals in the worlds grassland. Front. Ecol. Environ. 10:477-486.
- Deocampo, D. M. 2002. Sedimentary structures generated by Hippopotamus amphibius in a lake-margin wetland, Ngorongoro Crater, Tanzania. Palaios, 17:212-217.

- Doughty, C. E., Roman, J., Faurby, S., Wolf, A., Haque, A., Bakker, E. S., Malhi, Y., Dunning, J. B. and Svenning, J. C. 2015. Global nutrient transport in a world of giants. P. Natl. Acad. Sci. USA 113:868-873.
- Elser, J. J., Bracken, M. E.S., Cleland, E. E., Gruner, D. S., Harpole, W. S., Hillebrand, H., Ngai, J. T., Seabloom, E. W., Shurin, J. B. and Smith, J. E. 2007. Global analysis of nitrogen and phosphorus limitation of primary producers in freshwater, marine and terrestrial ecosystems. Ecol. Lett. 10:1135–1142.
- Ford, A. T., and Goheen, J. R. 2015. Trophic cascades by large carnivores: a case for strong inference and mechanism. Trends Ecol. Evol. 30:725-735.
- Frank, D. A. 2008. Ungulate and topographic control of nitrogen: phosphorus stoichiometry in a temperate grassland; soils, plants and mineralization rates. Oikos 117:591-601.
- Franklin, J. F. 2005. Spatial pattern and ecosystem function: reflections on current knowledge and future directions. In: Lovett G.M., Jones C. G., Turner, M. G., and Weathers, K. C. (eds.), Ecosystem function in heterogeneous landscapes. Springer, pp. 427-442.
- Gabet, E. J., Reichman, O. J., and Seabloom, E. W. 2003. The effects of bioturbation on soil processes and sediment transport. Annu. Rev. Earth Pl. Sc. 31:249-273.
- Gereta, E., and Wolanski, E. 1998. Wildlife-water quality interactions in the Serengeti National Park,

  Tanzania. Afr. J. Ecol. 36:1-14.
- Gordeev, M. I., and Sibataev, A. K. 1995. Influence of predatory plant bladderwort (Utricularia vulgaris) on the process of selection in malaria mosquito larvae. Russ. J. Ecol+. 26:216-220.
- Greenville, A. C., Wardle, G. M., Tamayo, B., and Dickman, C. R. 2014. Bottom-up and top-down processes interact to modify intraguild interactions in resource-pulse environments. Oecologia 175:1349-1358.
- Hansen, H. L., Krefting, L. W., and Kurmis, V. 1973. The forest of Isle Royale in relation to fire history and wildlife. St. Paul: Agricultural Experiment Station, University of Minnesota.
- Harms, S. 2002. The effect of bladderwort (Utricularia) predation on microcrustacean prey. Freshwater Biol. 47:1608–1617.
- Hobbs, N. T. 1996. Modification of ecosystems by ungulates. J. Wildlife Manage. 60:695-713.
- Hölker, F., Vanni, M. J., Kuiper, J. J., Meile, C., Grossart, H. P., Stief, P., Adrian, R., Lorke, A., Dellwig,
  O., Brand, A., Hupfer, M., Mooij, W. M., Nützmann, and Lewandowski, J. 2015. Tube dwelling invertebrates: tiny ecosystem engineers have large effects in lake ecosystems. –
  Ecological Monogr. 85:333-351.

- Holt, R.D. 2008. Theoretical perspectives on resource pulses. Ecology 89: 671-681.
- Hood, G. A., and Larson, D. G. 2015. Ecological engineering and aquatic connectivity: a new perspective from beaver - modified wetlands. – Freshwater Biol. 60:198-208.
- Huber, N. K. 1975. The geologic story of Isle Royale National Park. US Government Printing Office.
- Janetski, D.J., D.T. Chaloner, S.D. Tiegs, and G.A. Lamberti. 2009. Pacific salmon effects on stream ecosystems: a quantitative synthesis. Oecologia 159:583-595.
- Jansson, M., Olsson, H., and Pettersson, K. 1988. Phosphatases; origin, characteristics and function in lakes. Hydrobiologia 170:157-175.
- Karlsson, J., Byström, P., Ask, J., Ask, P., Persson, L. and Jansson, M., 2009. Light limitation of nutrient-poor lake ecosystems. Nature 460:506-509.
- Kibriya, S., and Jones, J. I. 2007. Nutrient availability and the carnivorous habit in Utricularia vulgaris. Freshwater Biol. 52:500-509.
- Kristensen, E., Penha-Lopes, G., Delefosse, M., Valdemarsen, T., Quintana, C. O., and Banta, G. T. 2011.

  What is bioturbation? The need for a precise definition for fauna in aquatic sciences. Mar.

  Ecol-Progress Ser. 446:285-302.
- Larsen, J.A. 1980. The Boreal Ecosystem. Academic Press London.
- Likens, G.E. (ed.). 1985. An ecosystem approach to aquatic ecology: Mirror Lake and its environment. Springer-Verlag.
- Leroux, S.J., and Loreau, M. 2010. Consumer-mediated recycling and cascading trophic interactions. Ecology 91:2162-2171.
- Levine S.N., and Schindler, D.W. 1992. Modification of the N:P ratio in lakes by in situ processes. Limnol. Oceanogr. 37:917-935.
- Levinton, J. 1995. Bioturbators as ecosystem engineers: control of the sediment fabric, inter-individual interactions, and material fluxes. In: Jones, C. and Lawton, J. H. (eds.), Linking Species and Ecosystems. Springer, pp. 29-36.
- Marcarelli, A. M., Baxter, C. V., Mineau, M. M., and Hall Jr, R. O. 2011. Quantity and quality: unifying food web and ecosystem perspectives on the role of resource subsidies in freshwaters. Ecology 92:1215-1225.
- McCarthy, T. S., Ellery, W. N., and Bloem, A. 1998. Some observations on the geomorphological impact of hippopotamus (Hippopotamus amphibius L.) in the Okavango Delta, Botswana. Afr. J. Ecol. 36:44-56.

- McIntyre, P.B., Flecker, A.S., Vanni, M.J., Hood, J.M., Taylor, B.W., and Thomas, S.A. 2008. Fish distributions and nutrient cycling in streams: can fish create biogeochemical hotspots? Ecology 89:2335-2346.
- Mermillod-Blondin, F. 2011. The functional significance of bioturbation and biodeposition on biogeochemical processes at the water–sediment interface in freshwater and marine ecosystems.
   J. N. Am. Benthol. Soc. 30:770-778.
- Mermillod-Blondin, F., and Rosenberg, R. 2006. Ecosystem engineering: the impact of bioturbation on biogeochemical processes in marine and freshwater benthic habitats. Aquat Sci. 68:434-442.
- Meunier, C.L., Gundale, M.J., Sánchez, I.S. and Liess, A. 2016. Impact of nitrogen deposition on forest and lake food webs in nitrogen limited environments. Glob. Change Biol. 22:164-179.
- Meysman, F. J., Middelburg, J. J., and Heip, C. H. 2006. Bioturbation: a fresh look at Darwin's last idea.

   Trends Ecol. Evol. 21:688-695.
- Middelboe, A. L., and Markager, S. 1997. Depth limits and minimum light requirements of freshwater macrophytes. Freshwater Biol. 37:553-568.
- Moore, J. W. 2006. Animal ecosystem engineers in streams. BioScience 56:237-246.
- Mosepele, K., Moyle, P. B., Merron G. S., Purkey, D. R., and Mosepele, B. 2009. Fish, floods, and ecosystem engineers: aquatic conservation in the Okavango Delta, Botswana. BioScience 59:53-64.
- Moss, B. 2015. Mammals, freshwater reference states, and the mitigation of climate change. Freshwater Biol. 60:1964–1976.
- Nowlin, W. H., Vanni, M. J., and Yang, L. H. 2008. Comparing resource pulses in aquatic and terrestrial ecosystems. Ecology 89:647-659.
- Pastor, J., Naiman, R. J., Dewey, B., and McInnes, P. 1988. Moose, microbes, and the boreal forest. BioScience 38:770-777.
- Peterson, R. O., Vucetich, J. A., Bump, J. M., and Smith, D. W. 2014. Trophic cascades in a multicausal world: Isle Royale and Yellowstone. Annu. Rev. Ecol. Evol. S. 45:325-345.
- Pickett, S., Ostfeld, R. S., Shachak, M., and Likens, G. 1997. The ecological basis of conservation: heterogeneity, ecosystems, and biodiversity. Springer.
- Pickett, S.T. and White, P.S. eds., 2013. The ecology of natural disturbance and patch dynamics. Elsevier.
- Polis, G. A., Power, M. E., and Huxel, G. R. 2004. Food webs at the landscape level. University of Chicago Press.

- Reichman, O. J., and Seabloom, E. W. 2002. The role of pocket gophers as subterranean ecosystem engineers. Trends Ecol. Evol. 17:44-49.
- Remaili, T. M., Simpson, S. L., Amato, E. D., Spadaro, D. A., Jarolimek, C. V., and Jolley, D. F. 2016.

  The impact of sediment bioturbation by secondary organisms on metal bioavailability,
  bioaccumulation and toxicity to target organisms in benthic bioassays: Implications for sediment
  quality assessment. Environ. Pollut. 208:590-599.
- Ritchie, M. E., Tilman, D., and Knops, J. M. 1998. Herbivore effects on plant and nitrogen dynamics in oak savanna. Ecology 79:165-177.
- Schindler, D.W., Newbury, R.W., Beaty, K.G., and Campbell, P. 1976. Natural water and chemical budgets for a small Precambrian lake basin in central Canada. Journal of the Fisheries Research Board of Canada, 33:2526-2543.
- Schmitz, O.J., Hawlena, D., Trussell, G.C. 2010. Predator control of ecosystem nutrient dynamics. Ecology Letters 13:1199-1209.
- Singer, F. J., and Schoenecker, K. A. 2003. Do ungulates accelerate or decelerate nitrogen cycling? Forest Ecol. Manag. 181:189-204.
- Sinclair, A. R., Nkwabi, A. K., and Metzger, K. L. 2015. The Butterflies of Serengeti: Impact of Environmental Disturbance on Biodiversity. – In: Sinclair, A. R. E., Metzger, K. L., Mduma, S. A. R., and Fryxell, J. M. (eds.), Serengeti IV: Sustaining Biodiversity in a Coupled Human-Natural System, pp. 301-322.
- Søndergaard, M, Kristensen, P., and Jeppesen, E. 1992. Phosphorus release from resuspended sediment in the shallow and wind-exposed Lake Arresø, Denmark. Hydrobiologia 228:91-99.
- Søndergaard, M, Jensen, J. P, and Jeppesen, E. 2003. Role of sediment and internal loading of phosphorus in shallow lakes. Hydrobiologia 506:135-145.
- Stottlemyer R., Toczydlowski, D., Herrmann, R. 1998. Biogeochemistry of a mature boreal ecosystem:

  Isle Royale National Park, Michigan. Scientific Monograph NPS/NRUSGS/NRSM-98/01. –

  United States Department of the Interior, National Park Service.
- Subalusky, A. L., Dutton, C. L., Rosi Marshall, E. J., and Post, D. M. 2015. The hippopotamus conveyor belt: vectors of carbon and nutrients from terrestrial grasslands to aquatic systems in sub Saharan Africa. Freshwater Biol. 60:512-525.
- Takafumi, H., Matsumoto, A., Aotani, K., and Yoshida, T. 2015. The cross-ecosystem impact of deer on an endangered submerged macrophyte, Ranunculus nipponicus var. submersus. Glob. Ecol. Cons. 4:581-588.

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- Takimoto G, Iwata T, Murakami M. 2009. Timescale Hierarchy Determines the Indirect Effects of Fluctuating Subsidy Inputs on In Situ Resources. Am. Nat. 173:200-211
- Thayer, C. W. 1979. Biological bulldozers and the evolution of marine benthic communities. Science 203:458-461.
- Tischler, K.D. 2004. Aquatic plant nutritional quality and contribution to moose diet at Isle Royale

  National Park. Michigan Technological University.
- Turner, M.G., 2010. Disturbance and landscape dynamics in a changing world 1. Ecology 91:2833-2849.
- Turner, M. G., and Gardner, R. H. 2015. Ecosystem Processes in Heterogeneous Landscapes. In:

  Turner, M. G., and Gardner, R. H. (eds.), Landscape Ecology in Theory and Practice. Springer,

  pp. 287-332.
- Usio, N., and Townsend, C. R. 2004. Roles of crayfish: consequences of predation and bioturbation for stream invertebrates. Ecology 85:807-822.
- Valentine, J. F., Heck, K. L., Harper, P., and Beck, M. 1994. Effects of bioturbation in controlling turtlegrass (Thalassia testudinum Banks ex König) abundance: evidence from field enclosures and observations in the Northern Gulf of Mexico. J. Exp. Mar. Biol. Ecol. 178:181-192.
- Vanni, M. J. 2002. Nutrient cycling by animals in freshwater ecosystems. Annu. Rev. Ecol. Syst. 33:341-370.
- Vanni, M.J., Flecker, A.S., Hood, J.M. and Headworth, J.L. 2002. Stoichiometry of nutrient recycling by vertebrates in a tropical stream: linking species identity and ecosystem processes. – Ecol. Lett. 5:285-293.
- Vanni, M. J., Boros, G., and McIntyre, P. B. 2013. When are fish sources vs. sinks of nutrients in lake ecosystems? Ecology 94:2195-2206.
- Walker, L.R. 2012. The biology of disturbed habitats. Oxford University Press.
- Whalen, S.C., and Cornwell, J.C. 1985. Nitrogen, phosphorus, and organic carbon cycling in an arctic lake. Can. J. Fish. Aquatic Sci. 42:797-808.
- Wetzel, R. G. 1992. Uptake of dissolved inorganic and organic phosphorus compounds by phytoplankton and bacterioplankton. Limnol. Oceanogr. 37:232-243.
- Wilkinson, M. T., Richards, P. J., and Humphreys, G. S. 2009. Breaking ground: pedological, geological, and ecological implications of soil bioturbation. Earth-Sci. Rev. 97:257-272.
- Wilmers, C.C., and Getz, W.M. 2004. Simulating the effects of wolf-elk population dynamics on resource flow to scavengers in Yellowstone National Park. Ecol. Model. 177: 193-208.

- Winter, T.C. and Likens, G.E. 2009. Mirror Lake: Interactions among Air, Land, and Water. University of California Press.
- Wisdom, M. J., Vavra, M., Boyd, J. M., Hemstrom, M. A., Ager, A. A., and Johnson, B. K. 2006.
  Understanding ungulate herbivory-episodic disturbance effects on vegetation dynamics:
  knowledge gaps and management needs. Wildlife Soc. B. 34:283-292.
- Wolanski, E., and Gereta, E. 1999. Oxygen cycle in a hippo pool, Serengeti National Park, Tanzania. Afr. J. Ecol. 37:419-423.
- Yang, L. H., Bastow, J. L., Spence, K. O., and Wright, A. N. 2008. What can we learn from resource pulses. Ecology 89:621-634.
- Yang, L.H., Edwards, K. F., Byrnes, J. E., Bastow, J. L., Wright, A. N., and Spence, K. O. 2010. A metaanalysis of resource pulse-consumer interactions. – Ecol. Monogr. 80:125-151.
- Zar, J. H. 2010. Biostatistical analysis. Pearson Education.

FIG. 1







