Divergent selection in low and high elevation populations of a perennial herb in the Swiss Alps

Esther R. Frei · Thomas Hahn · Jaboury Ghazoul · Andrea R. Pluess

Abstract Mountain plant species with wide elevational ranges are expected to be exposed to different selection pressures at low and high elevation. Estimating the extent of genetic population differentiation contributes to our understanding of selective forces shaping phenotypic variation of plants in response to changing climate conditions. Using a common garden experiment, we measured narrow-sense heritability ($h^2$) and quantitative trait differentiation ($Q_{ST}$) in several growth-related, reproductive and phenological traits among low and high elevation populations of the semi-dry grassland species *Ranunculus bulbosus* from the Swiss Alps. $Q_{ST}$ values were compared to neutral genetic differentiation ($F_{ST}$) based on AFLP markers and indicated divergent selection in most traits among all populations as well as among low and high elevation populations separately. Furthermore, pairwise $Q_{ST}$ and $F_{ST}$ estimates were not correlated suggesting that neutral marker differentiation is not a valuable proxy for quantitative trait differentiation. Neutral molecular differentiation increased with geographical distance whereas quantitative genetic differentiation did not follow an isolation by distance pattern. Plant traits did mostly not differ among low and high populations and pairwise $Q_{ST}$–$F_{ST}$ comparisons indicated no divergent selection between the two elevations. Differing levels of trait differentiation in low and high elevation populations, however, indicated that different selective forces acted on plants at contrasting elevations suggesting that selection regimes might alter under climate warming.

Keywords AFLP markers · Climate change · Isolation by distance · Mantel correlations · Narrow-sense heritability · Phenotypic traits · $Q_{ST}$–$F_{ST}$ comparisons

Introduction

In mountain habitats strong environmental variation over relatively short geographical distances (Körner 2003) is associated with highly divergent selective pressures on plant species and the emergence of locally adapted traits (Linhart and Grant 1996; Kawecki and Ebert 2004). Both abiotic factors, such as temperature, UV radiation, precipitation, and substrate, as well as biotic factors, such as competition, vary across elevations and potentially act as selective agents leading to plant differentiation (Choler et al. 2001; Till-Bottraud and Gaudeul 2002; Körner 2003; Blume et al. 2010). Besides the important role of natural selection in shaping phenotypic variation, neutral genetic processes, such as genetic drift, gene flow and random mutation, are also likely to influence population differentiation (Wright 1951; Slatkin 1977; Lande 1992). On the one hand, gene flow through pollen or seed dispersal is facilitated by the close proximity of populations along elevational gradients
and has the potential to counteract the effects of divergent selection in populations, and thus to prevent local adaptation (reviewed in Slatkin 1987; Lenormand 2002; Raesaenen and Hendry 2008). Genetic drift, on the other hand, can induce random genetic variation through random changes in allele frequencies, especially in small populations (Spitze 1993; Lynch and Milligan 1994).

The relative importance of selection and non-selective neutral processes for among-population differentiation in quantitative traits can be evaluated by comparing quantitative genetic differentiation ($Q_{ST}$; Spitze 1993) with neutral marker differentiation ($F_{ST}$; Wright 1951). Higher relative divergence in the quantitative traits than in neutral markers (i.e. $Q_{ST} < F_{ST}$) indicates stabilizing selection favouring different genotypes in different populations whereas the opposite (i.e. $Q_{ST} > F_{ST}$) indicates divergent selection favouring similar plant trait values are selected for. If the two measures do not significantly differ (i.e. $Q_{ST} = F_{ST}$), neutral processes (drift, migration and mutation) alone may be sufficient to explain patterns of differentiation, although the relative effects of drift and natural selection on population differentiation cannot be separated (Merilä and Crnokrak 2001; McKay and Latta 2002; O’Hara and Merilä 2005; Whitlock 2008). Studies comparing $Q_{ST}$ and $F_{ST}$ allow eliminating neutral genetic processes as an explanation for divergence and have already provided valuable insights into plant trait responses to environmental heterogeneity (reviewed in McKay and Latta 2002; Leinonen et al. 2008; De Kort et al. 2013). For a more comprehensive understanding of the nature of selection, however, additional measures are needed, such as correlations of divergence with environmental differences (Whitlock 2008) or spatial patterns of differentiation, i.e. isolation by distance (IBD; Hutchison and Templeton 1999). Non-adaptive isolation by distance patterns may arise from decreasing homogenizing effects of gene flow with increasing geographical distance and lead to phenotypic divergence among geographically distant populations. The relative contributions of selection and neutral processes should then also depend on the spatial correlation between the selective environment and the geographical distance between populations.

Mountain ecosystems are considered vulnerable to the impacts of land use change (Fischer et al. 2008) and a rapidly changing climate (Begert et al. 2005; IPCC 2013). Owing to steep environmental gradients, they are particularly suited to investigate population differentiation and diversity (Till-Bottraud and Gaudeul 2002). Transplant experiments along elevational gradients have revealed local adaptation in fitness-related traits (Byars et al. 2007; Gonzalo-Turpin and Hazard 2009) and in leaf and reproductive phenological traits (Scheepens et al. 2010; Alberto et al. 2011). While several species have shown neutral genetic differentiation across elevational gradients (reviewed in Ohsawa and Ide 2008), only little differentiation was found between low and high elevation populations of three semi-dry nutrient-poor grassland species, probably due to high gene flow (Hahn et al. 2012; Matter et al. 2013). Yet, genetic differentiation was slightly greater among high than among low elevation populations (Hahn et al. 2012) as shown also for other species (Young et al. 2002; Herrera and Bazaga 2008; Byars et al. 2009) suggesting that gene flow underlies different processes at high and low elevation. Likewise, differentiation in quantitative traits and the magnitude of heritability might change along elevational gradients. Understanding the interplay of natural selection and neutral genetic processes as well as their variation at low and high elevations sheds light on the driving forces behind intraspecific differentiation and will gain in importance in the context of global change.

In this study, we assessed how natural selection and neutral processes influence genetic differentiation in low and high elevation populations of *Ranunculus bulbosus*. In a common garden experiment we grew seed families of *R. bulbosus* from 14 populations originating at 1,200 m and 1,800 m a.s.l. Vegetative and reproductive traits as well as flowering phenological traits were measured. Subsequently, narrow-sense heritability ($h^2$) and quantitative trait differentiation ($Q_{ST}$) among all populations, as well as among low and high elevation populations separately, were determined. Neutral genetic differentiation ($F_{ST}$) was calculated based on amplified fragment length polymorphism (AFLP) marker data of a previous project, which analysed genetic variation within and among a larger number of populations of *R. bulbosus* (Hahn et al. 2012) whereas our molecular analysis was based on a subset identical with the seed sampling populations.

Comparisons of overall $Q_{ST}$ and $F_{ST}$ estimates allowed testing the null hypothesis of neutrality in each trait. Furthermore, by comparing pairwise $Q^i_{ST}$ and $F^i_{ST}$ estimates of all low with all high elevation populations, we tested whether there was divergent selection between low and high elevation populations. Mantel correlations of pairwise $Q^i_{ST}$ and $F^i_{ST}$ estimates were used to test whether selection and drift are acting in the same direction (e.g. Leinonen et al. 2008). Finally, Mantel correlations between geographical distance and neutral molecular marker as well as quantitative trait differentiation were used to investigate isolation by distance patterns (Hutchison and Templeton 1999).

### Materials and methods

#### Study species and plant material

The perennial herb *Ranunculus bulbosus* L. is a characteristic plant of semi-dry nutrient-poor calcareous grasslands
ranging from 400 to 2,000 m a.s.l (colline to subalpine) in the European Alps with a distribution center in the montane belt (Aeschimann et al. 2004). *Ranunculus bulbosus* is an early-flowering insect-pollinated geophytic herb (Coles 1973; Sarukhan and Harper 1973; Steinbach and Gottsberger 1994).

In summer 2008, seeds were sampled from seven regions across the Swiss Alps, each containing one low elevation population at 1,200 m a.s.l. and one high elevation population at 1,800 m a.s.l. resulting in a total of 14 populations (Fig. 1; Table S1). The horizontal distance between the two sampling locations of a region was 1–18 km (median: 4.5 km; Table 1). Population sizes and plant densities at low and high elevation were similar (Hahn et al. 2012). Seeds of nine maternal plants (seed families) of each population were collected at a distance of at least four metres among individuals. They were air-dried and stored in paper bags at 4 °C. In spring 2009, seeds were germinated in a greenhouse at the ETH Zurich field station Lindau/Eschikon. Plants were raised in randomly arranged individual pots (800 cm$^3$), filled with a 3:2 mixture of nutrient-poor commercial soil and sand.

**Fig. 1** Fourteen populations of *Ranunculus bulbosus* sampled across the Swiss Alps. *Open triangles* represent low elevation populations (1,200 m a.s.l) and *filled triangles* high elevation populations (1,800 m a.s.l). Regions are encoded corresponding to Table S1. Map copyright: © swisstopo

**Table 1** Geographical distances in km (below the diagonal) and pairwise $F_{ST}$ (above the diagonal) for 14 populations of *Ranunculus bulbosus*

<table>
<thead>
<tr>
<th></th>
<th>Chat12</th>
<th>Chat18</th>
<th>Hasl12</th>
<th>Hasl18</th>
<th>Luet12</th>
<th>Luet18</th>
<th>Matt12</th>
<th>Matt18</th>
<th>Obwa12</th>
<th>Obwa18</th>
<th>Unwa12</th>
<th>Unwa18</th>
<th>Valhe12</th>
<th>Valhe18</th>
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<tbody>
<tr>
<td>Chat12</td>
<td>–</td>
<td>0.047</td>
<td>0.116</td>
<td>0.101</td>
<td>0.102</td>
<td>0.116</td>
<td>0.098</td>
<td>0.131</td>
<td>0.131</td>
<td>0.129</td>
<td>0.091</td>
<td>0.114</td>
<td>0.091</td>
<td>0.099</td>
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<td>–</td>
<td>0.130</td>
<td>0.113</td>
<td>0.099</td>
<td>0.127</td>
<td>0.077</td>
<td>0.087</td>
<td>0.108</td>
<td>0.117</td>
<td>0.074</td>
<td>0.091</td>
<td>0.091</td>
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<td>89.391</td>
<td>–</td>
<td>0.101</td>
<td>0.081</td>
<td>0.130</td>
<td>0.164</td>
<td>0.176</td>
<td>0.174</td>
<td>0.181</td>
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<td>0.164</td>
<td>0.156</td>
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<td>Hasl18</td>
<td>89.553</td>
<td>88.346</td>
<td>1.648</td>
<td>–</td>
<td>0.077</td>
<td>0.080</td>
<td>0.175</td>
<td>0.174</td>
<td>0.191</td>
<td>0.182</td>
<td>0.146</td>
<td>0.159</td>
<td>0.148</td>
<td>0.148</td>
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<tr>
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<td>67.023</td>
<td>65.618</td>
<td>23.773</td>
<td>22.753</td>
<td>–</td>
<td>0.030</td>
<td>0.163</td>
<td>0.153</td>
<td>0.187</td>
<td>0.176</td>
<td>0.137</td>
<td>0.157</td>
<td>0.142</td>
<td>0.141</td>
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<td>67.076</td>
<td>65.756</td>
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<td>22.591</td>
<td>0.793</td>
<td>–</td>
<td>0.193</td>
<td>0.193</td>
<td>0.205</td>
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<td>0.181</td>
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<td>0.175</td>
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<td>50.115</td>
<td>59.694</td>
<td>59.497</td>
<td>41.532</td>
<td>42.218</td>
<td>–</td>
<td>0.049</td>
<td>0.077</td>
<td>0.094</td>
<td>0.069</td>
<td>0.104</td>
<td>0.075</td>
<td>0.049</td>
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<td>58.736</td>
<td>44.143</td>
<td>44.905</td>
<td>10.775</td>
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<td>0.072</td>
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<td>0.087</td>
<td>0.068</td>
<td>0.035</td>
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<td>62.976</td>
<td>62.551</td>
<td>42.407</td>
<td>42.990</td>
<td>9.164</td>
<td>19.912</td>
<td>–</td>
<td>0.040</td>
<td>0.091</td>
<td>0.106</td>
<td>0.086</td>
<td>0.069</td>
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<td>39.387</td>
<td>62.455</td>
<td>61.977</td>
<td>41.434</td>
<td>41.993</td>
<td>10.899</td>
<td>21.673</td>
<td>2.147</td>
<td>–</td>
<td>0.068</td>
<td>0.087</td>
<td>0.067</td>
<td>0.060</td>
</tr>
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<td>Unwa12</td>
<td>52.399</td>
<td>46.000</td>
<td>115.103</td>
<td>114.602</td>
<td>93.432</td>
<td>93.909</td>
<td>57.411</td>
<td>63.787</td>
<td>52.248</td>
<td>52.653</td>
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<td>0.043</td>
<td>0.055</td>
<td>0.044</td>
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<td>Unwa18</td>
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<td>30.513</td>
<td>99.103</td>
<td>98.495</td>
<td>76.800</td>
<td>77.232</td>
<td>43.872</td>
<td>52.053</td>
<td>37.187</td>
<td>37.164</td>
<td>17.647</td>
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<td>0.070</td>
<td>0.070</td>
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<td>Valhe12</td>
<td>38.040</td>
<td>30.832</td>
<td>85.757</td>
<td>85.217</td>
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<td>38.606</td>
<td>23.333</td>
<td>23.446</td>
<td>29.466</td>
<td>13.921</td>
<td>–</td>
<td>0.030</td>
</tr>
<tr>
<td>Valhe18</td>
<td>47.928</td>
<td>40.683</td>
<td>88.944</td>
<td>88.577</td>
<td>68.475</td>
<td>69.037</td>
<td>30.135</td>
<td>36.442</td>
<td>26.139</td>
<td>27.045</td>
<td>27.442</td>
<td>17.486</td>
<td>10.016</td>
<td>–</td>
</tr>
</tbody>
</table>

Population-pairwise $F_{ST}$ values were calculated with Arlequin 3.5 (Excoffier et al. 2005).
Leaf material for the molecular analyses was collected from the same populations but from different mother plants than the seeds for the common garden experiment (Table S1). Circa 30 plants of each population were sampled with distances among individuals of at least four metres. Sampled leaf material was immediately dried with silica gel.

Quantitative genetic data

In March 2010, a common garden experiment was set up to determine quantitative genetic differentiation ($Q_{ST}$) among populations. In this experiment, plants were cultivated under controlled environmental conditions using a half-sib design (Evanno et al. 2006), and thus the variation of phenotypes reflected their genetic differences (Briggs and Walters 1997; Petit et al. 2001). Three to four half-sibs of each of the nine $R. bulbosus$ seed families per population (440 individuals in total) were arranged randomly in the greenhouse and rotated weekly.

At the start of the experiment, initial plant size was assessed as number of leaves, and length and width of the longest leaf. The product of them was used as covariate. From April to June 2010, flowering phenology of each plant was monitored weekly to record the date of the first open flower, the last open flower and the first mature fruit. Flowering duration was calculated as number of days between the first and last open flower of a plant. In July 2010, flowers were counted and length of the longest flower stalk was measured. In October 2010, length of the longest leaf was measured and specific leaf area (SLA) determined. The fresh leaf blades were scanned (HP All-In-One colour Scanner, Hewlett-Packard GmbH, Düben-dorf, Switzerland) and their area was determined using LAMINA version 1.0.2 (Bylesjoe et al. 2008). The scanned leaf blades were weighed after oven drying at 60 °C for 48 h. SLA was calculated as the ratio of leaf area to dry weight of the leaf blade (Cornelissen et al. 2003). Moreover, leaf and bulb biomass were harvested, oven-dried and weighed.

Molecular genetic data

Population differentiation based on neutral genetic processes was determined using putatively neutral amplified fragment length polymorphism (AFLP) markers (Vos et al. 1995). For further details of genotyping, see Hahn et al. (2012).

Only polymorphic loci and loci with an allele frequency smaller than $1 – (3/N)$, where $N$ is the total number of sampled individuals, were included in the analysis (Lynch and Milligan 1994). The data set for all populations comprised 125 loci, the separate sets for low elevation populations 121 loci and high elevation populations 122 loci. Because AFLP markers might also be part of or linked to coding gene regions, they might be under selection (Stinchcombe and Hoekstra 2008). Such markers need to be excluded because $Q_{ST} – F_{ST}$ comparisons assume that $F_{ST}$ reflects neutral processes. A Bayesian outlier analysis in BayeScan 2.1 (Foll and Gaggiotti 2008), with 5,000 iterations and a thinning interval of 10, revealed that no loci were under divergent selection. The same result was previously also found for a slightly larger dataset (Hahn et al. 2012). All loci were therefore kept for subsequent analyses.

Statistical analyses

All statistical analyses were performed in R 2.15.0 (R Development Core Team 2012) with a full data set including all 14 populations as well as two data subsets including low and high elevation populations separately. To estimate between and within-population variances of quantitative traits, we ran linear mixed-effects models using the function lmer and the restricted maximum likelihood estimation (REML) in the R package lme4 (Bates et al. 2012). To meet the assumptions of normality of residuals and homoscedasticity, variables were transformed appropriately. The models included ‘population’ and ‘seed family’ nested within ‘population’ as random factors. Because data showed strong correlations with initial plant size, we used this variable as fixed covariate (see also Chun et al. 2011). Moreover, based on the full data set, we assessed genetic differentiation between low and high elevation plants using the same model with the addition of ‘altitude of origin’ as fixed factor.

Estimation of heritability, $Q_{ST}$ and $F_{ST}$

Narrow-sense heritabilities ($h^2$) were calculated according to Petit et al. (2001) as

$$h^2 = \frac{V_A}{V_A + V_e}$$

where $V_A$ is the additive genetic variance, which is the average genetic variance within populations. It was estimated by $V_A \approx 4 \cdot V_{FAM}$ where $V_{FAM}$ is the variance among seed families (Petit et al. 2001). Quantitative genetic differentiation ($Q_{ST}$) for each trait was calculated as

$$Q_{ST} = \frac{V_{POP}}{2V_{GEN} + V_{POP}}$$

where $V_{POP}$ is the among-population variance. Accounting for the half-sib design, $V_{GEN}$ was estimated by $V_{GEN} = V_A \approx 4 \cdot V_{FAM}$. The 95% confidence intervals of $h^2$ and $Q_{ST}$ estimates were calculated using a jack-knife procedure over seed families.
As a measure of neutral molecular differentiation, $F_{ST}$ was calculated (Wright 1951). Single locus $F_{ST}$ values were determined using BayeScan 2.1 (Foll and Gaggiotti 2008) and a bootstrap approach over loci with 5,000 iterations was applied to estimate the 95 % CI of the mean $F_{ST}$ across loci.

Overall $Q_{ST} – F_{ST}$ comparisons

To test whether quantitative trait differentiation was significantly different from neutral genetic differentiation, an approach similar to the one proposed by Whitlock and Guillaume (2009) was applied. Analogously to an equivalence test, the difference of each resampled $Q_{ST}$ (based on the distribution of the jack-knife procedure over seed families) and resampled single marker $F_{ST}$ was calculated. After repeating this 126 times (equivalent to the total number of seed families), the sampling distribution of $Q_{ST} – F_{ST}$ was generated. $Q_{ST}$ is significantly different from its expected value under neutral divergence among populations ($F_{ST}$) if the 95 % confidence interval of the $Q_{ST} – F_{ST}$ estimate does not contain zero.

Pairwise comparisons and Mantel tests

Population-pairwise $Q_{ST}$ values were calculated using Eq. 2, but with the among-population variance $V_{POP}$ calculated as the variance of the estimates of the population means of the two populations, i.e.

$$V_{POP} = \frac{c^2}{2}$$

where $c$ is the contrast between the two populations in a mixed-effects model containing ‘population’ as a fixed effect (Palo et al. 2003; personal communication B. O’Hara). Population-pairwise $F_{ST}$ values were calculated with Arlequin 3.5 (Excoffier et al. 2005).

First, population-pairwise $Q_{ST}^{ij}$ values were compared to pairwise $F_{ST}^{ij}$ in all low vs. high elevation population pairs as well as in all low vs. low and high vs. high pairs. Second, correlations between geographical distance, genetic ($F_{ST}$) and phenotypic ($Q_{ST}$) components were analysed using Mantel tests (Mantel 1967; Antoniazza et al. 2010). All Mantel tests were computed based on Legendre and Legendre (2012) with the R package vegan (Oksanen et al. 2013) and 1,000 permutations to assess the significance of the Mantel correlation coefficients ($r$). For Mantel tests with geographical distances, $F_{ST}^{ij}/(1 – F_{ST}^{ij})$ respectively $Q_{ST}^{ij}/ (1 – Q_{ST}^{ij})$ were used while geographical distances were log-transformed (Rousset 1997).

Results

Plant growth and phenology

Based on the results of linear mixed-effects models, none of the investigated quantitative traits showed genetic differentiation between altitudes of origin ($P > 0.05$; Table 2), except for bulb biomass, which was greater in low than high elevation plants ($P = 0.01$). The variances due to the random effects population and seed family accounted for 10–72 % and 4–20 %, respectively, in the different traits.

Heritability, $Q_{ST}$ and $F_{ST}$

The observed narrow-sense heritabilities ($h^2$) ranged from 0.291 to 0.675 in all populations (Table 3). Heritability of low and high elevation plants was similar in leaf length, bulb biomass, flowering start and flowering duration, but differed in the other six traits: SLA, number of flowers and fruiting start had larger $h^2$ whereas number of leaves, leaf biomass

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Table 2  Trait means and standard errors (SE) grouped by altitude of origin, as well as the effect of altitude of origin ($P$) for the quantitative traits of *Ranunculus bulbosus* in the common garden experiments

<table>
<thead>
<tr>
<th>Trait</th>
<th>Mean Orig1200</th>
<th>SE Orig1200</th>
<th>Mean Orig1800</th>
<th>SE Orig1800</th>
<th>$P$ Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number of leaves</td>
<td>12.66</td>
<td>0.46</td>
<td>13.28</td>
<td>0.51</td>
<td>0.74</td>
</tr>
<tr>
<td>Leaf length (cm)</td>
<td>10.34</td>
<td>0.21</td>
<td>9.53</td>
<td>0.18</td>
<td>0.09</td>
</tr>
<tr>
<td>Leaf biomass (g)</td>
<td>0.34</td>
<td>0.01</td>
<td>0.32</td>
<td>0.01</td>
<td>0.71</td>
</tr>
<tr>
<td>Bulb biomass (g)</td>
<td>0.60</td>
<td>0.02</td>
<td>0.40</td>
<td>0.02</td>
<td>0.01*</td>
</tr>
<tr>
<td>SLA (g/cm²)</td>
<td>243.95</td>
<td>2.47</td>
<td>248.26</td>
<td>3.18</td>
<td>0.48</td>
</tr>
<tr>
<td>Number of flowers</td>
<td>16.69</td>
<td>0.43</td>
<td>17.60</td>
<td>0.42</td>
<td>0.44</td>
</tr>
<tr>
<td>Flower stalk length (cm)</td>
<td>55.75</td>
<td>0.97</td>
<td>47.26</td>
<td>0.73</td>
<td>0.05(*)</td>
</tr>
<tr>
<td>Flowering start (JD)</td>
<td>103.81</td>
<td>0.68</td>
<td>107.93</td>
<td>0.69</td>
<td>0.45</td>
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<tr>
<td>Fruiting start (JD)</td>
<td>111.54</td>
<td>0.69</td>
<td>111.54</td>
<td>0.68</td>
<td>0.41</td>
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<tr>
<td>Flowering duration (days)</td>
<td>33.76</td>
<td>1.24</td>
<td>33.63</td>
<td>1.31</td>
<td>0.93</td>
</tr>
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</table>

$P$-values based on linear mixed-effect models including ‘plant size’ as covariate, ‘altitude of origin’ as fixed effect, as well as the random effects ‘population’ and ‘seed family’. Significance levels for altitude of origin (corrected for anti-conservative $P$-values): (*) $P < 0.05$; * $P < 0.01$; ** $P < 0.001$; *** $P < 0.0001$. Orig1200, low elevation populations (1200 m a.s.l.). Orig1800, high elevation populations (1800 a.s.l.)
and flower stalk length had smaller $h^2$ in low as compared to high elevation populations.

The observed $Q_{ST}$ values ranged from 0.103 for leaf length to 0.474 for flowering start in all populations (Fig. 2; Table S2). All traits showed significant among-population differentiation ($Q_{ST} > 0$) except for SLA in low elevation plants ($Q_{ST} = 0.003; 95 \%$ confidence interval (CI): [0.000, 0.007]). $Q_{ST}$ values were larger in five (leaf biomass, bulb biomass, flower stalk length, flowering start and fruiting start) and smaller in three traits (SLA, number of flowers and flowering duration) in low as compared to high elevation populations. $Q_{ST}$ values of number of leaves and leaf length did not differ between altitudes of origin.

From the AFLP data, we obtained an $F_{ST}$ value, averaged over loci, of 0.085 (CI: [0.080, 0.091]) in all populations. The $F_{ST}$ of low elevation populations ($F_{ST} = 0.093; CI: [0.090, 0.097]$) was similar to the $F_{ST}$ of high elevation populations ($F_{ST} = 0.095; CI: [0.092, 0.100]$).

Overall $Q_{ST}$–$F_{ST}$ comparisons

Considering the full data set with all populations, overall $Q_{ST}$ exceeded $F_{ST}$ in all measured traits with estimates ranging from 0.019 to 0.389 (average: 0.189; Fig. 3; Table S3). Also in the two subsets, $Q_{ST}$ exceeded $F_{ST}$ in all traits except for both subsets in leaf length and bulb biomass as well as SLA of low elevation populations. $Q_{ST}$–$F_{ST}$ estimates in low elevation populations ranged from $-0.090$ to $0.447$ with an average of $0.184$ and in high elevation populations from $-0.078$ to $0.392$ with an average of $0.183$. In most traits, except for number of leaves and leaf length, $Q_{ST}$–$F_{ST}$ estimates differed between low and high elevation populations indicated by non-overlapping CI’s (Fig. 3; Table S3).

Pairwise $Q_{ST}^{ij}$ and $F_{ST}^{ij}$ comparisons and Mantel tests

Pairwise $Q_{ST}^{ij}$ estimates ranged from 0.030 to 0.205 (Table 1). Considering pairwise $Q_{ST}^{ij}$ and $F_{ST}^{ij}$ estimates of all low with all high elevation populations, as well as all low vs. low and high vs. high pairs, $Q_{ST}^{ij}$ did not differ from $F_{ST}^{ij}$ in any of the measured traits (Table S4). Pairwise $Q_{ST}^{ij}$ values were not correlated with pairwise $F_{ST}^{ij}$ values in any of the investigated traits ($P > 0.23$; Table 4A). Pairwise $Q_{ST}^{ij}$ were not correlated with geographical distance in any of the investigated traits ($P > 0.21$; Table 4B). In contrast, pairwise $F_{ST}^{ij}$ were positively correlated with geographical distance in all populations (Mantel’s $r = 0.56$, $P < 0.01$; Tables 2 and 4C), as well as in the subsets of low (Mantel’s $r = 0.49$, $P < 0.05$) and high elevation populations (Mantel’s $r = 0.60$, $P < 0.05$).

Discussion

The main objective of this study was to evaluate the relative importance of natural selection vs. genetic drift in determining population differentiation in low and high populations in a semi-dry grassland species. Random genetic drift is expected to affect all loci equally. Thus, we used AFLP markers and confirmed their selective neutrality in our data set, which was in line with the findings of Hahn et al. (2012). Since the overall $Q_{ST}$–$F_{ST}$ comparisons may not adequately reflect variation in evolutionary forces acting at the inter-population level (Gutiérrez 2008), we combined this approach with population-pairwise $Q_{ST}^{ij}$–$F_{ST}^{ij}$ comparisons.

Table 3 Observed narrow-sense heritabilities ($h^2$) with their 95 \% confidence intervals (CI) for growth, reproductive and phenological traits of Ranunculus bulbosus

<table>
<thead>
<tr>
<th>Trait</th>
<th>All</th>
<th>Orig1200</th>
<th>Orig1800</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>$h^2$ CI 2.5% CI 97.5%</td>
<td>$h^2$ CI 2.5% CI 97.5%</td>
<td>$h^2$ CI 2.5% CI 97.5%</td>
</tr>
<tr>
<td>Number of leaves</td>
<td>0.4164 0.3922 0.4328</td>
<td>0.2467 0.2041 0.2876</td>
<td>0.5156 0.4960 0.5325</td>
</tr>
<tr>
<td>Leaf length</td>
<td>0.3975 0.3821 0.4097</td>
<td>0.3847 0.3591 0.4077</td>
<td>0.4236 0.3888 0.4382</td>
</tr>
<tr>
<td>Leaf biomass</td>
<td>0.4590 0.4364 0.4708</td>
<td>0.2929 0.2600 0.3100</td>
<td>0.5816 0.5445 0.5969</td>
</tr>
<tr>
<td>Bulb biomass</td>
<td>0.5278 0.5139 0.5405</td>
<td>0.5316 0.5138 0.5448</td>
<td>0.5349 0.5073 0.5576</td>
</tr>
<tr>
<td>SLA</td>
<td>0.4303 0.4087 0.4458</td>
<td>0.5319 0.5042 0.5453</td>
<td>0.3100 0.2588 0.3445</td>
</tr>
<tr>
<td>Number of flowers</td>
<td>0.3935 0.3678 0.4112</td>
<td>0.5171 0.5085 0.5413</td>
<td>0.2665 0.2213 0.2991</td>
</tr>
<tr>
<td>Flower stalk length</td>
<td>0.4276 0.4019 0.4438</td>
<td>0.3358 0.0000 0.3535</td>
<td>0.4999 0.4604 0.5230</td>
</tr>
<tr>
<td>Flowering start</td>
<td>0.6747 0.6632 0.6834</td>
<td>0.6765 0.6585 0.6884</td>
<td>0.6782 0.6377 0.6923</td>
</tr>
<tr>
<td>Fruiting start</td>
<td>0.6453 0.6346 0.6540</td>
<td>0.7137 0.6959 0.7265</td>
<td>0.5917 0.5656 0.6065</td>
</tr>
<tr>
<td>Flowering duration</td>
<td>0.2907 0.2647 0.3113</td>
<td>0.3147 0.2410 0.3459</td>
<td>0.2928 0.2492 0.3311</td>
</tr>
</tbody>
</table>

CI 95 \% confidence intervals obtained by jack-knifing over seed families. All among all populations. Orig1200 among low elevation populations (1,200 m a.s.l.), Orig1800 among high elevation populations (1,800 a.s.l).
Evidence for divergent selection

Considering low and high elevation populations together, all traits revealed significant heritability (Table 3) indicating evolutionary potential (Silvertown and Charlesworth 2001). $Q_{ST}$ values exceeded $F_{ST}$ values in all measured traits (Fig. 3), which indicates divergent selection and implies that different phenotypes are favoured in different populations (Merilä and Crnokrak 2001). This result is in line with a meta-analysis including 50 plant and animal species, which revealed that 70% of the $Q_{ST}$ values exceeded their associated $F_{ST}$ values (Leinonen et al. 2008). The average $Q_{ST}$–$F_{ST}$ difference among all populations in our study of 0.19 (Fig. 3; Table S3) is about 60% higher than the average of 0.12 reported in the review of Leinonen et al. (2008) but substantially less than the difference of 0.58 in Rana temporaria (Palo et al. 2003) or 0.74 in Arabis fecunda (McKay et al. 2001). The authors of the latter study attributed the large estimate to geographic isolation and small effective population sizes of their rare endemic study species whereas R. bulbosus is a rather common species and distances among the sampled populations in the Swiss Alps are much shorter. These dissimilarities in abundance and sampling design might explain the smaller average $Q_{ST}$–$F_{ST}$ differences in our study. While overall $Q_{ST}$–$F_{ST}$ differences in half of the traits of R. bulbosus were larger than 0.21, they were very small in leaf length, bulb biomass and SLA. The latter two traits are expected to have smaller $Q_{ST}$–$F_{ST}$ values because their generally relatively complex genetic background constrains strong evolutionary changes (De Kort et al. 2013) or it might indicate that these traits evolve neutrally. Highest divergent selection was found in reproductive phenology (flowering start and fruiting start). Phenological traits often vary across environmental gradients, such as advanced phenology at low as compared to high elevations, and hence are expected to be under divergent selection (Linhart and Grant 1996). Moreover, these traits are often associated with few large-effect genes (De Kort et al. 2013).

In line with the findings of Hahn et al. (2012), mean $F_{ST}$ values were generally low in all three data sets and showed a weak but significant IBD pattern (Table 4C). Moreover, a study on pollen flow in R. bulbosus along an elevation gradient in the Swiss Alps suggests high levels of contemporary gene flow, at least across small scales of $<1$ km (Matter et al. 2013). These findings suggest that gene flow constrained neutral genetic differentiation between more closely located populations. Under high levels of gene flow selection pressures or genetic drift must be very strong to leave a genetic imprint (Slatkin 1987) resulting in high $Q_{ST}$ values (De Kort et al. 2013). In our study, average $Q_{ST}$ of 0.27 was within the range of other insect-pollinated herbs such as Ranunculus reptans, Silene diclinis or Scabiosa columbaria (reviewed in Leinonen et al. 2008) but far below those of the annuals.
Table 4 Mantel correlation coefficients (r) testing pairwise correlations of (A) $Q_{ST}^F$ vs. $F_{ST}^F$, (B) $Q_{ST}^F$/(1 - $Q_{ST}^F$) vs. log10(geographical distance) and (C) $F_{ST}^Q$ vs. $GeoDist$ in *Ranunculus bulbosus*

<table>
<thead>
<tr>
<th></th>
<th>All</th>
<th>Orig1200</th>
<th>Orig1800</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>r</td>
<td>$P_{Bonf}$</td>
<td>r</td>
</tr>
<tr>
<td>A) $Q_{ST}^F$ vs. $F_{ST}^F$</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Number of leaves</td>
<td>-0.07</td>
<td>&gt;0.99</td>
<td>0.00</td>
</tr>
<tr>
<td>Leaf length</td>
<td>0.10</td>
<td>&gt;0.99</td>
<td>0.29</td>
</tr>
<tr>
<td>Leaf biomass</td>
<td>0.17</td>
<td>&gt;0.99</td>
<td>0.24</td>
</tr>
<tr>
<td>Bulb biomass</td>
<td>0.17</td>
<td>&gt;0.99</td>
<td>0.34</td>
</tr>
<tr>
<td>SLA</td>
<td>-0.17</td>
<td>&gt;0.99</td>
<td>-0.18</td>
</tr>
<tr>
<td>Number of flowers</td>
<td>-0.01</td>
<td>&gt;0.99</td>
<td>-0.02</td>
</tr>
<tr>
<td>Flower stalk length</td>
<td>0.10</td>
<td>&gt;0.99</td>
<td>0.07</td>
</tr>
<tr>
<td>Flowering start</td>
<td>0.20</td>
<td>&gt;0.99</td>
<td>-0.20</td>
</tr>
<tr>
<td>Fruiting start</td>
<td>0.25</td>
<td>0.37</td>
<td>0.34</td>
</tr>
<tr>
<td>Flowering duration</td>
<td>0.09</td>
<td>&gt;0.99</td>
<td>-0.04</td>
</tr>
<tr>
<td>B) $Q_{ST}^F$ vs. $GeoDist$</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Number of leaves</td>
<td>0.01</td>
<td>&gt;0.99</td>
<td>0.06</td>
</tr>
<tr>
<td>Leaf length</td>
<td>0.06</td>
<td>&gt;0.99</td>
<td>0.08</td>
</tr>
<tr>
<td>Leaf biomass</td>
<td>-0.41</td>
<td>&gt;0.99</td>
<td>-0.11</td>
</tr>
<tr>
<td>Bulb biomass</td>
<td>0.14</td>
<td>0.21</td>
<td>0.13</td>
</tr>
<tr>
<td>SLA</td>
<td>0.00</td>
<td>&gt;0.99</td>
<td>0.06</td>
</tr>
<tr>
<td>Number of flowers</td>
<td>0.03</td>
<td>&gt;0.99</td>
<td>0.01</td>
</tr>
<tr>
<td>Flower stalk length</td>
<td>0.08</td>
<td>&gt;0.99</td>
<td>-0.01</td>
</tr>
<tr>
<td>Flowering start</td>
<td>0.11</td>
<td>&gt;0.99</td>
<td>-0.28</td>
</tr>
<tr>
<td>Fruiting start</td>
<td>0.04</td>
<td>&gt;0.99</td>
<td>0.10</td>
</tr>
<tr>
<td>Flowering duration</td>
<td>-0.41</td>
<td>&gt;0.99</td>
<td>0.02</td>
</tr>
<tr>
<td>C) $F_{ST}^Q$ vs. $GeoDist$</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Neutral markers</td>
<td>0.56</td>
<td><strong>0.001</strong></td>
<td>0.49</td>
</tr>
</tbody>
</table>

$P_{Bonf}$: Bonferroni corrected $P$-values for multiple comparisons. Significances are obtained by 1,000 permutations. Bold values indicate significant correlations ($P < 0.05$)

All among all populations, *Orig1200* among low elevation populations (1,200 m a.s.l.), *Orig1800* among high elevation populations (1,800 m a.s.l).

Arabidopsis thaliana ($Q_{ST} = 0.88$; Kuittinen et al. 1997) and Arabis fecunda ($Q_{ST} = 0.94$; McKay et al. 2001).

While neutral genetic markers followed an IBD pattern, quantitative genetic differentiation was not correlated with geographical distance in any of the investigated traits (Table 4B) indicating that geographical distance is not the driving selection agent in our study system. In contrast, quantitative genetic differentiation in amphibians increased with geographical distance among populations in Scandinavia (Palo et al. 2003; Hangartner et al. 2012). In these studies, geographical distance was also correlated with environmental clines (e.g. latitude or acidity) whereas in our paired sampling approach with evenly spread low and high elevation populations elevation was not correlated with geographical distance. Therefore, other factors which do not vary clinally with geographical distance seem to be responsible for the observed differentiation among populations, e.g. differences in land use (Stöcklin et al. 2009; Völler et al. 2013) or competition (Bischoff et al. 2006).

A positive correlation of $Q_{ST}$ and $F_{ST}$ would indicate that the degree of genetic differentiation in neutral markers reflects the degree of genetic differentiation in quantitative traits. Such correlations were found in a meta-analysis across different species (Leinonen et al. 2008). However, relatively few studies investigated the intraspecific correlation of pairwise $Q_{ST}^F$ with $F_{ST}^F$. Some authors found positive correlations between pairwise $Q_{ST}^F$ and $F_{ST}^F$ (Morgan et al. 2001; Steinger et al. 2002) whereas others reported correlations that were only weak and inconsistent among traits (Palo et al. 2003; Hangartner et al. 2012). Likewise our results showed no $Q_{ST}^F$-$F_{ST}^F$ correlation (Table 4A) indicating that selection, drift and gene flow did not influence molecular and quantitative differentiation in the same way. This limited predictability of quantitative traits from molecular markers might have important implications on conservation biological issues since management decisions are increasingly based purely on molecular analyses (e.g. Reed and Frankham 2001). Management plans should therefore also consider variation in quantitative traits, which might better reflect variation important for fitness (Lynch 1996).

Selection differences between low and high elevation populations

In addition to the divergent selection among all populations, we observed that the magnitude of selection differed between low and high elevation populations in eight out of ten traits (Fig. 3; Table S3). In number of flowers and flowering duration, divergent selection was stronger in high as compared to low elevation populations, which could be due to adaptation to the more heterogeneous local conditions at high elevations (Körner 2003). For SLA, high elevation populations were also under divergent selection whereas low elevation populations showed stabilizing selection (i.e. a negative $Q_{ST}^F$-$F_{ST}^F$ estimate). Furthermore, the stronger differentiation in leaf biomass, flower stalk length, flowering start and fruiting start in low as compared to high elevation populations might be shaped by greater heterogeneity in competition or land use among low elevation habitats (Choler et al. 2001; Rudmann-Maurer et al. 2008). Differing patterns of divergent selection in low and high elevation populations might influence plant responses to climate change. As a consequence of shifting temperature regimes,
current levels of quantitative differentiation at 1,200 m a.s.l. are expected to be observed at 1,800 m a.s.l. under predicted climate warming.

In contrast to the observed patterns of divergent selection within low and high elevation populations, comparisons of pairwise $Q_{ST} - F_{ST}$ values in low vs. high elevation populations indicated no divergent selection between low and high elevation populations (Table S4). This is in line with the lack of genetic differentiation between altitudes of origin in the majority of mean trait values (Table 2). Therefore, elevation is likely not the main selective agent driving the observed quantitative differentiation.

An alternative approach to test for divergent selection as the main driver of phenotypic differentiation is the reciprocal transplantation of plants across a species range (e.g. Joshi et al. 2001). Individuals performing better under home-site conditions provide evidence that natural selection resulted in local adaptation, which has been considered a widespread phenomenon across climatic gradients (Santamaria et al. 2003; Becker et al. 2006; Macel et al. 2007) and also along elevational gradients (Byars et al. 2007; Gonzalo-Turpin and Hazard 2009). In contrast, but in line with the findings of this study, a transplant experiment with R. bulbosus between low and high elevation sites revealed no evidence for local adaptation although several traits showed quantitative genetic differentiation possibly due to the co-occurrence of environmental factors, other than elevation (Frei et al. 2014).

Accuracy of $Q_{ST} - F_{ST}$ comparisons

Although $Q_{ST} - F_{ST}$ comparisons provide a valuable tool to separate neutral processes from natural selection, they are not without criticism. To mention a few points among others, first, the indication of divergent selection is conservative, because several non-additive components of variance, such as maternal and dominance effects, may lower the $Q_{ST}$ estimate relative to $F_{ST}$ (Lopez-Fanjul et al. 2003; Goudet and Buchi 2006). While full-sib approaches contain twice the environmental component of variance common to family members as well as part of the dominance and epistatic components of variance, the half-sib design used here reduces this bias (Lynch and Walsh 1998; Merilä and Crnokrak 2001; Lopez-Fanjul et al. 2003). Furthermore, biased estimates of within-population components of variance through maternal effects might be of minor importance in our study because these effects generally diminish with plant age (Roach and Wulff 1987) and our plants were about one year old at the start of the experiment. Second, environmental effects, that mask underlying genetic divergence, might lead to under- or overestimation of quantitative trait differentiation (Conover et al. 2009). In our common garden experiment, we can exclude such direct environmental effects typical in studies on individuals collected from the wild (Pujol et al. 2008). Third, the precision of $Q_{ST}$ estimates increases with the number of populations investigated (O’Hara and Merila 2005). Indeed, the confidence intervals of our $Q_{ST}$ estimates across all populations were often smaller than the ones calculated per elevation (Fig. 2; Table S2). Since $Q_{ST}$ estimates are rather high in most of the studied traits, we are confident that potential inaccuracies in the estimates of $Q_{ST}$ would not influence the results qualitatively.

Conclusions

By comparing quantitative trait to neutral molecular marker differentiation, we demonstrated that relatively strong divergent selection acted on several morphological and phenological traits among low and high elevation populations of the common semi-dry grassland species R. bulbosus from the Swiss Alps. Although pairwise $Q_{ST} - F_{ST}$ comparisons indicated no divergent selection between the two elevations, varying degrees of trait differentiation within each elevation indicated differences in selective forces between low and high elevation habitats. Differing selection pressures at contrasting elevations, and thereby under different temperature regimes, imply that climate change might alter selection processes. If so, the considerable levels of heritability might facilitate respective plant responses and thereby foster population persistence in the longer term.

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